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1 **Title: Genomic and morphometric evidence for Austronesian-mediated pig translocation in the Pacific**

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Abstract: Several millennia of human mediated translocation of non-native pig species (genus *Sus*) to the islands of Wallacea and Oceania has significantly altered local ecosystems. To investigate the timing and trajectory of these introductions, we conducted both genomic analyses of 576 pig nuclear genomes and a geometric morphometric analysis of 708 modern and ancient dental remains. Our analyses demonstrate that free-living and domestic pigs in Wallacea and Oceania possess diverse ancestries resulting from the introduction of multiple, sequential pig populations followed by gene flow. Despite the variability in their genomic ancestry, these pigs all possess a distinct tooth morphology, and a genetic link to the Chinese domestic pig populations that accompanied the dispersal of Austronesian language speakers ~4,000-3,000 years ago via Taiwan and the Philippines.

Introduction

People have transported animal species both involuntarily (e.g., commensal species such as rats and mice), and deliberately (e.g., as livestock or part of game management practices) beyond their native ranges for millennia. These introductions have often substantially altered local ecosystems, particularly on islands (1). The translocation of vertebrates eastwards across the Wallace Line (Fig. 1A) into Wallacea, a biogeographic region between the Asian and Australasian biotas, has had dramatic environmental impacts (2). For example, while the natural range of the genus *Sus* (pigs) is primarily located west of the Wallace Line, and limited to the westernmost part of Wallacea (i.e., Sulawesi and other islands between the Asian and Australasian continental regions), pigs are now widespread across archipelagoes in Wallacea, and Near and Remote Oceania (Fig. 1A). The geographic origin and ancestry of these pig populations across Wallacea and Oceania, however, remain unknown.

Previous genomic research has shown that pigs within the genus *Sus* can be broadly classified into three groups: Western Eurasian wild boar (*S. scrofa*, the ancestor of Western domestic pigs), Eastern Eurasian wild boar (*S. scrofa*, the ancestor of Eastern domestic pigs), and Island Southeast Asian (ISEA) endemic species found in Borneo and Sumatra (*S. barbatus*), Java (*S. verrucosus*), Sulawesi (*S. celebensis*), and the Philippines (*S. cebifrons*) (Fig. 1A; (3–6)). The human-mediated translocation of pigs east of the Wallace Line most likely began with *S. celebensis* in pre-Neolithic times >4,000 years ago, prior to the dispersal of Austronesian language speaking peoples and domestic animals into the region (7). The recent discovery on Sulawesi of the world’s oldest figurative artwork (>50,000 years) that depicts the local endemic warty pig species (*S. celebensis*) demonstrates the deep-rooted relationship between humans and pigs in the region (8, 9). Mitochondrial DNA and zooarchaeological analyses have identified *S. celebensis* remains from pre-Neolithic layers (purportedly older than 4,000 years) in Liang Bua Cave, Flores (10–12), though these have not yet been directly dated. If confirmed by radiocarbon dating, the presence of this species in pre-Neolithic deposits, ~300 km from its natural range in Sulawesi across the Flores Sea, would suggest these pigs were intentionally translocated by people.

Mitochondrial analyses indicated that the Eurasian pig (*S. scrofa*) also occurred in later deposits at Liang Bua Cave (younger than 4,000 years ago), suggesting *S. scrofa* was introduced to Flores subsequent to the introduction of *S. celebensis* (10–12). In fact, analyses of pig mitochondrial data from Wallacea and Oceania showed that introduction of *S. scrofa* was considerably more widespread than *S. celebensis*, since populations of *S. scrofa* are now free-living throughout these regions (Fig. 1; (10)). These *S. scrofa* populations were likely first introduced east of the Wallace Line 4,000-3,000 years ago during the expansion of Austronesian speakers from mainland East Asia and Taiwan, through the Philippines and Northern Wallacea, into Melanesia and Polynesia (13). This is supported by the remains of pigs (~2,900 years before present [BP]) excavated in Vanuatu from Lapita archaeological contexts associated with the Austronesian expansion (14–16).

117 Pigs from Oceania and Wallacea predominantly possess both a particular tooth shape and a distinct
118 mitochondrial haplogroup known as the “Pacific Clade” (10, 17). This mtDNA signature has also been
119 identified in free-living *S. scrofa* individuals from Java, Sumatra, and mainland Southeast Asia, including
120 Laos, Vietnam, and Yunnan Province in southwestern China. This distribution has been interpreted to mean
121 that pigs introduced to Wallacea likely originated from peninsular Southeast Asia (10, 17). In contrast,
122 modern domestic pigs found along the hypothesized route of the Austronesian expansion in Southeast China,
123 Taiwan, and the Philippines possess distinct Lanyu (known as the “Lanyu Clade”) and East Asian
124 mitochondrial signatures (10, 17). These data suggest that although Austronesian speakers may have
125 travelled with pigs that possessed the Lanyu Clade signature as they arrived in the Philippines, they only
126 acquired pigs with the Pacific Clade mitochondrial haplotype, as well as a variety of horticultural and
127 arboreal crops, including banana, sugarcane and yam (18), after leaving the Philippines.

128 West of the Wallace Line, however, the Pacific Clade mitochondrial haplogroup has only been identified in
129 free-living pigs (10, 17). The absence of this haplogroup in modern domestic pigs thus challenges the idea
130 that it represents a marker of domestic pigs introduced from mainland Asia. Furthermore, recent nuclear
131 genome analyses have shown that, although wild *S. scrofa* populations are primarily found in mainland
132 Eurasia, some *S. scrofa* populations naturally occur in Western Indonesia (Sumatra, and Java; Fig. 1A; (3,
133 4)). While these populations west of the Wallace Line mostly possess haplogroups typically associated with
134 ISEA-endemic *Sus* (due to gene flow; (3, 4)), some individuals possess the Pacific Clade haplogroup (10,
135 17). It thus remains unclear whether free-living pigs found east of the Wallace Line represent *S. scrofa*
136 populations that dispersed naturally (analogous to *S. celebensis*), or were translocated eastward by people.

137 The successful natural dispersal of the ancestor of another ISEA-endemic *Sus*, *S. celebensis*, from Borneo to
138 Sulawesi (4), and its subsequent human-mediated translocation to the Lesser Sunda Islands suggests that a
139 combination of natural and human-mediated dispersal of wild *S. scrofa* across the Wallace Line and onward
140 toward Oceania is plausible. In this scenario, the pigs discovered in Lapita archaeological contexts may have
141 been the descendants of domestic pigs introduced by Austronesian-speaking groups from mainland Asia, and
142 that they acquired mitochondrial haplotypes belonging to the Pacific Clade through gene flow with other pig
143 populations that were already present in Wallacea.

144
145 To establish the geographic origin of the pig populations in Wallacea, Melanesia, Micronesia and Polynesia,
146 we sequenced 117 *Sus* nuclear genomes and generated geometric morphometric data (from the third lower
147 molar) for 401 modern and 313 archaeological specimens. The genomic data generated here included 54
148 modern genomes, 60 historical genomes (1871-1993) and three ancient genomes from the Philippines (~500
149 years BP), Palau (Micronesia; ~1400 years BP), and the Lapita archaeological site of Teouma on Efate
150 Island, Vanuatu (~2,900 years BP). We analyzed these data alongside publicly available data that included
151 400 nuclear genomes (Fig. 1A) and ~700 mtDNA control region sequences.

152 153 **Results and Discussion**

154 ***The origin of the Pacific Clade mitochondrial haplogroup and associated morphometric signature***

155 We assessed the spatiotemporal frequency of both the Pacific Clade and Lanyu Clade across ISEA and the
156 Pacific, based on a data set consisting of 585 mtDNA whole genome (>10x coverage; Fig. S3), 703 control
157 region fragment sequences including historical and ancient specimens (10, 17), and recently published
158 datasets of 356 modern individuals from the Philippines (19, 20). We identified 58 individuals that possessed
159 mitochondrial sequences belonging to the Lanyu haplogroup, all of which were found in the Philippines
160 (n=52, of which n=7 from Palawan) and Taiwan (n=6). Of the 90 individuals that possessed a Pacific Clade
161 signature (Fig. 1B), the majority (n=55) were located east of the Wallace Line in Wallacea, New Guinea,
162 Melanesia, Polynesia, and in a single domestic pig from the Philippines (out of 356). The majority (~77%) of
163 pigs east of Wallace line possessed the Pacific Clade, while ~22% (16/72) possessed haplotypes belonging to

164 the East Asian haplogroup and one individual from New Caledonia possessed a haplotype belonging to the
165 European clade. The remaining individuals with a Pacific Clade haplogroup were found across South and
166 Peninsular Southeast Asia (Fig. 1B).

167
168 These results suggest that the distinctive tooth shape (Pacific shape) previously associated with pigs carrying
169 a Pacific Clade mtDNA signature may also be present west of the Wallace Line. To address this, we
170 conducted a morphometric analysis of pig teeth (third lower molar) based on 307 archaeological and 401
171 modern specimens. We used 12 landmarks and 87 sliding semilandmarks to capture tooth shape variation.
172 The Pacific shape was identified in pigs east of the Wallace Line including modern and ancient individuals
173 from Papua New Guinea (PNG), Vanuatu and the Marquesas Islands (Fig. 1C). This tooth shape is distinct
174 from the shape observed in archaeological pigs from mainland East Asia (correct cross-validation = 97%
175 (CI: 92.3-100%); Data S3).

176
177 None of the 36 archaeological individuals from the Philippines possessed the Pacific shape, while four
178 archaeological out of 35 individuals in total (both modern and archaeological) from Taiwan were identified
179 as having this shape with high probability (>92%; Data S3). Nine out of 14 individuals from Sarawak
180 (Borneo) were also found to possess the Pacific shape. These results demonstrate that multiple pig
181 populations found west of the Wallace line in Taiwan, the Philippines, Borneo, and mainland Asia carry the
182 distinctive Pacific mtDNA and tooth shape signatures. This geographic distribution, and the possibility of
183 morphological convergence, complicates the use of these markers for determining the origins of pigs found
184 east of the Wallace Line.

185 186 ***The ancestry of pigs east of the Wallace Line***

187 To further investigate the origin of pigs east of the Wallace Line, we analyzed 576 nuclear genomes
188 sequenced to >0.1x coverage (Data S1). We first conducted a Principal Component Analysis (PCA) that
189 corroborated the existence of three major ancestry clines (Fig. 1D) observed in previous publications (4, 21,
190 22). Principal Component 1 (PC1) differentiated the reciprocally monophyletic *S. scrofa* (wild and domestic)
191 and wild ISEA-endemic *Sus* species: *S. celebensis*, *S. cebifrons*, *S. philippensis*, *S. barbatus* and *S.*
192 *verrucosus* (Fig. 1D). PC2 differentiated European (wild and domestic) and Asian *S. scrofa* populations
193 (wild and domestic; Fig. 1D). *S. scrofa* pigs from ISEA (including Wallacea), Micronesia, Melanesia, and
194 Polynesia (represented by red triangles in Fig. 1A & D) are broadly distributed across PC1 and PC2,
195 indicating that they possess mixed ancestry that includes ISEA-endemic *Sus*, Asian *S. scrofa* and European
196 *S. scrofa*. These results are consistent with the observation that the majority of free-living *S. scrofa*
197 individuals west of the Wallace Line from Sumatra, Java, the Malay Peninsula and nearby islands, also
198 possessed ISEA-endemic *Sus* mtDNA haplogroups (Fig. 1B; Fig. S3).

199
200 To quantify the proportion of ancestry derived from ISEA-endemic *Sus*, Asian *S. scrofa* and European *S.*
201 *scrofa* proportions in individual genomes, we used a combination of analyses including ADMIXTURE and
202 D-statistics. We first employed an unsupervised ADMIXTURE analysis (K=3) based on 50 wild *S. scrofa*
203 individuals from Europe and 40 from East Asia (China and Korea), in addition to the 30 ISEA-endemic *Sus*
204 genomes in our dataset. This facilitated the identification of representative genomes for each ancestry (i.e.,
205 those that possessed >99% from only one source; Fig. S6). These representatives served as sources in a
206 supervised ADMIXTURE (K=3) analysis that included the 576 genomes with >0.1x coverage. We then
207 decomposed ISEA-endemic *Sus* ancestry into one of four components (*S. celebensis*, *S. cebifrons/S.*
208 *philippensis*, *S. barbatus*, and *S. verrucosus*) using D-statistics (see Supplementary Material).

209
210 These analyses confirmed the widespread of both ISEA-endemic *Sus* and Asian *S. scrofa* nuclear ancestry in
211 free-living *S. scrofa* populations east and west of the Wallace Line (4). Many of these samples also
212 possessed ISEA-endemic *Sus* mtDNA haplogroups, particularly in Wallacea, Sumatra, Java, and the Malay

213 Peninsula (Fig. 1B; Fig. 2). There was also a strong concordance between geographic location and ISEA-
214 endemic *Sus* ancestry: 18 *S. scrofa* individuals from Java and Sumatra possessed between ~13-37% ancestry
215 derived from either *S. verrucosus* (endemic to Java) or *S. barbatus* (endemic to Sumatra and Borneo), while
216 eight admixed individuals east of the Wallace Line possessed between ~15-80% *S. celebensis* ancestry
217 (endemic to Sulawesi, east of the Wallace Line; Fig. 2). We found little to no *S. cebifrons/S. philippensis*
218 ancestry (endemic to the Philippines) in 14 modern domestic pigs (*S. scrofa*) from the Philippines (Fig. 2).

219
220 Most free-living *S. scrofa* from Sumatra, Java, and nearby islands possessed similar levels of *S. verrucosus*
221 ancestry (13-18%). This finding supports previous studies indicating that wild *S. scrofa* from Sumatra
222 interbred with *S. verrucosus* during the Pleistocene when low sea levels allowed for the connection of
223 Sumatra, Borneo, and Java via the exposed Sunda Shelf, which facilitated migration and population
224 exchange (3, 4). Three wild pigs from Simeulue Island, northwest of Sumatra, which were suspected to have
225 been part of a population translocated from Sulawesi based on their morphological classification as *S.*
226 *celebensis* (23), possessed ~30% *S. barbatus* and ~70% Asian *S. scrofa* ancestry. These results suggest that
227 gene flow can lead to the generation of hybrid populations that can then confuse morphological analyses,
228 resulting in spurious species identifications.

229
230 Although all 12 individuals in Wallacea i.e. free-living pigs from Maluku, the Lesser Sundas and New
231 Guinea are outside of the natural range of *Sus* species they all possessed some degree of Asian *S. scrofa*
232 and/or *S. celebensis* ancestry, though the proportion of each varied widely between islands (Fig. 2). In the
233 Maluku archipelago northeast of Sulawesi, *S. celebensis* ancestry in free-living pigs ranged from 80% on
234 Halmahera to ~5% on Morotai, just a few kilometers north. Despite its high proportion of *S. celebensis*
235 ancestry, this 1979 individual from Halmahera was classified and accessioned as *S. scrofa* at the Smithsonian
236 Natural History Museum (24). Additionally, *S. celebensis* ancestry was absent in free-living pigs from the
237 Central Maluku islands of Buru and Seram, despite previous surveys suggesting the presence of *S. celebensis*
238 on Buru (25). This lack of *S. celebensis* ancestry on Buru is also puzzling considering the presence of
239 babirusa (*Babyrusa babyrussa*) on the island, which are thought have originated from Sulawesi, where *S.*
240 *celebensis* also occurs.

241
242 A similar pattern was observed in the Lesser Sunda region of Wallacea including the islands of Timor, where
243 *S. celebensis* ancestry in free-living pigs ranged from 100% to 29%, and Flores, where it ranged from 16% to
244 5%. These results demonstrate that *S. celebensis* populations occur well beyond its native range on Sulawesi.
245 While natural dispersal to some islands cannot be ruled out, the open water distance between Sulawesi and
246 Flores suggests that seafaring humans deliberately translocated this species during or since the Late
247 Pleistocene (10, 11). Despite the significant variation in *S. celebensis* ancestry (0-80%) in individuals found
248 east of the Wallace Line, the consistent presence of a *S. scrofa* Pacific Clade mtDNA haplogroup across all
249 individuals, indicates a component of Asian *S. scrofa* ancestry (Fig. 1B).

250
251 We did not detect any ISEA-endemic *Sus* ancestry in domestic pigs from the Philippines, nor in free-living
252 or domestic pigs from Melanesia or Polynesia. European *S. scrofa* ancestry was also absent from historical,
253 free-living individuals from New Guinea dated to the early-to-mid-20th century, but ranged from 10% to
254 58% in modern domestic New Guinea populations. In modern New Caledonian free-living pigs, European
255 ancestry ranged from ~43-57%, and a free-living pig from Hawaii, sampled in 1937, possessed 58%
256 European ancestry. All recent domestic pigs from the Philippines possessed between 27-90% European
257 ancestry. We also detected ~42% European ancestry in a free-living pig from Sumba, a Wallacean island
258 south of Flores.

259
260 Patterns of European and Asian *S. scrofa* ancestry covariance across the genome based on DATES (26)
261 indicated that European *S. scrofa* ancestry was introduced to these islands during the 19th and early 20th

262 centuries (Supplementary Material). In addition, ancient genomes from Vanuatu (~2,900 BP), the Philippines
263 (~500 BP), and Palau (~700 BP) showed no evidence of European ancestry. These results demonstrate that
264 European *S. scrofa* ancestry was introduced to these regions, likely via European domestic pigs during and
265 since the colonial period, and has since risen to substantial proportions in both free-living and domestic pig
266 populations across the region.

267 268 ***The geographic origin of Asian S. scrofa genomic ancestry east of the Wallace Line***

269 Although multiple historical/modern free-living and domestic pigs east of the Wallace Line possess a diverse
270 mix of ancestry, including *S. celebensis* and European *S. scrofa*, they all possess some degree of Asian *S.*
271 *scrofa* ancestry, and often possess a *S. scrofa*-like mtDNA haplogroup (Fig. 1B). Assessing the geographic
272 origin of this Asian *S. scrofa* ancestry is challenging, however, particularly in mixed genomes that also
273 feature European *S. scrofa* or ISEA-endemic *Sus* ancestry. In addition, the Asian *S. scrofa* lineage is
274 widespread and encompasses wild boar from North and South China, Korea, Japan, ISEA (Sumatra and
275 Java), as well as ancient, historical, and modern pigs from the Philippines, Near and Remote Oceania, and
276 East Asian domestic pigs.

277
278 To overcome these constraints, we performed ancestry deconvolution, a process by which specific ancestries
279 in admixed individuals can be identified and isolated using local ancestry inference. We first phased all
280 genomes with >8x coverage using SHAPEIT v4 (27), and imputed the genomes with >1x coverage using
281 GLIMPSE (28). We then used GNOMIX (29) to identify ancestry blocks from Asian *S. scrofa*, European *S.*
282 *scrofa*, and ISEA-endemic *Sus* ancestry, using wild, un-admixed individuals as reference populations (see
283 Supplementary Material). Downsampling showed that GNOMIX local ancestry inference is highly accurate
284 (94-99%) for genomes over 0.1x coverage (Fig. S5).

285
286 For each admixed genome, we filtered ancestry blocks assigned to European *S. scrofa* and ISEA-endemic
287 *Sus* ancestry. We then computed f4 ratios to compare levels of non-Asian *S. scrofa* ancestry before and after
288 filtering out both European *S. scrofa* and ISEA-endemic *Sus* ancestry blocks (Fig. 3A). European *S. scrofa*
289 ancestry in admixed individuals from New Caledonia, Hawaii, Papua New Guinea and Taiwan (Lanyu pigs)
290 decreased from ~25-50% to 0-5%, while European *S. scrofa* ancestry in domestic pigs from the Philippines
291 decreased from ~30-70% to 0-15%. ISEA-endemic *Sus* (*S. celebensis*) ancestry in individuals from
292 Wallacea, including Flores, Timor, and Halmahera, decreased from between ~40-80% to below 5%.

293
294 ISEA-endemic *Sus* (*S. verrucosus*) ancestry in wild pigs from west of the Wallace Line, in Java, Sumatra,
295 and nearby islands, however, decreased from between 5-14% to ~5%. This slightly lower deconvolution
296 success in these individuals can be explained by ancient admixture that took place during the Pleistocene (3,
297 4). Because this gene flow took place earlier in time, the ISEA-endemic *Sus* ancestry across the genome now
298 exists in blocks that are too small to identify. As a result of its more recent introduction to the region, the
299 European *S. scrofa* ancestry is present in larger blocks (Fig. 3B), and is thus more easily identified. The wild
300 pigs from west of the Wallace Line likely possess *S. scrofa* ancestry that is not well represented in our
301 panels, which could make it harder to identify by GNOMIX (see Supplementary Material).

302
303 Principal component analysis using deconvoluted genomes indicated that populations east of the Wallace
304 Line possessed a closer genetic affinity with mainland domestic populations—such as breeds from Southeast
305 China (such as Luchuan, Wuzhishan, and Bamaxiang)—as well as with Lanyu pigs from Taiwan and
306 domestic pigs from the Philippines, which aligns with the hypothesized route of the dispersal of
307 Austronesian language speakers (13).

308
309 We then assessed the genetic similarities between the Asian-derived ancestry of *S. scrofa* individuals east of
310 the Wallace Line and a 0.7x coverage genome from a ~2,900-year-old pig specimen (POU_Vanuatu_3 in

311 Fig. 1A) excavated at a Lapita archaeological site (associated with Austronesian speakers) in Vanuatu (14).
312 The relatively low coverage of this ancient individual (0.7x) makes direct comparison with deconvoluted
313 genomes challenging, since the deconvolution process itself (that removes European *S. scrofa* or ISEA-
314 endemic *Sus* ancestry) further reduces the effective coverage.

315
316 To circumvent this issue, we first computed outgroup-f3 statistics comparing POU_Vanuatu_3 with modern
317 samples that only possessed Asian *S. scrofa* ancestry. The objective of this step was to identify a proxy
318 individual with >1x coverage suitable for subsequent comparisons against all samples, including the
319 deconvoluted individuals. We found POU_Vanuatu_3 shared most drift with other Wallacean individuals
320 and domestic pigs from Southeast China (such as Luchuan, Wuzhishan, Bamaxiang), and was closest to
321 ISW_PNG_18, a ~100-year-old free-living pig from PNG (Fig. S11). Morphometric analyses further
322 demonstrated that both ancient Vanuatu pigs from similar contexts and ISW_PNG_18 possessed the
323 distinctive Pacific Shape tooth morphology (Fig. 1C; Data S1).

324
325 We therefore chose ISW_PNG_18 as a high-coverage proxy for the ancient Vanuatu specimen's ancestry.
326 Outgroup-f3 analyses performed on the deconvoluted genome dataset, which included all Asian (mainland
327 and ISEA) and Oceanian pigs, demonstrated that all *S. scrofa* individuals from Wallacea, Micronesia,
328 Melanesia, and Polynesia share significantly stronger genetic affinities with the ISW_PNG_18 proxy
329 compared to ISEA *S. scrofa* from west of the Wallace Line (Fig. 4). ISW_PNG_18 also shared more drift
330 with ISU_UchuPalau_1, a 672-year-old Micronesian pig from Palau and ISU_Philippines_1, a 500-year-old
331 individual from the Batanes Islands compared to individuals west and north of the Wallace line. This proxy
332 also shared more drift with domestic pigs from Taiwan (Lanyu pigs), and South China (e.g., Bamaxiang,
333 Luchuan and Wuzhishan pigs from Hainan Island and adjacent mainland Chinese provinces; underlined in
334 Fig. 4) than with domestic pigs from Northern China and Vietnam, or ISEA populations west of the Wallace
335 Line and wild Chinese *S. scrofa* (Fig. 4).

336
337 These results indicate that the domestic and free-living pigs found today beyond the natural range of *S.*
338 *scrofa* in Wallacea and the Pacific, are closer genetically to ancient pigs from Vanuatu (from early Lapita
339 archaeological contexts), and the Philippines, and modern domestic pigs in Taiwan, than to modern and
340 historical individuals from peninsular Southeast Asia and western Indonesia. This indicates that pigs in the
341 Pacific are, at least partly, descendants of domestic pigs introduced to the region via the Philippines, most
342 probably by Austronesian-speaking groups who possessed East Asian human ancestry (30).

343
344 This demonstration of shared ancestry undermines the previously mitochondrially-based suggestion (10, 17)
345 that Polynesian pigs were not part of the Austronesian expansion since they were thought to have originated
346 in Peninsular East Asia and dispersed through ISEA. The spatiotemporal distributions of the Pacific and
347 Lanyu mitochondrial clades still show a substantial turnover in the mitochondrial haplotypes associated with
348 Austronesian and Polynesian pigs. It is possible that both haplotypes were present (but as yet undetected)
349 during the early phase of the Austronesian expansion, but due to recurrent bottlenecks associated with their
350 dispersal, the Lanyu haplotypes vanished while the Pacific Clade haplotype became fixed.

351
352 Conditional heterozygosity analysis reveals that free-living and domestic pig populations east of the Wallace
353 Line, and in Taiwan (i.e. Lanyu pigs) exhibit significantly lower heterozygosity compared to their mainland
354 domestic ancestors (Fig. S14; $p < 0.001$; Supplementary Material), suggesting that they likely experienced a
355 bottleneck associated with founder events during their dispersal. Sequential bottlenecking could also explain
356 the fixation of the rarer Pacific Clade haplotype. Alternatively, it is possible that the Pacific Clade haplotype
357 was initially only present in native *Sus scrofa* populations in western ISEA. It then dispersed eastwards in
358 populations that then hybridised with the Austronesian East Asian pigs whereupon the Pacific Clade

359 haplotype replaced the Lanyu signature. Additional ancient nuclear and mitochondrial DNA from ancient
360 pigs across the region and from East Asia is necessary to determine which of these explanations is correct.

361
362 ***Multiple waves of dispersal and introduction in the Lesser Sunda Islands***

363 It is noteworthy that a 1929 free-living pig from Sumbawa (ISW_Sumbawa_1), a few hundred kilometres
364 east of the Wallace Line, possesses a mixture of Asian *S. scrofa* and *S. verrucosus* ancestry (endemic to
365 Java), similar to pigs in Bali. Pigs from Flores and Timor (Fig. 2), however, possessed a mixture of Asian *S.*
366 *scrofa* and *S. celebensis* ancestry (endemic to Sulawesi). Admixture modeling further suggests that at least
367 part of the *S. scrofa* component in the Sumbawa pig is derived from a sister-lineage to Asian and European
368 *S. scrofa*, similar to that observed in ISEA populations west of the Wallace Line (Fig. S9). These results
369 suggest that Sumbawa pigs may possess ancestry from the western Lesser Sunda Islands, suggesting a
370 possible natural and/or human-mediated translocation of pigs across the Wallace Line into the Lesser Sunda
371 Islands. This ancestry pattern in the Lesser Sunda Islands illustrates the complex evolutionary history of pigs
372 east of the Wallace Line.

373
374 Combined, our results suggest multiple dispersal processes in the region that possibly started with a natural
375 and/or human-mediated translocation of ISEA pig populations that possessed *S. scrofa* and *S. verrucosus*
376 ancestry from Bali moving east across the Wallace Line. This was likely followed by the introduction of *S.*
377 *celebensis* from Sulawesi to Lesser Sunda Islands such as Flores and Timor, potentially by hunter-gatherer
378 populations. Subsequently, domestic *S. scrofa* pigs derived from populations in mainland Asia were
379 introduced into the region from northern Wallacea (following their introduction from southern China and the
380 Philippines) sometime after 3,500-3,000 years ago. Many of these introduced pigs later became feral on
381 islands across the region. Finally, pigs with European ancestry were introduced broadly across the region,
382 including into the Lesser Sundas to the island of Sumba during the colonial period.

383
384 **Conclusions**

385 We found that most free-living and domestic pigs across a vast geographical region from the Philippines to
386 Hawaii possess a degree of ancestry linked to domestic pigs (*S. scrofa*) likely introduced during the spread of
387 Austronesian-speaking groups from Southeast China and Taiwan via the Philippines ~4,000 years ago.
388 Interestingly, pigs in Near and Remote Oceania lack any detectable ancestry from wild endemic *Sus* species
389 present along the Austronesian dispersal route in the Philippines, Sulawesi and other islands. This pattern
390 suggests that the first wave of introduced domestic pigs did not interbreed with local, indigenous, *Sus*
391 populations. Only their descendants left behind in Wallacea, and which subsequently became feral, had the
392 opportunity to hybridize with indigenous wild species such as *S. celebensis* (Fig. 1A; Fig. 2).

393
394 The lack of gene flow between local *Sus* species and the first domestic pigs introduced to Wallacea is
395 consistent with the pattern observed in the human populations who likely transported these pigs, and who did
396 not mix with local groups during the initial phases of their dispersal to Melanesia (30). The continuing
397 movement of Austronesian speakers eastwards across the islands of Oceania then resulted in the first
398 establishment of domestic and feral pig (*S. scrofa*) populations across the region, where no pig species were
399 previously present.

400
401 Their peculiar tooth morphology, and initial genetic isolation from local wild species they encountered in
402 ISEA and Wallacea, may reflect the fact that these introduced *S. scrofa* pigs possessed specific domestic
403 traits that facilitated their transport and management by Austronesian-speaking groups. Transporting these
404 animals between islands resulted in a unique evolutionary history characterized by serial founder effects,
405 gene flow from divergent lineages, and likely selection for specific traits that facilitated the establishment of
406 feral populations. Future research utilizing high-coverage genomes from modern, historical, and
407 archaeological samples, combined with functional assays and detailed morphometric analyses, will be crucial

408 to elucidate how these domestic pigs, introduced from mainland Asia, became so successful in establishing
409 feral populations across ISEA and the Pacific.

410

411 The Materials and Methods are available in the Supplementary Materials.

412

413 **References and Notes**

414

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648 **Competing interests:** Authors declare that they have no competing interests.

649

650 **Data and materials availability:**

651 All genomics data have been deposited at the European Nucleotide Archive (ENA) with project number
652 PRJEB83975. All morphometric data is available as in Data S3.

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655 **Supplementary Materials**

656 Materials and Methods

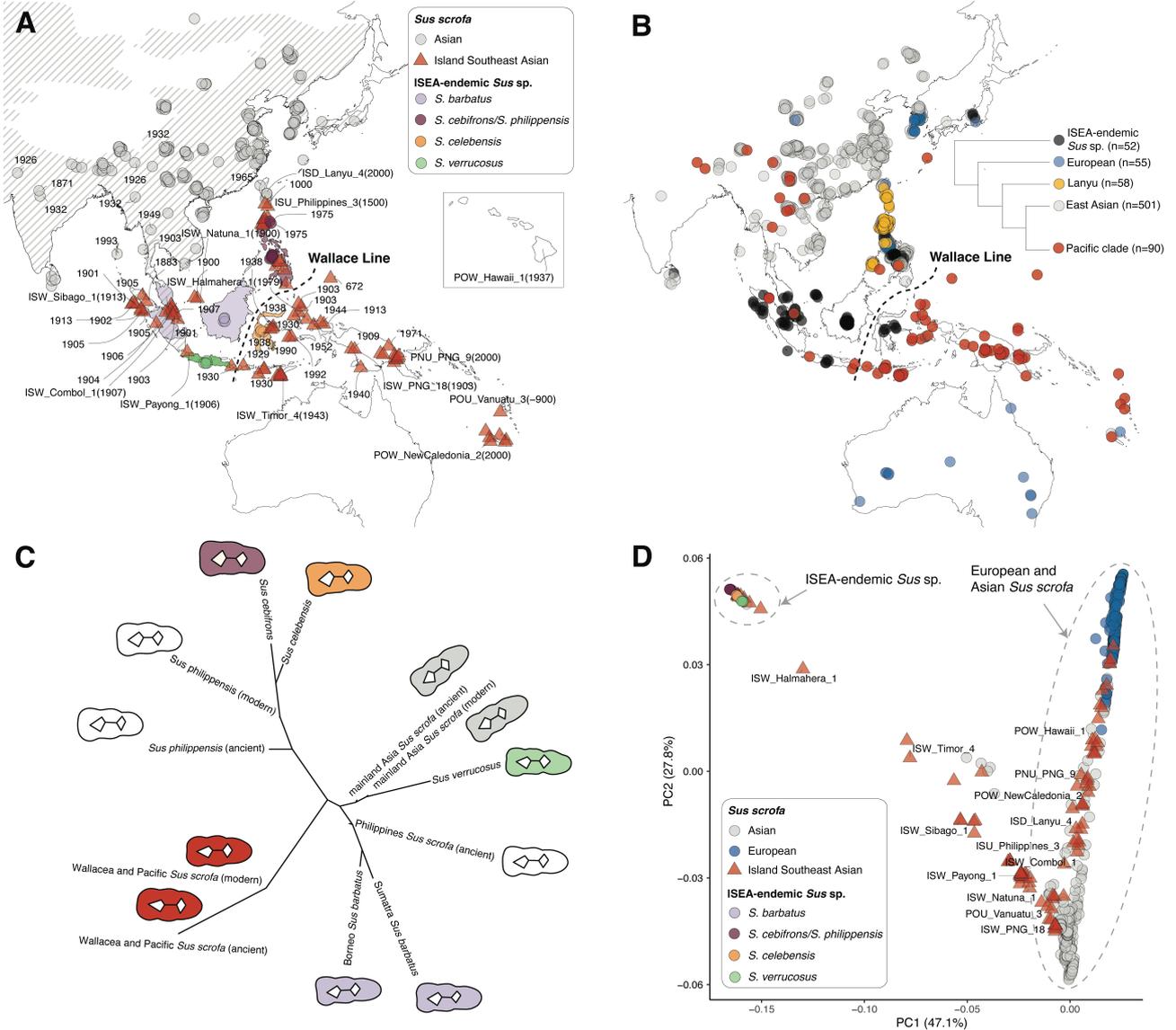
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658 Figs. S1 to S15

659 Data S1 and S3

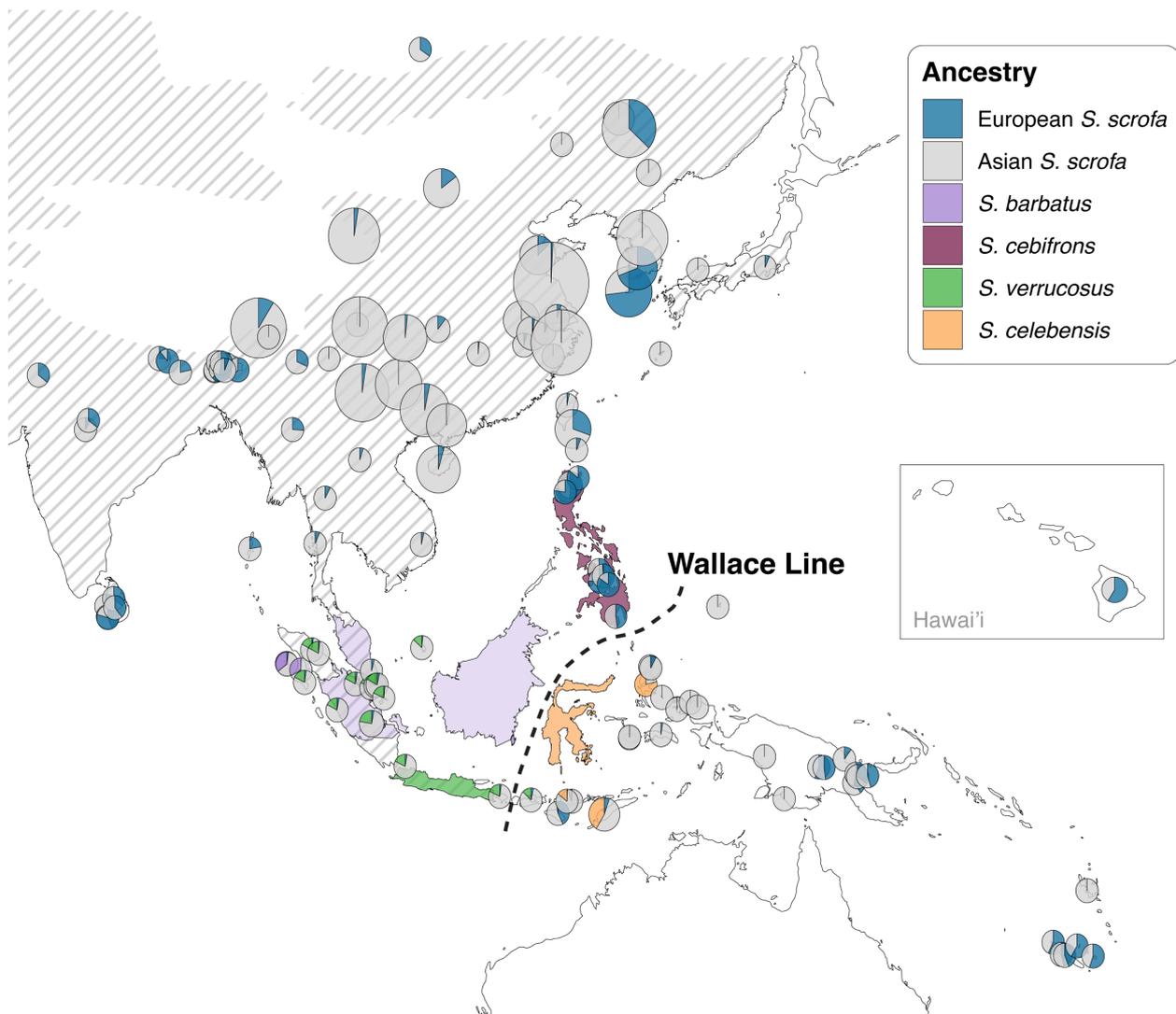
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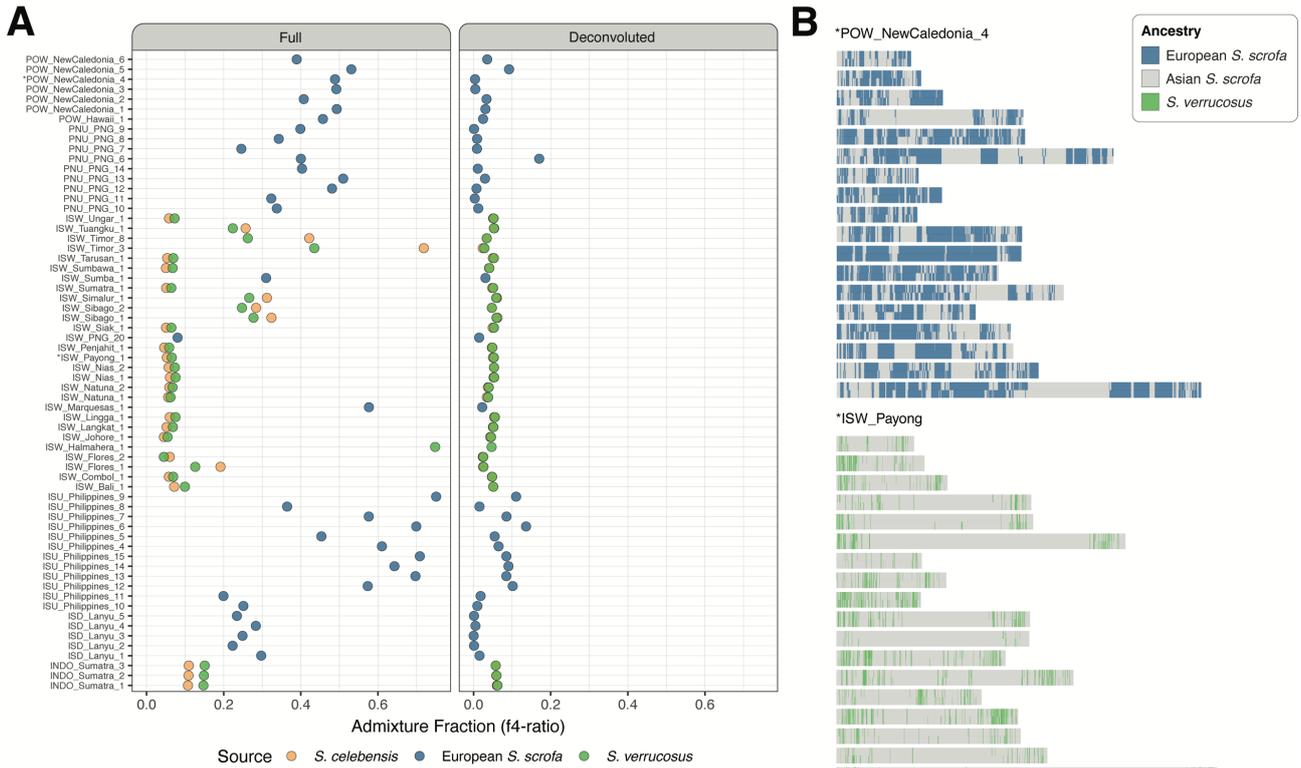


663
 664 **Fig. 1 A. Map of sampling locations.** The gray hatched area on the map represents the natural distribution
 665 of *Sus scrofa* (the ancestor of domestic pigs) in mainland Asia and Island Southeast Asia (ISEA)
 666 (specifically Sumatra). The light purple area on the map (Borneo and Sumatra) indicates the natural
 667 distribution of *S. barbatus*, the dark purple area shows *S. cebifrons* in the Philippines, the orange area
 668 represents *S. celebensis* on Sulawesi, and the green area is for *Sus verrucosus* on Java. The dashed line
 669 between Sulawesi and Borneo represents the Wallace Line. Red triangles indicate 49 newly sequenced
 670 historical *S. scrofa* specimens (dating from 1871-1994) from ISEA (cataloged as *S. scrofa* in museum
 671 collections) and three archeological specimens from the Philippines (~1500 AD), Palau (Micronesia; ~672
 672 AD), and the Lapita culture site of Teouma on Efate Island, Vanuatu. Gray dots represent individual *Sus*
 673 *scrofa* genomes (some of which are newly sequenced; see Data S1) from mainland Asia (both South Asia
 674 and East Asia). The locations of the ISEA endemic *Sus* species individual genome analyzed in this study are
 675 noted in purple (*S. barbatus*; Borneo), cream (*S. cebifrons*), orange (*S. celebensis*), and green (*S.*
 676 *verrucosus*). **B. Mitochondrial analysis.** Geographic distribution of major mtDNA haplogroups based on
 677 data from ~705 control regions and ~323 whole mitochondrial genome data. The tree on the left is a
 678 schematic representation illustrating the phylogenetic relationships between these major haplogroups. The
 679 complete phylogenetic tree, based on whole mitochondrial genome data, is available in Fig. S3. This figure
 680 illustrates the distribution of the red dots, representing the "Pacific Clade," extending west of the Wallace
 681 Line into Western Indonesia, the Philippines, and mainland Asia (as shown in the full mitogenome tree). **C.**

682 **Morphometric analysis.** This neighbour-joining network illustrates the morphometric similarities in lower
683 third molar (m3) shape among *Sus scrofa* and ISEA endemic *Sus* species, based on Mahalanobis distances.
684 The network was generated following a discriminant analysis that utilized the first 15 principal components
685 from a preceding Principal Component Analysis (PCA). These selected components explain 88.7% of the
686 total shape variance and were chosen to maximize the morphological distinctions between the groups. The
687 tooth outlines shown represent the mean shape for each population, derived from separate superimposition
688 analyses. **D. Principal component analysis (PCA).** This PCA, based on pseudohaploid genotype from >500
689 individuals, illustrates the genetic variation among Eurasian *S. scrofa* and ISEA-endemic *Sus* species. Red
690 triangles represent newly sequenced *S. scrofa* genomes from ISEA (museum specimens). Gray dots indicate
691 individual *S. scrofa* genomes from mainland Asia (both South Asia and East Asia; Fig 1A), blue dots
692 represent European *S. scrofa*, while purple (*S. barbatus*; Borneo), cream (*S. cebifrons*), orange (*S.*
693 *celebensis*), and green (*S. verrucosus*) represent ISEA-endemic species. The first principal component (PC1)
694 clearly separates the ISEA-endemic *Sus* species from *S. scrofa* individuals, while the second principal
695 component (PC2) distinguishes between European and Asian *S. scrofa*. The red triangles (*S. scrofa* genomes
696 from ISEA) are scattered across the PC1-2 space, reflecting their complex ancestry, which includes
697 contributions from Asian and European *S. scrofa* as well as ISEA-endemic *Sus* ancestry.
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 700 **Fig. 2. Ancestry of pigs inferred using ADMIXTURE and D-statistics.** The ancestry of each individual
 701 Asian pig in our dataset was decomposed into five components using a combination of supervised
 702 ADMIXTURE analysis and D-statistics. Initially, the ancestry was decomposed with K=3 in ADMIXTURE
 703 (see Fig. S6), representing European *S. scrofa*, Asian *S. scrofa*, and ISEA-endemic *Sus* ancestry. The ISEA-
 704 endemic *Sus* ancestry component was further resolved using D-statistics (see Supplementary Material). Each
 705 pie chart on the figure represents a sampling location, and its size is proportional to the number of samples
 706 from that location.



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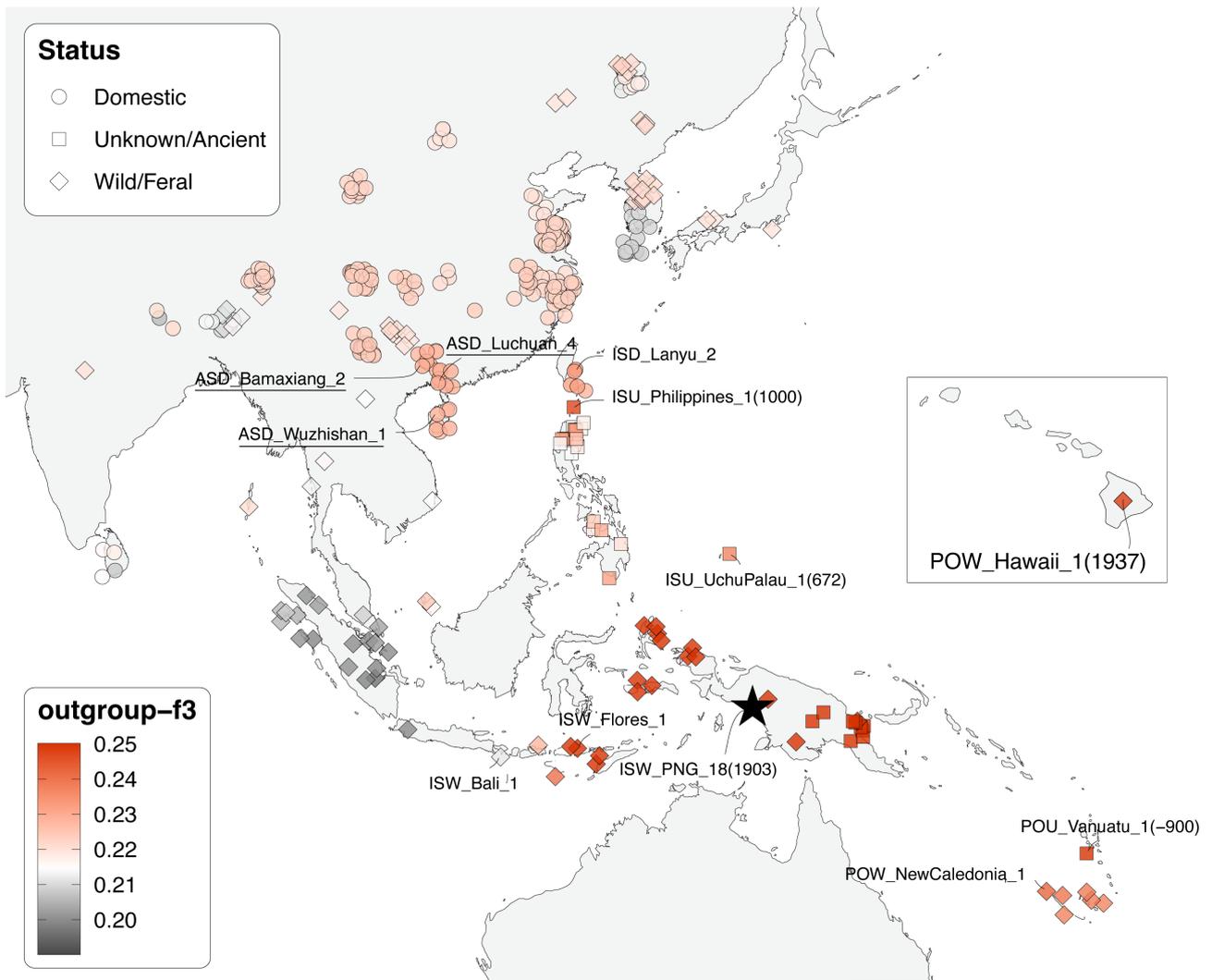
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Fig. 3A. Accuracy of ancestry deconvolution. European *S. scrofa* / ISEA-endemic *Sus* ancestry proportion in genomes of mixed ISEA individuals before (full) and after (deconvoluted) ancestry, with deconvolution calculated using the f_4 ratio (see Supplementary Material). Values close to 0 in the right panel (deconvoluted) indicate successful deconvolution, meaning the European *S. scrofa* / ISEA-endemic *Sus* ancestry was effectively removed from the genome. **B.** Blocks of European *S. scrofa* and *S. verrucosus* ancestry across chromosomes from a New Caledonian wild-caught individual and a wild pig from Payong, a small island a few km north of Java (Fig. 1A). This figure illustrates that the European *S. scrofa* ancestry blocks in the New Caledonian pig genome are considerably larger than the *S. verrucosus* ancestry blocks in the Payong pig genome. This difference indicates a more recent admixture event in the New Caledonian pig (dated to ~1922 using DATES (26); see Supplementary Material).



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Fig. 4. Outgroup-f3 after deconvolution. Outgroup-f3 (shared drift) of the form (Pygmy hog [outgroup], X, ISW_PNG_18) plotted on a map, where X represents any Asian individual in our dataset. Prior to this analysis, European *S. scrofa* and ISEA endemic *Sus* ancestry was removed from the genomes of admixed individuals using ancestry deconvolution. ISW_PNG_18 was selected as a proxy for the approximately 2,900-year-old Vanuatu pig specimen excavated from a Lapita culture archaeological site (see Fig. S11). Higher outgroup-f3 values indicate a greater degree of genetic affinity with the ISW_PNG_18 individual, which serves as a representative of pig ancestry associated with the dispersal of Austronesian language speakers. This figure demonstrates a strong genetic affinity between pigs east of the Wallace Line and those in mainland Asia (i.e. those underlined) to those introduced by Austronesian language speakers, rather than with pigs from west of the Wallace Line in ISEA.

Supplementary Materials for

Genomic and morphometric evidence for Austronesian-mediated pig translocation in the Pacific

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The PDF file includes:

Materials and Methods
Supplementary Text
Figs. S1 to S16

Other Supplementary Materials for this manuscript include the following:

Data S1 to S3

Materials and Methods

Archeological specimens and site description of DNA samples

Uripiv, Vanuatu (2900-2000 BP; POU_Vanuatu_3; contact: Stuart Bedford, School of Archaeology and Anthropology, College of Arts and Social Sciences, Australian National University, Canberra, ACT, Australia)

The small islands of Vao and Uripiv are located off the northeast coast of Malekula Island in northern Vanuatu. Excavations on uplifted beach terraces on the sheltered coast of the islands revealed dense concentrations of midden deposits associated with the colonising Lapita phase (c. 2900 BP) through to c. 2000 BP. The sites are rich in faunal remains, both terrestrial and marine, shellfish, pottery and other artefacts. The sites on both islands were deeply stratified, up to 1.8 m deep to the sterile beach sand in some places, and generally very well preserved showing limited disturbance.

Teouma, Vanuatu (3000-2600 BP; POU_Vanuatu_4, POU_Vanuatu_5; contact: Stuart Bedford, School of Archaeology and Anthropology, College of Arts and Social Sciences, Australian National University, Canberra, ACT, Australia)

The Teouma Lapita site is located on the south coast of Efate Island, Vanuatu, and is currently 800 metres from Teouma Bay. Tectonic uplift, volcanic ashfall prior to and during the period of human utilisation of the site, and alluvial deposition from an adjacent stream have altered its immediate coastal location during the Lapita period at c. 3000 BP. Excavations by a joint ANU-Vanuatu National Museum team took place between 2004 and 2010, during which 68 burial features, including remains of just over 100 individuals were found. Teouma has numerous indicators of being an initial colonisation site for Efate, including extinct faunal remains, early ceramic forms and decoration, and exotic New Britain obsidian. This combined with the extensive and previously very rare Lapita skeletal remains underline its significance to those investigating colonisation in this region. There is evidence of continuing occupation at the site to c. 2600 BP. During the Lapita phase the site was composed of a cemetery and a contemporary settlement with a midden deposit. During later phases settlement expanded across the cemetery area.

Wanigela, Papua New Guinea (600-700 BP; PNU_PNG_2, PNU_PNG_3; contact: Geoffrey Clark, Archaeology and Natural History, School of Culture, History and Language, Australian National University, Canberra, Australia)

The 14C sample was an adult pig tooth and fragment of mandible from Oreresan Village, Mound B (Spit 8 mixed) dated by Wk-30430, CRA 675 +/- 30, 13C -20.1, C:N 3.3. All the aDNA samples are from the Mound B excavation which is located in Oreresan Village located in coastal Eastern New Guinea in Collingwood Bay. The mound was excavated by Brian Egloff in the 1960s and site details are in his thesis publication (31).

Mound B is an oval-shaped mound near the Sasap River that is around 40 m in length and 17 m in width with a maximum height of 1.4 m. Excavations at the site totalled 22 m² with sediments divided into eight zones and subzones – Zone I (top), Zone IIA, IIB, IIC, Zone III, Zone IVA, IVB (base). Pig bones were common in all excavation levels and comprised 80% of the identifiable vertebrate remains (31). Many pigs had been killed before their third molars had erupted when the body weight was estimated to be less than 80-120 kg. Four radiocarbon results obtained by Egloff and suggested that the Mound B materials were deposited between AD 100 and AD 1450. The large amount of pig remains in Mound B might represent debris from ceremonial feasting indicating that the importance of pigs in traditional societies in New Guinea dates to AD 1300-1400 (Calib 7.10 result for Wk-30430 at 95.4% is AD 1270-1390).

Uchul A Rois, Palau (900-1450 AD; ISU_UchuPalau_1, ISU_UchuPalau_2; contact: Geoffrey Clark, Archaeology and Natural History, School of Culture, History and Language, Australian National University, Canberra, Australia)

Excavations of ~4 m² in Uchularois Cave by Masse (1989) (32) recovered an assemblage of pig bones totalling 110 identified specimens. Radiocarbon dates from Uchularois Cave have been recently revised and suggest that midden deposition began at AD 900, with site abandonment by AD 1450 (33). ISU_UchuPalau_1 was radiocarbon dated to ~1,300 calAD (OxA-44064).

Pamayan, Philippines (1420–1630 AD; ISU_Philippines_3; contact: Peter Bellwood, School of Archaeology and Anthropology, College of Arts and Social Sciences, Australian National University, Canberra, ACT, Australia)

Pamayan, Savidug village, Savidug Island, Batanes Islands, northern Philippines (Late prehistoric, overlapping with early Spanish period). The Pamayan shell midden lies at the back of Savidug village, exposed by a road cutting along the base of a limestone hill. A surface presence of red-slipped pottery indicated the existence of a potentially interesting site, so two small excavation sections of 60 by 80 cm were excavated 16 m apart back into the section exposed in the road cutting, exposing shell midden to a maximum depth of 105 cm beneath the topsoil (34). However, the recency of the Pamayan midden was rendered obvious by the occurrence of a few imported Chinese ceramic sherds down to 95 cm depth. A C14 date of 418±41 uncal. BP (WK 13091) from the base of the cultural layer indicates that the midden overlaps in its age of formation with the arrival of Spanish missionaries on the island, although there is a strong possibility that the archaeological materials have migrated downslope from a former site on top of the hill, a circumstance which makes the precise date of the pig bone sample uncertain. The pig bone was therefore dated using radiocarbon dating to 1475 calAD (OxA-43990).

Savidug Dune Site, Philippines (600-1000 BP; ISU_Philippines_1; contact: Peter Bellwood, School of Archaeology and Anthropology, College of Arts and Social Sciences, Australian National University, Canberra, ACT, Australia)

Savidug Dune Site, Savidug village, Savidug Island, Batanes Islands, northern Philippines. The Savidug Dune Site has two stratigraphic and cultural phases of usage (34). An upper cultural layer dated c.1000 to 600 BP, with imported ceramics and iron, hence overlapping in date with the formation of the Pamayan shell midden. A lower cultural layer dated c.3200 to 2000/1500 BP, which can be divided into an earlier and a later phase. The earlier phase is associated with rare examples of circle-stamped pottery, dated 3200 to 2700 BP in this site and from parallels at Sunget on Batan Island and Anaro on Itbayat Island. The later phase, dated c.2700 to 2000/1500 BP, is associated with jar burial and Taiwan nephrite.

Radiocarbon dating

Two samples were radiocarbon dated at the Oxford Radiocarbon Accelerator Unit (ORAU) The dates were calibrated on OxCal using the IntCal20 curve (35):

| Sample ID | Site, country | Uncalibrated radiocarbon age BP | Error (+/-) | Calibrated age BP | Calibration curve | Lab code |
|-------------------|----------------------|---------------------------------|-------------|-------------------|-------------------|-----------|
| ISU_UchuPalau_1 | Uchul a Rois, Palau | 1370 | 18 | 1308-1279 | IntCal20 | OxA-44064 |
| ISU_Philippines_3 | Pamayan, Philippines | 424 | 16 | 511-475 | IntCal20 | OxA-43990 |

Museum specimens

Museum samples for DNA were collected from the AMNH (American Museum of Natural History, New York, USA), the Australian Museum (Sydney, Australia), FMNH (Field Museum of Natural History, Chicago, USA), the Long An Museum (Vietnam), the NHM (Natural History Museum, London, UK), the NMS (National Museum of Scotland, Edinburgh, Scotland), the Naturalis (Naturalis Leiden, The Netherlands), the OUM (Oxford University Museum, Oxford, UK), and the USNM (Smithsonian National Museum of Natural History, Washington DC, USA). More information, including their museum identification for these samples can be obtained from Data S1.

Archeological and Museum specimen extraction and library

DNA extraction

Samples were extracted in two different ancient DNA facilities - Durham University and the University of Oxford - using conventional set-up for ancient and degraded genomic material (strictly isolated pre- and post-PCR rooms, use of full PPE with overshoes and double set of gloves [nitrile and latex], systematic extraction and library blanks was added to each batch of 12 samples. All equipment and work surfaces were cleaned before and after each use with a dilute solution of bleach (5-10%) followed by water and ethanol (99%). Pipettes and plastic racks were UV-irradiated in a crosslinker (254 nm wavelength) prior to and after use and/or decontaminated using “DNA AWAY” (Fisher Scientific SAS) or bleach followed by purified water. In both laboratories the ancient pig remains were prepared for DNA extraction by removing approximately two a millimetres layer of the outer bone surface by abrasion using a Dremel drill with clean cut-off wheels (Dremel no 409), targeting compact cortical bone or dentine. The bone was then pulverised in a Micro-dismembrator (Sartorius-Stedim Biotech), followed by collection in 15 mL Grainer tubes (Durham) or in 2 mL ependorf tubes (Oxford). Milling containers and grinding balls were subsequently suspended and cleaned in 1% virkon (Durham) or “DNA AWAY” (Oxford) and rinsed in absolute ethanol.

In Durham, the samples were extracted using the protocol described in (36). Bone powder (100-400 mg) was digested in 0.425 M EDTA, 0.05% SDS, 0.05 M Tris-HCl and 0.333 mg/mL proteinase K and incubated overnight (18-24 hours) on a rotator at 50 °C, or until fully dissolved. The digestion buffer, excluding proteinase K, was UV-irradiated (254 nm wavelength) for an hour in a dedicated cross-linker prior to use. 2mL of extract solution was then concentrated in a Millipore Amicon Ultra-4 30 KDa MWCO (Millipore) to a final volume of 100µL. The concentrated extract was purified using silica spin-columns (QIAquick PCR Purification Kit, Qiagen) following manufacturers recommendations, except that the final elution step was performed twice to produce a final volume of 100µL. One in five or one in ten negative extraction controls were performed alongside the ancient bone samples.

In Oxford, we used 80-120 mg of bone or dentine powder that was first pre-digested in 1 mL a buffer made of EDTA and proteinase K for 1h on a rotator at 37 °C, to remove extracellular DNA and some contaminants ((37). After centrifugation, this first lysis was removed and stored in the freezer. The pellets were then fully digested in 1.8 mL of the same digestion buffer, incubated overnight on a rotator at 50 °C. The DNA was extracted from the resulting lysis using a modified (38) protocol. In brief, the 1.8 mL of solution were mixed to 50 ml of the extraction buffer 0.5 M EDTA, pH 8.0 to a final concentration of 0.45 M, 10 mg/ml Proteinase K to a final concentration of 0.25 mg/ml and Tween 20 to a final concentration of 0.05%) and centrifuged through a silica spin column (Qiagen) using an extender. Once bound to the silica filter, the DNA was purified using the commercially provided buffer (PE) and eluted in 42 µL of TET.

Library building and sequencing

The libraries were built in Oxford from the DNA extracts obtained in Durham or Oxford, without distinctions, using the Blunt End Single Tube (BEST) protocol (39). The amplification of the indexed DNA was optimised using quantitative PCR to estimate the best number of cycles. As expected, the number of required cycles varied largely between archaeological and museum samples. In some instances, we performed multiple parallel amplifications to increase the resulting amount of endogenous DNA. Sequencing of selected libraries was performed on a NovaSeq S4 platform (2×150 bp) at Novogene and Macrogen.

Whole genome enrichment and sequencing

Libraries from the following individuals: ASU_AnSon_1, ASU_ManBac_1, ASU_Taiwan_1, ISU_Philippines_1, ISU_Philippines_3, ISU_UchuPalau_1, ISW_Moluccas_1, POU_Vanuatu_3, POU_Vanuatu_5 were subjected to whole genome enrichment (WGE). Genomic DNA from 3 individuals: i.e. a wild boar from Japan (*Sus scrofa*), a European wild boar (*Sus scrofa*), and a bearded pig (*Sus barbatus*) from the Department of Animal Sciences, University of Illinois, was provided to Daicel Arbor Bioscience for baits construction. The gDNA was visually inspected for color and viscosity and the total DNA was quantified via a spectrofluorimetric assay. WGE baits were prepared from equal masses of the 3 gDNA samples.

Libraries were provided to Daicel Arbor Bioscience for capture. They were quantified via quantitative PCR (qPCR). Based on the quantification results (ranging from 1.9ng-445.6ng), the libraries were reamplified for 4-8 cycles to increase the mass, targeting approximately 1000ng per library. Amplified libraries were quantified again via qPCR. For libraries prepared from the same individual, equal masses of each library were pooled for capture.

Each capture pool was dried down to 7 uL by vacuum centrifugation. Captures were performed following the myBaits v5.02 protocol using the custom Pig WGE baits with an overnight hybridization and washes at 60C. Post-capture, the reactions were amplified for 10-18 cycles and were quantified with a spectrofluorimetric assay. A second round of capture was performed in the same manner. The post-capture material was visualized using the TapeStation 4200 (Agilent) platform with a High Sensitivity D1000 tape. Captures that contained adapter artifact underwent gel excision to remove the artifact. The captures were pooled in approximately equimolar ratios. Samples were screen sequenced on the Illumina NovaSeq 6000 platform on a S4 PE150 lane.

Modern samples

Papua New Guinea, Bhutan, and Sri Lankan (contact: Jaime Gongora)

DNA samples from Bhutanese (SAD_Bhutan_1-4; SAW_Bhutan_6-11), Nepalese (ASD_Nepal_1-3), and PNG (PNU_PNG_6-14) pigs were extracted from whole blood using the QIAamp® DNA Blood Mini Kit; and from hair samples using the Blood and Body Fluid Spin Protocol. DNA from Sri Lankan pigs (SAD_SirLanka_1-4; SAW_SirLanka_5) was extracted using the salting method as described by Miller et al. (40). DNA concentration was determined using ethidium bromide staining and a NanoPhotometer, and ranged from 25-200 ng/μl. All sampling and DNA extraction were done in the respective countries in collaboration with relevant institutions.

With the exception of seven samples (described below), libraries were prepared by the Australian Genome Research Facility (AGRF) as a commercial service using the Illumina Nextera DNA Flex library preparation protocol. Library quality was assessed using a

TapeStation and qPCR, and sequenced by AGRF on a NovaSeq 6000 S4-300 flow cell, v1.5 chemistry.

The remaining seven samples (SAW_Bhutan_6-11; SAW_SriLanka_5) were wild boars from Bhutan and Sri Lanka, collected opportunistically from dead, slaughtered, or captured individuals. DNA from these seven samples was potentially degraded because the tissue was not freshly collected. Consequently, aliquots of purified DNA extracted from these samples were sent from the University of Sydney to the Australian Centre for Ancient DNA (ACAD), University of Adelaide, where libraries were created in a clean room using a protocol optimised for fragmented, low-quality DNA. Specifically, DNA was blunt-ended and had truncated Illumina TruSeq adapters ligated following the protocol of (41). Full length Illumina TruSeq adapters (with a unique non-overlapping 7nt i5/i7 index combination per sample) were added via PCR using primer sequences from (42).

Each library was amplified in eight separate 25 μ L reactions, each comprising 3 μ L of undiluted library, 1 x Platinum Taq DNA Polymerase High Fidelity buffer (ThermoFisher Scientific), 2 mM MgSO₄ (ThermoFisher Scientific), 0.25 mM of each dNTP (ThermoFisher Scientific), 0.4 μ M of each primer, and 0.2 U of Platinum Taq DNA Polymerase High Fidelity (ThermoFisher Scientific), in laboratory grade water. Cycling conditions for the PCR were as follows: 94 °C for 2 min; 20 cycles of 94 °C for 30 s, 60 °C for 30 s, 68 °C for 40 s; and 68 °C for 10 min. PCR products from each library were pooled and purified using 1.1 x volume AxyPrep (Axygen), washed twice with 80% ethanol, and then resuspended in 30 μ L of buffer comprising 10 mM Tris, 0.1 mM EDTA, and 0.05% Tween-20. Library quality was assessed using a Fragment Analyzer (Agilent) and sequenced by AGRF on a NovaSeq 6000 S1-300 flow cell, v1.5 chemistry.

Lanyu pigs (Taiwan; contact: Larry Schook)

DNA was extracted using standard phenol-chloroform techniques. Library preparation and sequencing was carried out at Macrogen (Seoul, South Korea) using an Illumina PCR-free TruSeq library kit (Illumina, Inc) followed by 2x150 bp Paired End analysis on an Illumina NovaSeq instrument.

New Caledonia (contact: Patrick Barriere) and Philippines (contact: Michael Herrera)

DNA extraction was carried out at the PalaeoBARN, University of Oxford. Three hairs per sample were cut into approximately 1 cm long sections and washed in a 0.5% sodium hypochlorite solution. Bleach was removed by rinsing the hairs twice in HPLC grade water followed by two washes in UltraPure DNase/RNase-Free Distilled Water. DNA extraction was carried out using the DNeasy Blood & Tissue Kit (Qiagen), including digestion in Buffer ATL/Proteinase K/DTT solution overnight. Single-stranded DNA sequencing libraries were prepared at the Laboratory of Functional Genome Analysis, Genzentrum, University of Munich (LMU) Swift BioSciences 1S kit. Sequencing: screening on NextSeq at Genzentrum followed by deep sequencing on a 150 PE NovaSeq 6000 S4 flowcell at Macrogen (Seoul, South Korea).

Data processing

Modern genome

Raw reads from modern genomes from both publicly available samples and newly sequenced, were first processed (i.e. adapter/PolyG tail trimmed, quality/length filtered) with fastp using default settings (see <https://github.com/OpenGene/fastp>) (43). Reads were then aligned using BWA-MEM (44), to the Sscrofa11.1 reference genome, duplicates were removed using Picard MarkDuplicates.

Historical and ancient genomes

Historical and ancient genomes were processed separately using the nextflow eager pipeline (45) with default parameters except for the following options: --mapper bwamem, and --clip_readlength 30. This includes pre-processing of reads using AdapterRemoval (46) to remove adapter and collapse pairs and removing PCR duplicates using MarkDuplicates. MapDamage2 (47) was run on the resulting BAM files to assess the level of DNA damage (Fig. S1). Most samples do not exhibit significant deamination patterns, except for the ancient DNA samples, which include the ancient Philippines (ISU_Philippines_1, and 3), Vanuatu (POU_Vanuatu_3), and Palau (ISU_UchuPalau_1) pigs. However, the degradation in the historical samples is evident from the read length distribution (Fig. S2), which shows that most reads are shorter than 200bp.

Mitochondrial analyses

Mitochondrial genome analysis

We used htsbox (<https://github.com/lh3/htsbox>) to create a mitochondrial genome majority consensus fasta file based on BAM files (mapped to the Ssc11.1 reference genome which contains the mitochondrial DNA as a contig), using only reads with a mapping and base quality of 30, and at least 20 base pair long, the first and last 5 base pair were also removed for ancient and historical samples. Fasta files were merged into a single file which was filtered using a custom perl script. Sequences with less than 10x average depth of coverage, over 70% missing data were excluded from this analysis, and positions in the alignment with 30% missing data were also removed. This resulted in an alignment of 585 individuals and 15,523bp. A phylogenetic tree was then constructed with RAxML (48) using 100 (fast) bootstrap replicates and a GTRGAMMA substitution model (Fig. S3).

The Pacific Clade mitochondrial haplogroup was first described using short mtDNA control region sequences (10). To assess the monophyletic strength of this clade, we used complete mitogenomes to build a maximum likelihood phylogeny using 585 mtDNA genomes with >10x coverage. Previously described *S. scrofa* haplogroups, such as European, Near Eastern, East Asian, and a haplogroup including the mtDNA sequences from ISEA endemic *Sus* species (Fig. S3), all formed well-supported clades. We also identified two well-supported clades corresponding to the Pacific Clade and the Lanyu Clade, confirming their identification as monophyletic groups using short-fragment PCR data (10).

Published mtDNA control region dataset

Published mitochondrial control region sequences, many of which belong to the same individuals for which we obtained nuclear data in this study, (N=707, see Supplementary Data S2) were aligned using global iterative refinement (G-INS-i) implemented in MAFFT 7.526 (2024/Apr) (49). The alignment was visualised and a 621-bp segment of the control region was selected using Geneious Prime® 2024.0.7 (GraphPad LLC). IQ-TREE multicore version 1.6.12 (50) was used to construct a maximum likelihood tree under a HKY+F+R4 model of evolution, selected using ModelFit (51). Branch support was assessed using 1000 Ultrafast bootstrap pseudo-replicates (52). The bootstrap consensus tree was drawn in FigTree 1.4.5 (<https://github.com/rambaut/figtree>) and used to assign sequences to one of four haplogroups: Lanyu, Pacific Clade, Europe (European *S. scrofa* in Fig. S3), East Asia (Asian *S. scrofa* in Fig. S3), and ISEA (ISEA-endemic *Sus*, i.e. *S. barbatus*, *S. philipensis*/*S. cebifrons*, *S. verrucosus*, or *S. celebensis* in Fig. S3).

Nuclear analyses

Reference panel genotype call

Genotyping was performed using graph typer v2.7.2 (53) using genomes with at least 5x (500 individuals). The resulting VCF was filtered using the recommended following options (53):

```
vcffilter -f "ABHet < 0 | ABHet > 0.33 & ABHom < 0 | ABHom > 0.97 & MQ > 30"  
(requiring allelic balance of 33% for heterozygous and >97% for homozygous and a  
mapping quality of 30)
```

```
bcftools view -f "PASS" -m 2 -M 2 -v snps  
(selecting on biallelic SNPs flagged as PASS)
```

```
bcftools-1.11/bcftools filter --SnpGap 10 -i "FORMAT/GQ>30" -i "INFO/AC>4"  
(removing every SNPs within 10bp of an indel, and requiring genotyping quality of 30  
and allele count of at over 4 reads).
```

This resulted in a total of ~95M SNPs, including ~25M transversions.

Phasing and imputation

We used SHAPEIT v5 for phasing (27) the reference panel which contains 95M SNPs in 500 individuals. Phasing was performed in 1Mbp sliding windows (100Kb overlap) using the phase_common tool and a recombination map obtained from (54). The resulting BCF were ligated using the ligate tool.

We then used GLIMPSE 2 for imputation (28). We first used the GLIMPSE2_chunk tool to split the phased phase into imputation regions based on the recombination map obtained from (54). Each chunk was then converted into a binary reference format using the GLIMPSE2_split_reference tool. Each BAM file for samples between 1-5x was then imputed separately using GLIMPSE2_phase for each chunk, and chunks were ligated using GLIMPSE2_ligate. The resulting BCFs were then merged with the panel using bcftools. Imputation accuracy was estimated by first downsampling the BAM files of nine individuals to 0.1x, 0.5x, 1x, 2x and 4x using samtools, which were selected to represent the diverse ancestry in our dataset i.e.: ASD_Luchuan_3 and ISD_Lanyu_1 (Asian *S. scrofa*), EUD_Pietrain_4 (European domestic), CEL_Sus-Celebensis_1 (ISEA-endemic *Sus*), INDO_Sumatra_1 and ISW_Bali_1 (Indonesia *S. scrofa*/ISEA-endemic *Sus* hybrid), ISW_Buru_1 and PNU_PNG_10 (Wallacean *S. scrofa*) and POW_NewCaledonia_4 (hybrid Asian and European *S. scrofa*).

We then imputed each bam using a reference panel from which these six individuals were removed. We then calculated concordance between “true” genotypes and imputed genotypes (r^2) at different minor allele frequency (MAF) using the GLIMPSE_concordance tool (Fig. S4). Imputation accuracy was >95% in most individuals from at 2x and MAF>1%. Island Southeast Asian feral individuals from Bali (ISW_Bali_1, ISW_Buru_1), however, only achieved an accuracy of >90% at MAF>5%.

The primary objective of imputation in this analysis is to conduct ancestry deconvolution through local ancestry inference. In addition to testing accuracy of imputation on genotypes, we investigated how imputation influenced our ability to conduct local ancestry inference (LAI) using GNOMIX, to pinpoint genomic segments associated with specific ancestries (see below for details on GNOMIX analysis). We utilized the imputed VCFs derived from downsampled BAM files in the preceding step to conduct a LAI using GNOMIX. For each ancestry, we then compared the regions identified by gnomix in both the full coverage and downsampled data. The results revealed a high degree of overlap between

these regions, ranging from 94% to 99% (Fig. S5). This suggests that imputation minimally impacted the power of local ancestry inference in this context.

Pseudohaploid call

The 25M transversion SNPs discovered in the 5x data set were then genotyped in all historical and ancient genomes (i.e. those below 5x depth of coverage) not included in the panel using ANGSD (v0.933) with the following options: -dohaplocall 1 -minMinor -doCounts 1 -minMapQ 20 -minQ 20 -minInd 1 -setMinDepth 1 -trim 5. Positions at which the genotype did not match one of the two alleles in the 25M SNP panel were set as missing data. Genotype data were merged using plink v1.9 (55). Samples from the panel were pseudohaploised using a custom script in the resulting plink file.

Principal component analysis (PCA)

Principal component analysis (PCA) was carried out using emu (56) on pseudohaploid transversions. We filtered SNPs with MAF<5% and kept all samples with >10,000 SNPs covered (576 individuals).

ADMIXTURE

We first ran an unsupervised ADMIXTURE clustering based on plink file containing data from 120 pseudohaploid genomes from wild individuals from East Asia, Europe and ISEA to identify individual carrying >99% ancestry from three different clusters: Asian *S. scrofa*, European *S. scrofa* and ISEA-endemic *Sus* (Fig. S6). Individuals with at least 99% ancestry from one cluster were then used as a source in a supervised ADMIXTURE analysis (K=3 again), based on 576 individuals.

We also ran an unsupervised ADMIXTURE clustering based on 304 pseudohaploid genomes. All genomes were included in this analysis, except European domestic pigs and no more than 3 individuals per Asian breed. We used European wild as a proxy for European ancestry rather than European domestic pigs because many of the latter possess Asian ancestry (57) At K=3 (best K based on cross-validation), the results (Fig. S7) mirrored those in Fig. S6 (only using wild boar), showing the impact of European ancestry (blue) in Asian domestic (ASD) and some individuals East of the Wallace Line (see also the blue component in Fig. 2), and the impact of ISEA-endemic *Sus* ancestry (black) in ISEA individuals from both East and West of the Wallace Line (see also Fig. 2).

At K=5 (second best K based on cross-validation) this analysis separated ISEA individuals East (mostly possessing the red ancestry component) and West of the Wallace Line (mostly possessing the light green ancestry component).

D and F statistics

Our ADMIXTURE analysis with 120 wild pig genome did not possess enough power to split the ancestry of ISEA-endemic *Sus* individual i.e. *S. celebensis*, *S. cebifrons*, *S. verrucosus*, or *S. barbatus*. Reducing the number of *S. scrofa* individuals in ADMIXTURE to 25 allowed for the separation of each species ancestry at K=9 (Fig. S8) demonstrating that this lack of power in our full analysis is likely to be due to an ascertainment bias due to the comparatively smaller number of ISEA-endemic *Sus* genomes in our dataset (n=23).

Therefore, for each ISEA individual that possessed both Asian *S. scrofa* and ISEA-endemic *Sus* based on our ADMIXTURE analysis we first used D-statistic to assess which ISEA-endemic *Sus* best matched their non-*scrofa* ancestry. To do so we first validated that each individual yielded significant D-statistics of the form D(Babyrousa outgroup, ISEA-endemic *Sus*; Test, Asian *S. scrofa*), where ISEA-endemic *Sus* represent any genome of *S.*

celebensis, *S. cebifrons*, *S. verrucosus*, or *S. barbatus* and Asian *S. scrofa* any genome of individuals that possess >99% Asian *S. scrofa* ancestry in our ADMIXTURE analysis.

We then ran all combinations of D-statistics of the form D(B. babyrousa, Test; ISEA-endemic-Sus-Y, ISEA-endemic-Sus-Z), where Test is the individual possessing *S. scrofa* and ISEA-endemic *Sus* ancestry, and ISEA-endemic-Sus-Y, and ISEA-endemic-Sus-Z represent all combinations of ISEA-endemic *Sus* individual i.e. *S. celebensis*, *S. cebifrons*, *S. verrucosus*, or *S. barbatus*. We choose the individual species with the highest D value as the source of ISEA-endemic *Sus* ancestry which was used for the plot in Fig. 2.

Admixture graphs

We suspected that Western Indonesian mixed *S. scrofa* / ISEA-endemic *Sus* (e.g. from Sumatra and nearby islands) possess *S. scrofa* ancestry that is poorly represented by our panel. To address this, we constructed admixture graphs using Admixtools2 (<https://github.com/ukrmaiel/admixtools>), to assess the phylogenetic placement whether the *S. scrofa* ancestry found in these individuals is nested within Asian *S. scrofa* or a sister group to Asian and European *S. scrofa* (Fig. S9).

We constructed “skeleton graphs”, tailored to for each *S. scrofa* ISEA genome, informed by the D-statistic pipeline above. For individuals with no detectable I ISEA endemic-Sus ancestry, we constructed admixture graphs containing a *B. babyrousa* as an outgroup, an individual with Asian *S. scrofa* ancestry, an individual with European *S. scrofa* ancestry, and an individual with South Asian *S. scrofa* ancestry (i.e. Sri Lanka or India). The graph was created using the find_graphs function (plusminus_generations = 5, stop_gen = 50, stop_gen2 = 10). We consistently identified the same topology: (Babyrousa, (South Asian, (Asian, European)))

For individuals with detectable ISEA endemic-Sus ancestry we carried out the same as above, but included a genome corresponding to the relevant non-scrofa species (i.e. the species that was the best proxy for the ISEA endemic-Sus ancestry based on D-statistics). This also consistently identified the same topology: (Babyrousa, (ISEA endemic-Sus, (South Asian, (Asian, European))))

We then added each test genome (i.e. *S. scrofa* genome from ISEA) to its corresponding skeleton graph from 1a or 1b, above, in two different locations to create two admixture graphs for each test genome:

- basic graph 1a/1b-Asian: On the Asian branch, i.e. (Babyrousa, (South Asian, ((Asian, test), European))), 1a; or (Babyrousa, (ISEA endemic-Sus, (South Asian, ((Asian, test), European))))), 1b
- basic graph 1a/1b-Sister: On the basal branch, i.e. (Babyrousa, (ISEA endemic-Sus, (South Asian, (test, (Asian, European)))))) or (Babyrousa, (South Asian, (ISEA endemic-Sus, (Asian, European))))

For the basic graphs for individuals with detectable non-*scrofa* ancestry (basic graph 1b-Asian & 1b-Sister), we added an admixture edge between ISEA endemic-*Sus* and our test genome.

For each individual (test), we then assessed whether the Asian or Sister graph was the best fit, using 100 bootstrap-resampled graph fits, after computing out of sample scores to fairly compare models of different complexity to each other.

The ratio of scores for the two admixture graphs were plotted in Fig. S9, as:

$$(Score_Asian - Score_Sister) / Max[Score_Asian, Score_Sister]$$

Local ancestry reconstruction (GNOMIX)

Local ancestry reconstruction was performed using GNOMIX (29). Wild individuals with Asian *S. scrofa*, European *S. scrofa* and non-*scrofa* ancestry that were used as a source in the supervised ADMIXTURE above were used as reference for this analysis.

We trained two models, each with two sources:

- *S. scrofa* (using European *S. scrofa*) and non-*scrofa*
- European *S. scrofa* and Asian *S. scrofa*.

For model 1, we did not use Asian *S. scrofa* to discriminate between *scrofa* and non-*scrofa* ancestry as previous studies have suggested that mainland Southeast Asian wild boar (*S. scrofa*) possess some small degree of ISEA-endemic *Sus* ancestry (3, 4).

We ran GNOMIX with default settings except for the time of admixture used for validation (generation since admixture: 2, 4, 6, 8, 12, 16, 24, 48). Both models achieved training accuracy and validation accuracy > 99%. Local ancestry reconstruction was then performed on *S. scrofa* individuals from Asia that were either phased (>5x) or imputed (>1x) and which were showed to possess either European *S. scrofa* or non-*scrofa* ancestry (non possessed both), based on the ADMIXTURE analysis, using the appropriate model (model 1 for non-*scrofa* ancestry, model 2 in case of European *S. scrofa* ancestry). See the section on imputation above for details about the results of the test for accuracy of imputation for LAI.

Ancestry deconvolution

We used the GNOMIX results to identify regions of Asian *S. scrofa* ancestry in the genome of *S. scrofa* individuals with mixed ancestry, i.e. possessing either non-*scrofa* and Asian *S. scrofa* (GNOMIX model 1) or European *S. scrofa* and Asian *scrofa* ancestry (GNOMIX model 2). To do so, we selected regions which were identified with high confidence (probability >99%) by GNOMIX as either homozygous *S. scrofa* (model 1) or homozygous Asian *S. scrofa* (model 2). For each mixed individual, we extracted these regions from the pseudohaploid PLINK file used for PCA, ADMIXTURE, and D-statistics analyses described above.

We then calculated admixture fraction using f4 ratio to assess the proportion of non Asian *S. scrofa* ancestry left in the genomes of mixed individuals after deconvolution (Fig. F4-deconvolution). European ancestry in most (14/16) individuals from New Caledonia, Hawaii, and Papua New Guinea decreased from ~50-25% to 0-5%. European ancestry in one individual from New Caledonia, however, remained at ~10%, at ~20% in an individual from Papua New Guinea. European ancestry in Lanyu pigs decreased from ~25% to 0-1%, while European ancestry in domestic pigs from the Philippines decreased from 30-70% to 0-15%.

Non-*scrofa* ancestry in individuals with lower *S. scrofa* ancestry such as pigs from Sibago or pigs East of the Wallace line, including Flores, Timor, and Halmhaera decreased from between ~80-40% to below 5%. The non-*scrofa* ancestry in wild boars from Western Indonesia (West of the Wallace line), including Sumatra and nearby islands, however, decreased from between 14-5% to ~5%. The lower success of deconvolution in these individuals can likely be explained by ancient admixture processes (18) which would be found in smaller blocks of non-*scrofa* ancestry across the genome that are undetectable by GNOMIX. In contrast, due to its recent introduction in the region, European *S. scrofa* ancestry is found in larger blocks (Fig. 3B), which are more easily detectable.

We also conducted a PCA using both full genome and deconvoluted data using all *S. scrofa* individuals from East Asia (n=228). Fig. S10A, which includes all Asian *S. scrofa* individuals without ancestry deconvolution, reveals a separation between populations east and west of the Wallace Line. Some individuals from the East, however, such as individuals

from Timor and Halmhaera, cluster closer to pigs from west of the Wallace line likely reflecting the varying degrees of ISEA-endemic ancestry. These individuals instead plot close to pigs East of the Wallace Line following ancestry deconvolution (Fig. S10B).

This Fig. highlights that *S. scrofa* populations on either side of this biogeographic boundary possess markedly different ancestry profiles, despite their relative geographic proximity. Populations east of the Wallace Line show a closer affinity to mainland domestic populations, particularly those from Southeast China such as Luchuan, Wuzhishan, and Bamaxiang.

outgroup f3

We calculated outgroup-f3 statistics using calc-f3 from the structf4 package (58) and babirusa as an outgroup. The goal of this analysis is to assess the degree to which Lapita pigs from Melanesia share drift with other populations of *S. scrofa*. To do so we first chose a 2,500years old ancient Vanuatu genome (POU_Vanuatu_3; ~0.7x) as reference and computed all pairwise outgroup-f3 involving this individual.

We found that feral pigs from Papua New Guinea (e.g. ISW_PNG_18; ~2.2x) shared the most drift with this individual (Fig. S11). ADMIXTURE analysis of ISW_PNG_18 did not detect any evidence of European ancestry, making it a good proxy for Lapita pig ancestry. We used this individual as reference in subsequent outgroup-f3 analyses due to its higher coverage than POU_Vanuatu_3 to limit issues arising when comparing multiple individuals with low breadth of coverage, particularly after ancestry deconvolution. We then computed all pairwise outgroup-f3 involving ISW_PNG_18 using both the full and deconvoluted dataset.

Admixture time (DATES)

We used DATES (26) to estimate time of admixture in admixed Island Southeast Asian individuals using the recombination map obtained from (54) and the following parameters:

```
binsize: 0.001  
maxdis: 1  
jackknife: YES  
qbin: 10  
runfit: YES  
affit: YES  
loalfit: 0.45
```

The same individuals used to train the GNOMIX models were used as source for this analysis. We used a generation time of 3 years to convert into years - admixture date were subtracted to age of samples (specific age for historical genomes in Data S1 and using year 2000 for modern samples). Results were considered significant at first if Z-score > 2 and normalized root-mean-square deviation (NRMSD) < 0.7. We first assessed the accuracy of DATES using five European domestic individuals with known dates of admixture with Asian *S. scrofa* (as a result of 19th century improvement of pig stock in Europe). We modelled these individuals using the same source as for the ADMIXTURE and GNOMIX analyses.

All results were significant and visual inspection of the exponential distribution fitted and the covariance decay curve suggested a good fit (Fig. S12). The recovered admixture proportion and admixture dates admixture dates fitted well with the known history of these breeds (admixture date in the middle of the 19th century (59)):

| Sample ID | Sources | EUW | ASW | Mean time (generation) | SE | Z | NRMSD | Mean time (years) |
|------------------|---------|-------|-------|------------------------|------|-----|-------|-------------------|
| EUD_Duroc_1 | EUW-ASW | 0.78 | 0.22 | 55.5 | 15.0 | 3.7 | 0.083 | 1833 |
| EUD_Landrace_11 | EUW-ASW | 0.784 | 0.216 | 39.5 | 8.5 | 4.6 | 0.058 | 1881 |
| EUD_Pietrain_10 | EUW-ASW | 0.772 | 0.228 | 46.8 | 8.1 | 5.8 | 0.069 | 1860 |
| EUD_Tamworth_3 | EUW-ASW | 0.831 | 0.169 | 59.9 | 14.4 | 4.1 | 0.07 | 1820 |
| EUD_Yorkshire_19 | EUW-ASW | 0.758 | 0.242 | 53.7 | 7.8 | 6.9 | 0.073 | 1839 |

We modelled individual from Oceania, Melanesian and the Philippines using the same approach:

| Sample ID | Sources | EUW | ASW | Mean time (generation) | SE | Z | NRMSD | Mean time (years) |
|--------------------|---------|-------|-------|------------------------|--------|--------|-------|-------------------|
| ISD_Lanyu_1 | EUW-ASW | 0.351 | 0.649 | 6.221 | 1.987 | 3.131 | 0.137 | 1981 |
| ISD_Lanyu_2 | EUW-ASW | 0.29 | 0.71 | 8.63 | 7.832 | 1.102 | 0.077 | 1974 |
| ISD_Lanyu_3 | EUW-ASW | 0.319 | 0.681 | 3.147 | 6.363 | 0.495 | 0.094 | 1991 |
| ISD_Lanyu_4 | EUW-ASW | 0.342 | 0.658 | -2.435 | 10.307 | -0.236 | 0.136 | 2007 |
| ISD_Lanyu_5 | EUW-ASW | 0.308 | 0.692 | -5.063 | 13.104 | -0.386 | 0.115 | 2015 |
| ISU_Philippines_10 | EUW-ASW | 0.341 | 0.659 | 18.075 | 2.469 | 7.321 | 0.087 | 1946 |
| ISU_Philippines_11 | EUW-ASW | 0.304 | 0.696 | 38.298 | 15.299 | 2.503 | 0.107 | 1885 |
| ISU_Philippines_12 | EUW-ASW | 0.647 | 0.353 | 16.761 | 11.669 | 1.436 | 0.073 | 1950 |
| ISU_Philippines_13 | EUW-ASW | 0.739 | 0.261 | 24.639 | 3.955 | 6.229 | 0.091 | 1926 |
| ISU_Philippines_14 | EUW-ASW | 0.696 | 0.304 | 36.515 | 11.697 | 3.122 | 0.088 | 1890 |
| ISU_Philippines_15 | EUW-ASW | 0.753 | 0.247 | 18.08 | 9.237 | 1.957 | 0.078 | 1946 |
| ISU_Philippines_4 | EUW-ASW | 0.673 | 0.327 | 6.702 | 5.378 | 1.246 | 0.166 | 1980 |
| ISU_Philippines_5 | EUW-ASW | 0.54 | 0.46 | 18.577 | 8.305 | 2.237 | 0.077 | 1944 |
| ISU_Philippines_6 | EUW-ASW | 0.739 | 0.261 | 10.952 | 16.668 | 0.657 | 0.109 | 1967 |
| ISU_Philippines_7 | EUW-ASW | 0.648 | 0.352 | 40.351 | 11.847 | 3.406 | 0.087 | 1879 |
| ISU_Philippines_8 | EUW-ASW | 0.475 | 0.525 | 36.789 | 9.636 | 3.818 | 0.067 | 1890 |
| ISU_Philippines_9 | EUW-ASW | 0.791 | 0.209 | 63.41 | 11.33 | 5.597 | 0.05 | 1810 |
| ISW_Marquesas_1 | EUW-ASW | 0.639 | 0.361 | 34.074 | 9.877 | 3.45 | 0.096 | 1898 |
| ISW_Morotai_1 | EUW-ASW | 0.177 | 0.823 | 30.923 | 6.229 | 4.964 | 0.074 | 1810 |
| ISW_Morotai_2 | EUW-ASW | 0.186 | 0.814 | 30.347 | 6.181 | 4.91 | 0.072 | 1812 |
| ISW_PNG_20 | EUW-ASW | 0.208 | 0.792 | 16.317 | 5.572 | 2.929 | 0.091 | 1922 |

| | | | | | | | | |
|--------------------|---------|-------|-------|--------|--------|-------|-------|------|
| ISW_Sumba_1 | EUW-ASW | 0.453 | 0.547 | 17.65 | 2.946 | 5.991 | 0.189 | 1877 |
| PNU_PNG_10 | EUW-ASW | 0.424 | 0.576 | 26.165 | 4.892 | 5.349 | 0.058 | 1922 |
| PNU_PNG_11 | EUW-ASW | 0.421 | 0.579 | 7.609 | 8.702 | 0.874 | 0.108 | 1977 |
| PNU_PNG_12 | EUW-ASW | 0.544 | 0.456 | 25.227 | 3.32 | 7.599 | 0.117 | 1924 |
| PNU_PNG_13 | EUW-ASW | 0.571 | 0.429 | 25.036 | 8.263 | 3.03 | 0.076 | 1925 |
| PNU_PNG_14 | EUW-ASW | 0.482 | 0.518 | 17.289 | 8.536 | 2.025 | 0.078 | 1948 |
| PNU_PNG_6 | EUW-ASW | 0.487 | 0.513 | 6.311 | 17.201 | 0.367 | 0.101 | 1981 |
| PNU_PNG_7 | EUW-ASW | 0.334 | 0.666 | 21.447 | 10.853 | 1.976 | 0.077 | 1936 |
| PNU_PNG_8 | EUW-ASW | 0.418 | 0.582 | 30.253 | 13.904 | 2.176 | 0.073 | 1909 |
| PNU_PNG_9 | EUW-ASW | 0.488 | 0.512 | 14.085 | 4.373 | 3.221 | 0.13 | 1958 |
| POW_Hawaii_1 | EUW-ASW | 0.585 | 0.415 | 40.041 | 8.144 | 4.916 | 0.078 | 1817 |
| POW_NewCaledonia_1 | EUW-ASW | 0.57 | 0.43 | 23.775 | 4.149 | 5.73 | 0.081 | 1942 |
| POW_NewCaledonia_2 | EUW-ASW | 0.462 | 0.538 | 33.71 | 7.245 | 4.653 | 0.081 | 1912 |
| POW_NewCaledonia_3 | EUW-ASW | 0.544 | 0.456 | 41.317 | 8.477 | 4.874 | 0.071 | 1889 |
| POW_NewCaledonia_4 | EUW-ASW | 0.544 | 0.456 | 30.252 | 9.299 | 3.253 | 0.082 | 1922 |
| POW_NewCaledonia_5 | EUW-ASW | 0.587 | 0.413 | 23.563 | 15.314 | 1.539 | 0.08 | 1942 |
| POW_NewCaledonia_6 | EUW-ASW | 0.45 | 0.55 | 31.695 | 7.888 | 4.018 | 0.045 | 1918 |

Most significant ($Z > 2$) results points to dates in from the early 18th century to the mid 20th century. We attempted to estimate time of admixture between *S. scrofa* and endemic-Sus in Western Indonesia populations but visual inspection of the exponential distribution fitted and the covariance decay curve suggested a poor fit (Fig. S13). This is likely due to the fact that there are only the low number of individual genomes available for each ISEA-Endemic *Sus* species and possibly that the Asian *S. scrofa* population used here is a poor proxy for the ancestry in these populations;

Conditional heterozygosity

We calculated conditional heterozygosity (θ) to obtain estimates of heterozygosity. This approach offers the advantage of enabling comparisons across datasets with varying sequencing coverage. Conditional heterozygosity can also be computed post-ancestry deconvolution. To do so, we ascertained heterozygous sites in a $\sim 29x$ *S. cebifrons* genome (CEB_Sus-Cebifrons_10) to identify 554,543 heterozygous transversions. We chose this species as it is an outgroup to *S. scrofa* and not involved in admixture with *S. scrofa*. This strategy effectively mitigates potential biases arising from the variable levels of sequencing coverage inherent in our ancient, historical, and modern data.

For each *S. scrofa* (or hybrid) individual with $> 1x$ depth of coverage in our dataset, we randomly sampled two reads per site and computed the number of sites at which each alleles differed, excluding sites which possess alleles not present in the *S. cebifrons* individual. We recomputed this metric using homozygous tracks of Asian *S. scrofa* ancestry (i.e. masking regions of the genome with European *S. scrofa* or ISEA-endemic *Sus* identified by GNOMIX).

The result shows the influence of ancestry deconvolution, which substantially reduces heterozygosity levels (Fig. S14). This effect is particularly pronounced in individuals with ancestry derived from both Asian *S. scrofa* and ISEA-endemic *Sus* species, such as those from the islands of Timor (e.g. ISW_Timor_3 and ISW_Timor_8) or Halmahera (ISW_Halmahera_1). These individuals exhibit markedly inflated heterozygosity, which was effectively normalized when excluding their ISEA-endemic *Sus* ancestry, bringing their heterozygosity levels within the typical range of Eastern ISEA populations.

This analysis shows that with the exception of the insular domestic Lanyu pigs (a small population restricted to Taiwan), populations east of the Wallace Line possess lower heterozygosity compared to other Asian populations, including mainland Asian domestic pigs, from which they originated (wilcoxon test, $W=2332$, $p<0.001$).

Geometric Morphometric (GMM)

Data acquisition

Information on the specimens analysed with geometric morphometrics can be found in Data S3. This study used geometric morphometrics to analyze the shape and size of third lower molars. Digital images of the teeth were taken from above (occlusal view) using a standardized protocol (61). Twelve anatomical landmarks and 87 sliding semi-landmarks (Fig. S15) were then placed on these images using TPSdig software (62). Coordinates were superimposed using a General Procrustes Analysis (function ProcSym of the R package Morpho (63)). During this procedure the position of the SSL were adjusted using the Procrustes distance (bending = FALSE) and the original configuration was used in all iterations (recursive=FALSE). The centroid size and Procrustes residuals (coordinates after superimposition) were used respectively to assess tooth size and variation.

Predictive Discriminant Analysis

In order to identify the ancient teeth corresponding to *S. scrofa* several regional predictive linear discriminant analyses were performed. The identification was based on a resampling approach based on groups of the same size combined with a data reduction procedure selecting the first components (PC) of a Principal Component Analysis (PCA) that maximise the between group differentiation (64). Among the 100 analyses computed, only the discriminant analyses with the highest correct cross-validation percentages (CVP, jackknife) were retained (upper quartile of the distribution). The specimens of *S. scrofa* included in those identifications originated from China, India, Thailand, Vietnam, Malaysia, Burma, Sumatra and Java. We then classified individual samples from Island Southeast Asia using this approach.

In Sulawesi, the 4 ancient specimens could correspond either to *S. scrofa* (N=97) or *S. celebensis* (N=93). The two modern species have a correct cross-validation of 96% (90% confidence interval CI: 95.7%-96.8%) using the 17 first PCs carrying 91.3% of the total variance. For the prediction, 67 discriminant analyses were retained all with a CVP above 96%. The four specimens were identified as *Sus scrofa*.

In Sarawak (Borneo), 26 archaeological specimens were compared to 67 *S. barbatus* and 97 *S. scrofa*. The CVP of the two species is 90.7% (CI: 88.7-93.2) based on 12 PCS carrying 85.9% of the total variance. 39 discriminant analyses were retained, all with a CVP above 91%. 13 archaeological specimens were identified as *S. barbatus* and 13 to *S. scrofa*.

For the Philippines, the 48 archaeological specimens were identified as either *S. scrofa* (N=97) or ISEA endemic *Sus* (N=38, including *S. cebifrons* (N=8), *S. oliveri* (N=3), and *S. philippensis* (N=27)). The two modern taxa have a CVP of 92.7% (CI: 88.1-97.4%) based on 13 PCs (% of total variance). 33 discriminant analyses were retained resulting in the identification of 35 *S. scrofa* and 13 endemic *Sus*.

In Flores, 6 archaeological specimens were identified as either *S. celebensis* (N=93) or *S. scrofa* (N=97). The cross validation between the two species was 96.3% (CI: 96.2, 96.8%) based on 13 PCs (% of variance). 97 discriminant analyses were retained, with a CVP above 96.2%, that allowed to identify 5 *S. celebensis* and 1 *S. scrofa*.

The 147 archaeological specimens from mainland South-East Asia (China, Vietnam, Thailand) strongly differ from the 31 from the far east of the distribution (Marqueses, Vanuatu). Between those two groups the correct cross validation is 97% (CI: 92.3-100%) based on the first 10 PCs (83% of total variance). The Wallacean and Pacific specimens exhibited a third lower molar proportionally wider and shorter compared to the more rectangular shape of the specimens from mainland Asia (Fig. S16). Based on those PCs and the predictive discriminant analysis approach described above (see (64)) the remaining 102 specimens were identified based on 42 discriminant analyses with a CVP above 98%.

The morphometric proximity between ancient and modern groups from the various regions and taxonomy was visualised using Mahalanobis distances obtained from the 15 first PC scores (88.7% of total variance) that maximise the between group differences (Fig. 1C).

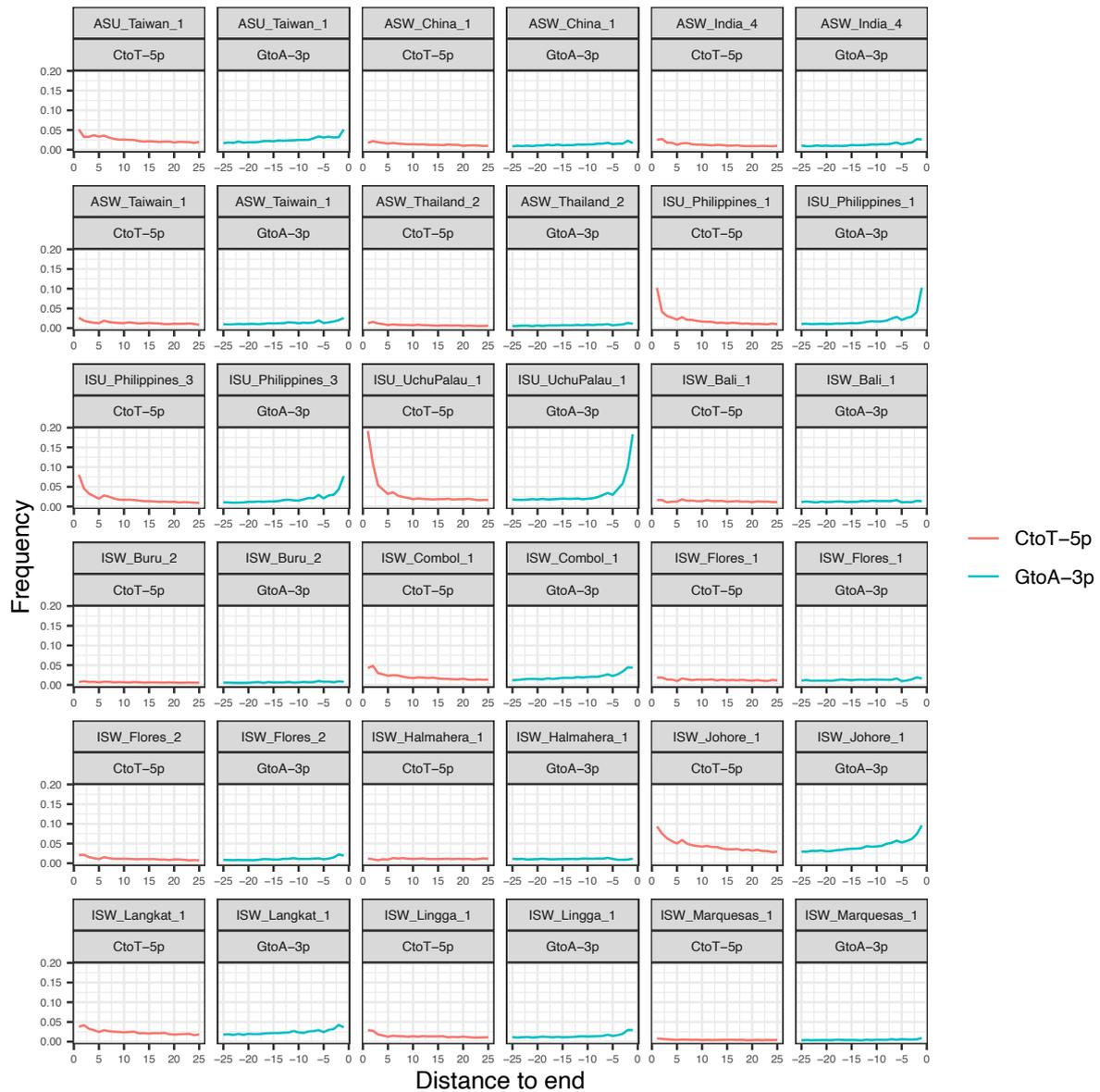


Fig. S1

C-to-T (left, red) and G-to-A (right, blue) substitution frequencies estimated for ancient and historical samples obtained from MapDamage2.

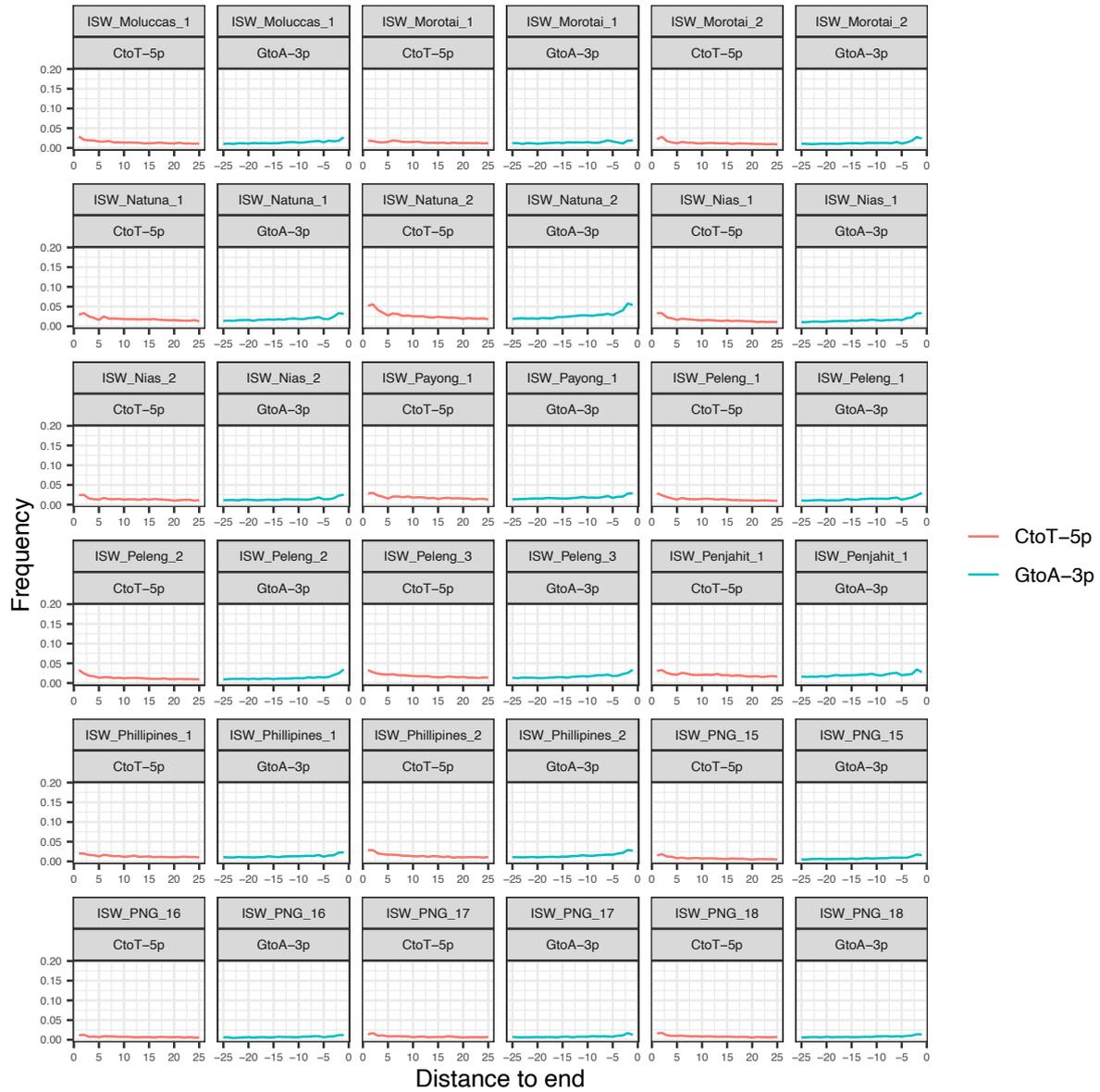


Fig. S1 (continued)

C-to-T (left, red) and G-to-A (right, blue) substitution frequencies estimated for ancient and historical samples obtained from MapDamage2.

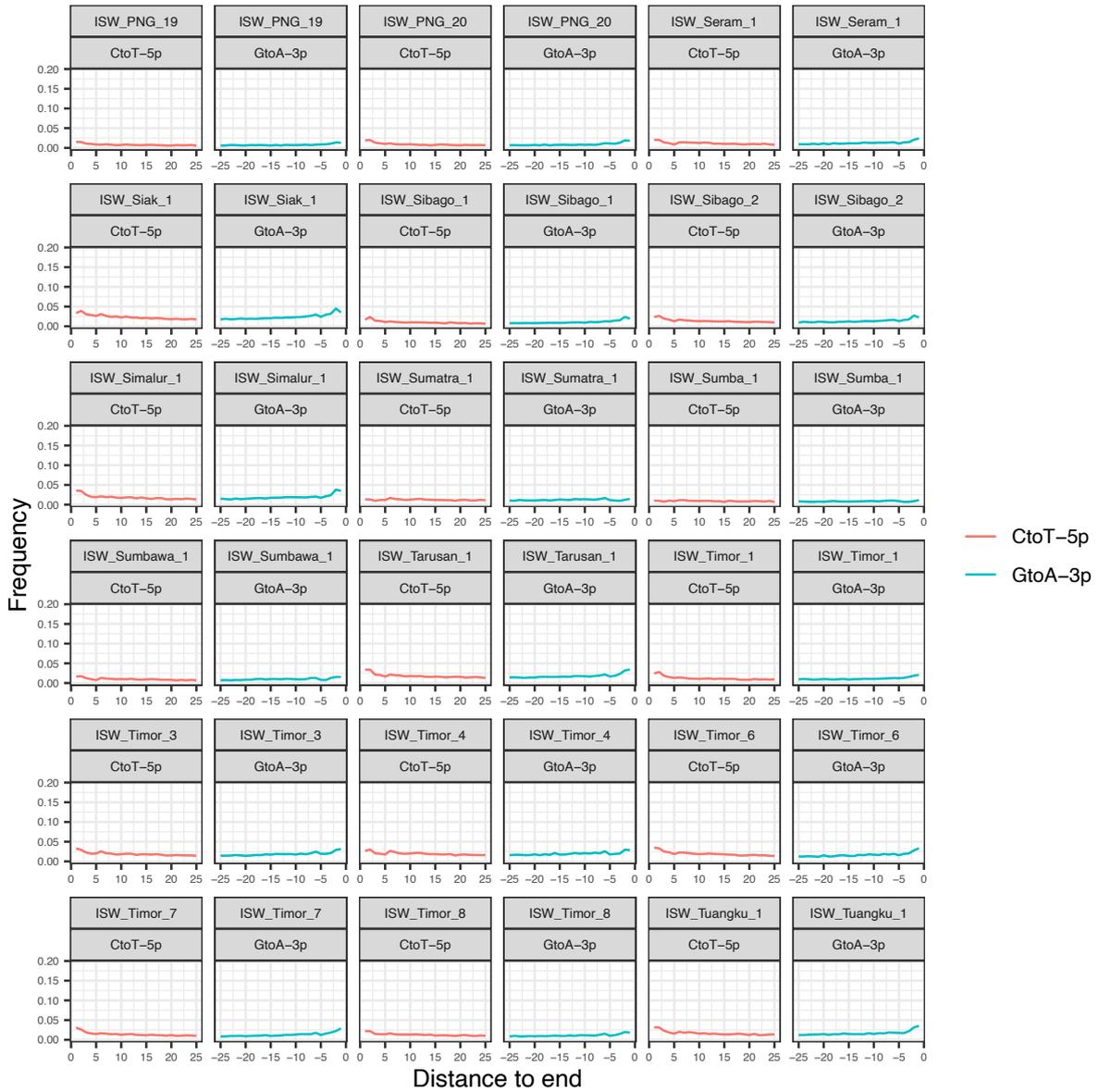
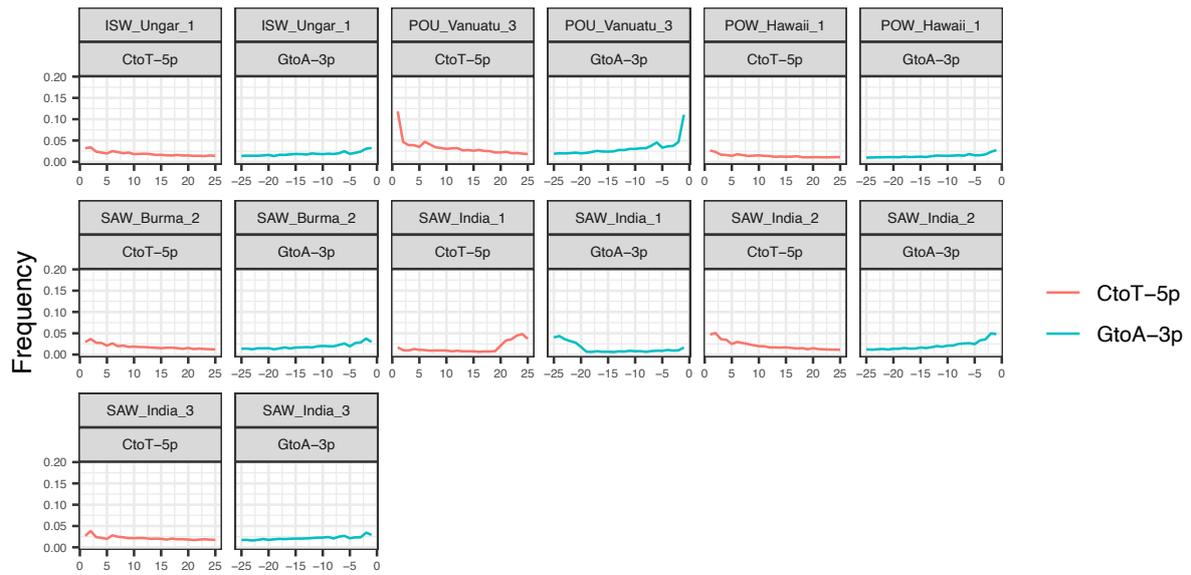


Fig. S1 (continued)

C-to-T (left, red) and G-to-A (right, blue) substitution frequencies estimated for ancient and historical samples obtained from MapDamage



Distance to end

Fig. S1 (continued)

C-to-T (left, red) and G-to-A (right, blue) substitution frequencies estimated for ancient and historical samples obtained from MapDamage2

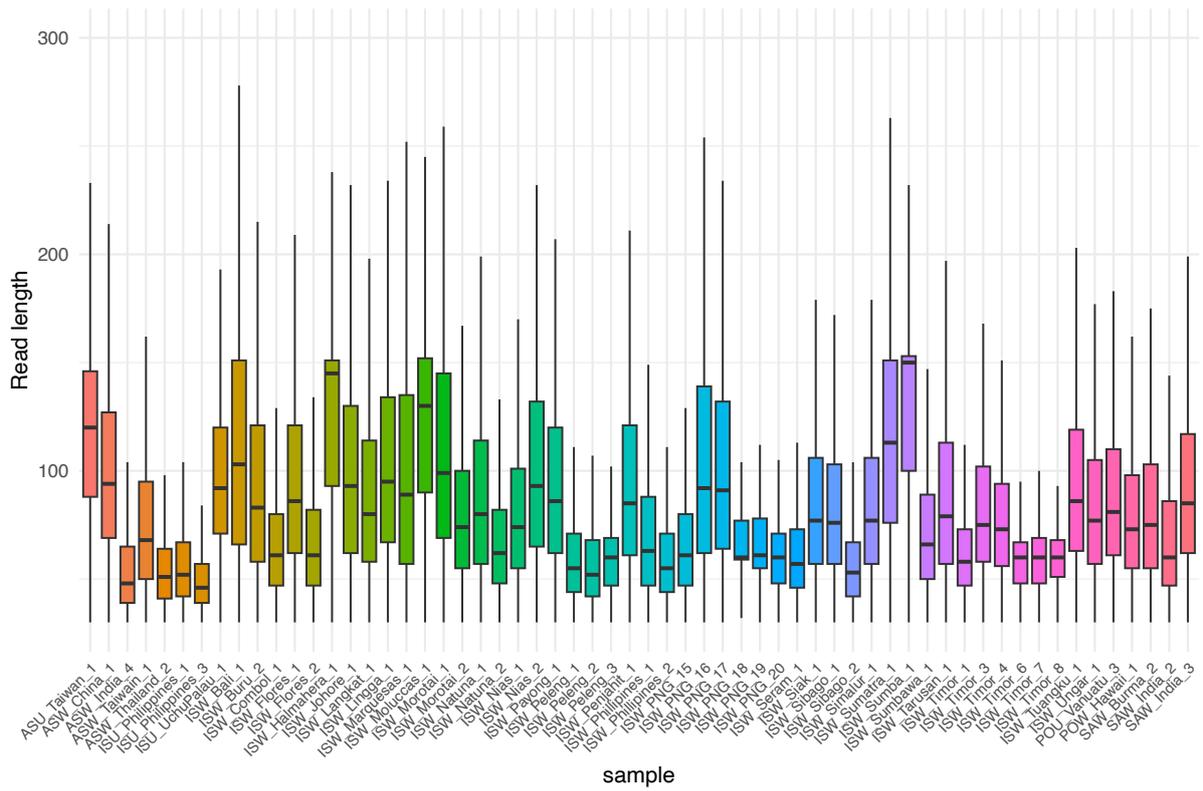


Fig. S2.
 Distribution of mapped read length for each ancient and historical sample (x-axis).

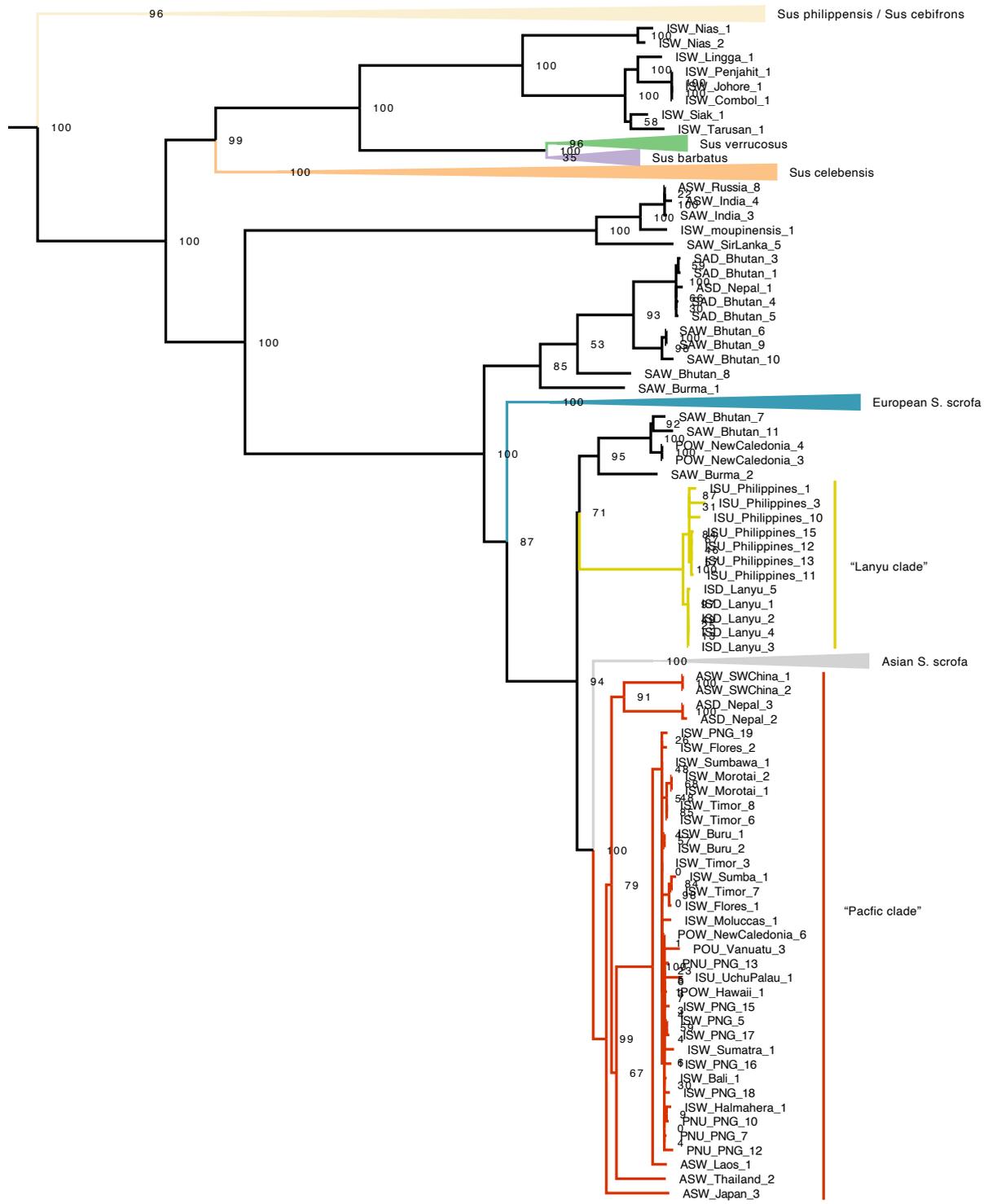


Fig. S3. Maximum likelihood tree based on 585 mitogenomes with bootstrap support.

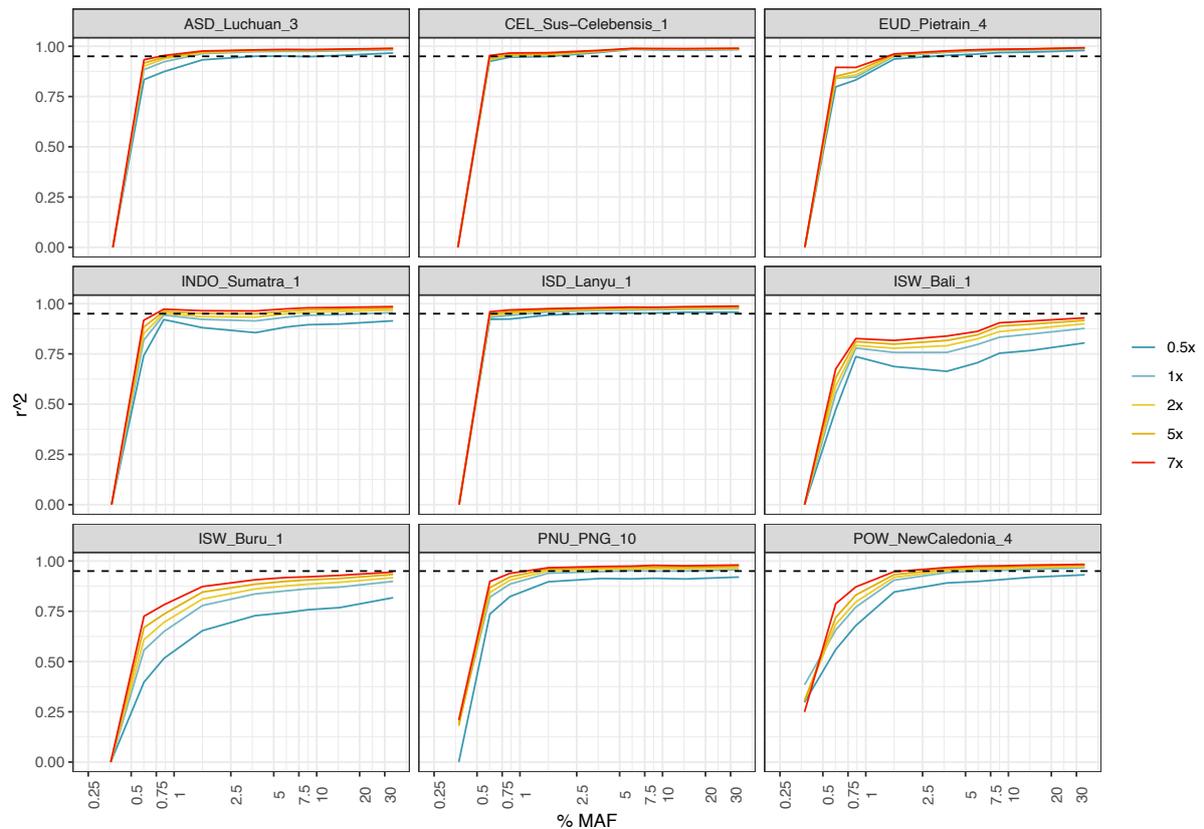


Fig. S4.

Imputation accuracy at different coverage in 9 individuals, ASD_Luchuan_4: Southeast Chinese domestic pig (*S. scrofa*), CEL_Sus-Celebensis_1: *S. celebenis*, EUD_Pietrain_4: European domestic pig (*S. scrofa*), INDO_Sumatra_1: Sumatra wild pig (*S. scrofa*), ISD_Lanyu_1: Lanyu (domestic) pig (*S. scrofa*), ISW_Bali_1: Bali wild pig (*S. scrofa*), ISW_Buru1: Buru (Wallacea), wild pig (*S. scrofa*), POW_NewCaledonia_4: New Caledonian wild pig (*S. scrofa*). The x-axis represents MAF (minor allele frequency), y-axis represents concordance between “true” genotypes and imputed genotypes (r^2) at different downsampled coverage (0.5-7x) as computed by the GLIMPSE_concordance tool.

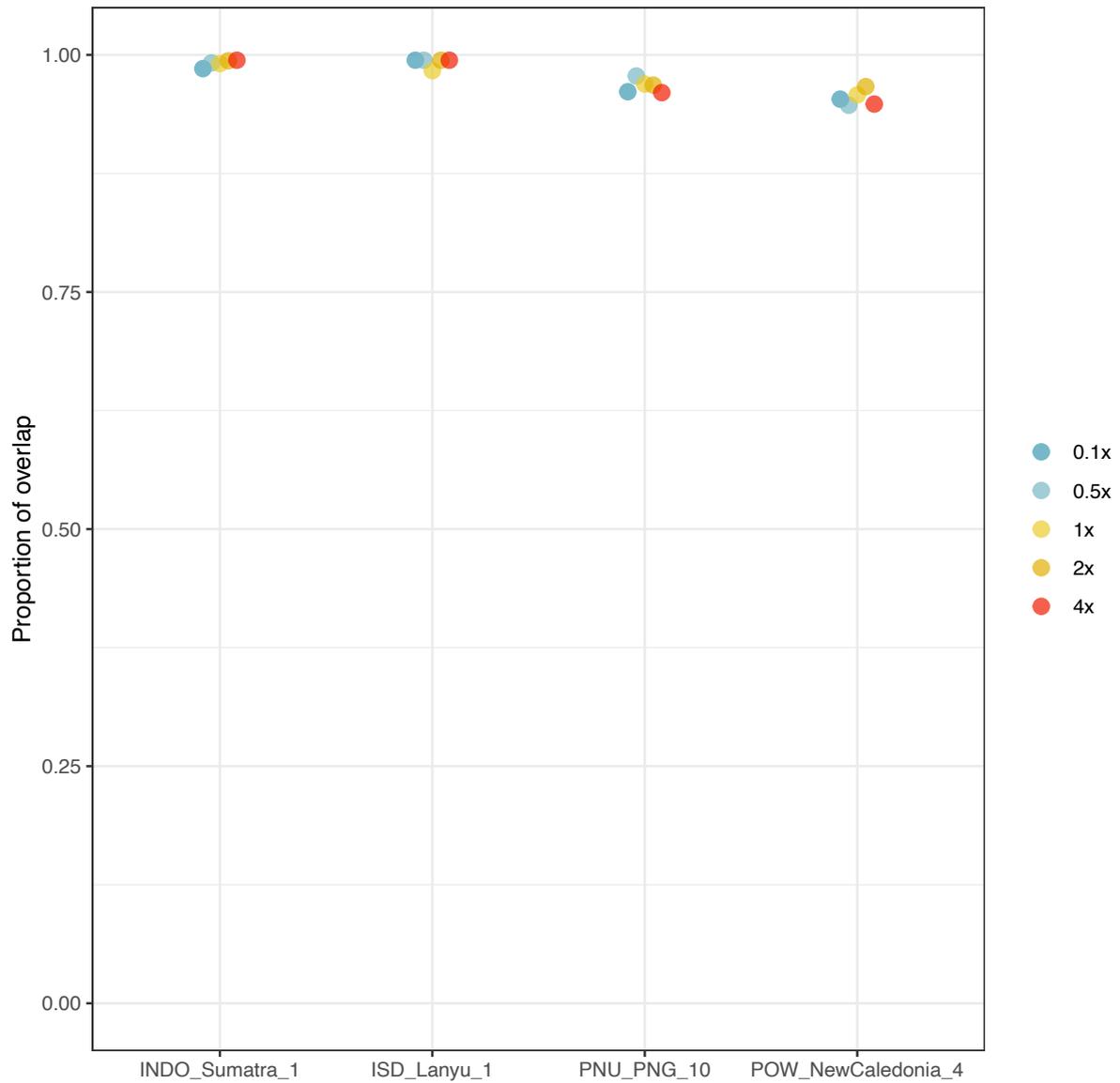


Fig. S5.

Accuracy of local ancestry inference across different sequencing coverages. The y-axis shows the proportion of matching ancestry calls by GNOMIX between high-coverage data and downsampled data 0.5x to 4x coverage.

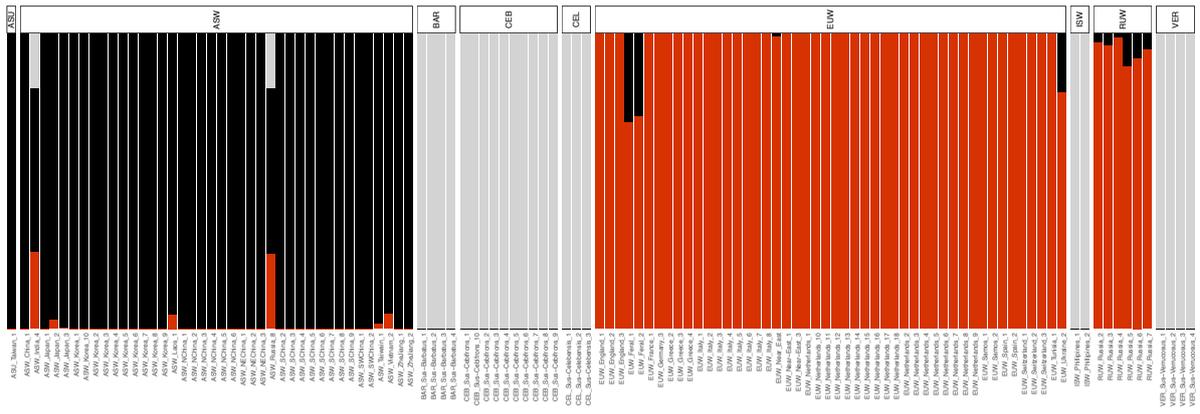


Fig. S6.
Results of unsupervised ADMIXTURE analysis using K=3 based 120 wild boar genomes.

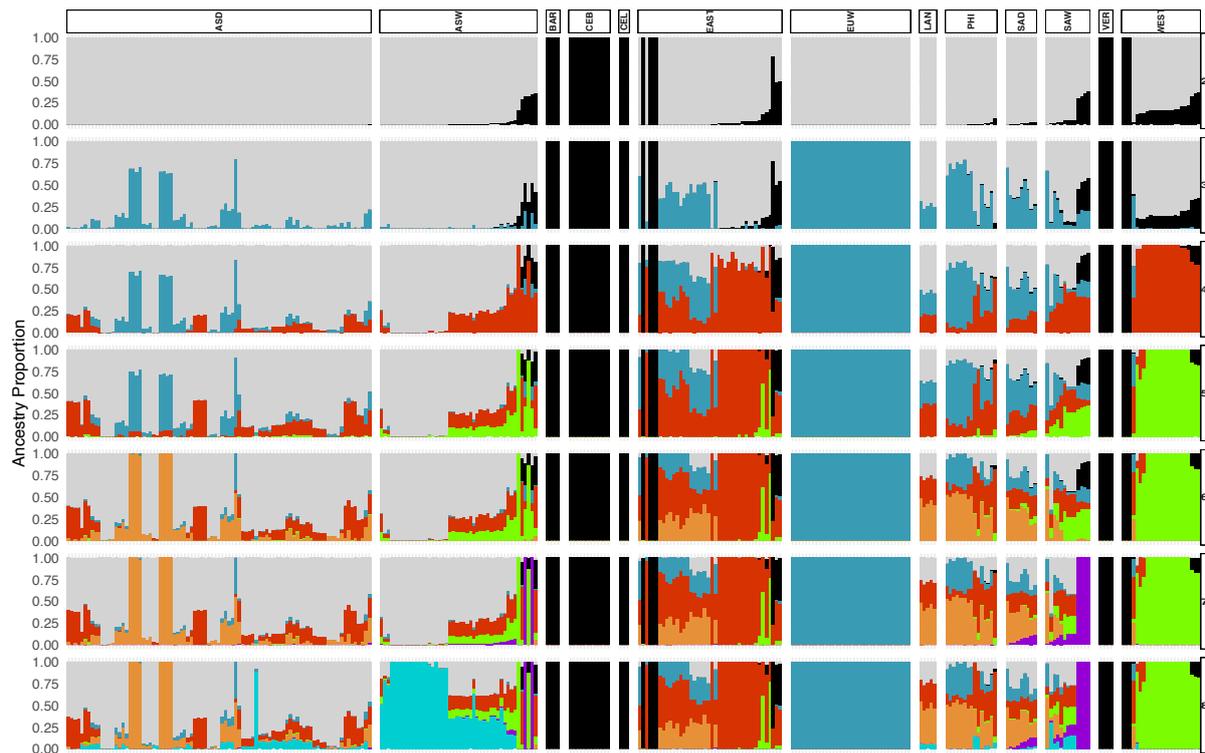


Fig. S7.

Results of unsupervised ADMIXTURE analysis using K=2-8 based on 304 genomes. The following acronym were used to separate individuals into groups: ASD=mainland East Asian domestic, ASW=mainland East Asian , BAR=*S. barbatus*, CEB=*S. cebifrons*, CEL=*S. celebensis*, EAST=ISEA East of the Wallace Line, EUW=European wild, LAN=Lanyu, PHI=domestic pigs from the Philippines, SAD=South Asia domestic (e.g. Bhutan and Nepal), SAW=South Asian wild (e.g. Buthan, India, Nepal), VER=*S. verrucosus*, WEST=ISEA West of the Wallace Line.

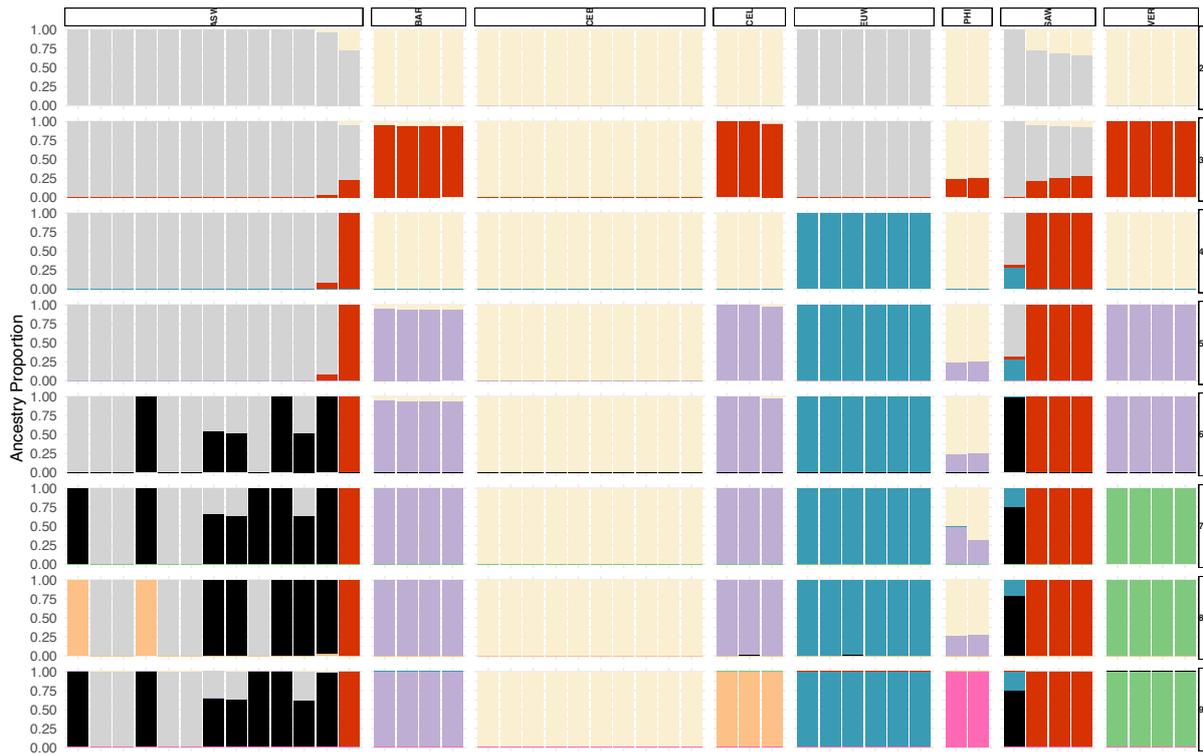


Fig. S8.

Results of unsupervised ADMIXTURE analysis using K=2-8 based on 46 genomes. The following acronym were used to separate individuals into groups: AASW=mainland East Asian , BAR=*S. barbatus*, CEB=*S. cebifrons*, CEL=*S. celebensis*, PHI=*S. Philippensis*, SAW=South Asian wild (e.g. Buthan, India, Nepal), VER=*S. verrucosus*.

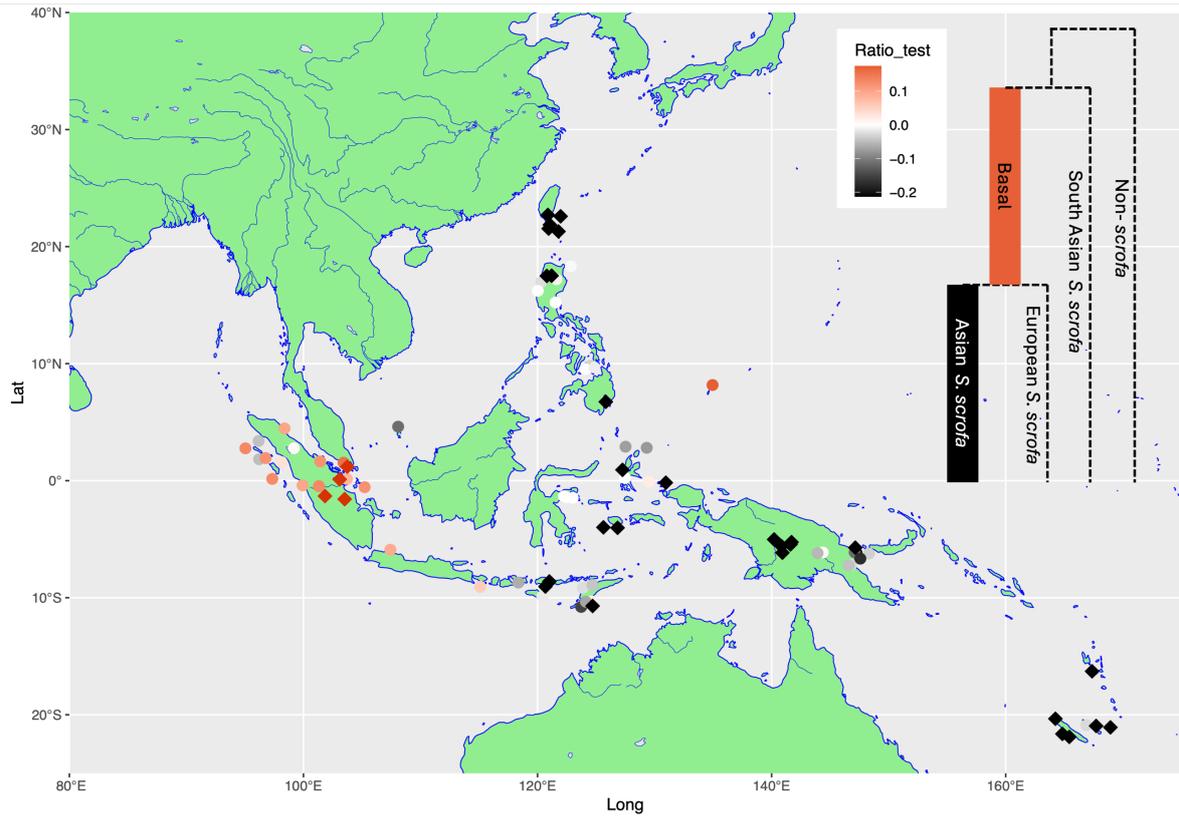


Fig. S9.

Figure showing the fit of admixture graphs assuming basal *S. scrofa* ancestry versus Asian *S. scrofa* ancestry for each sample, accounting for non-*S. scrofa* admixture. Models supporting basal *S. scrofa* ancestry are shown in red, and those supporting Asian ancestry in black. Instances when this support is significant at $p < 0.05$ is shown by diamonds.

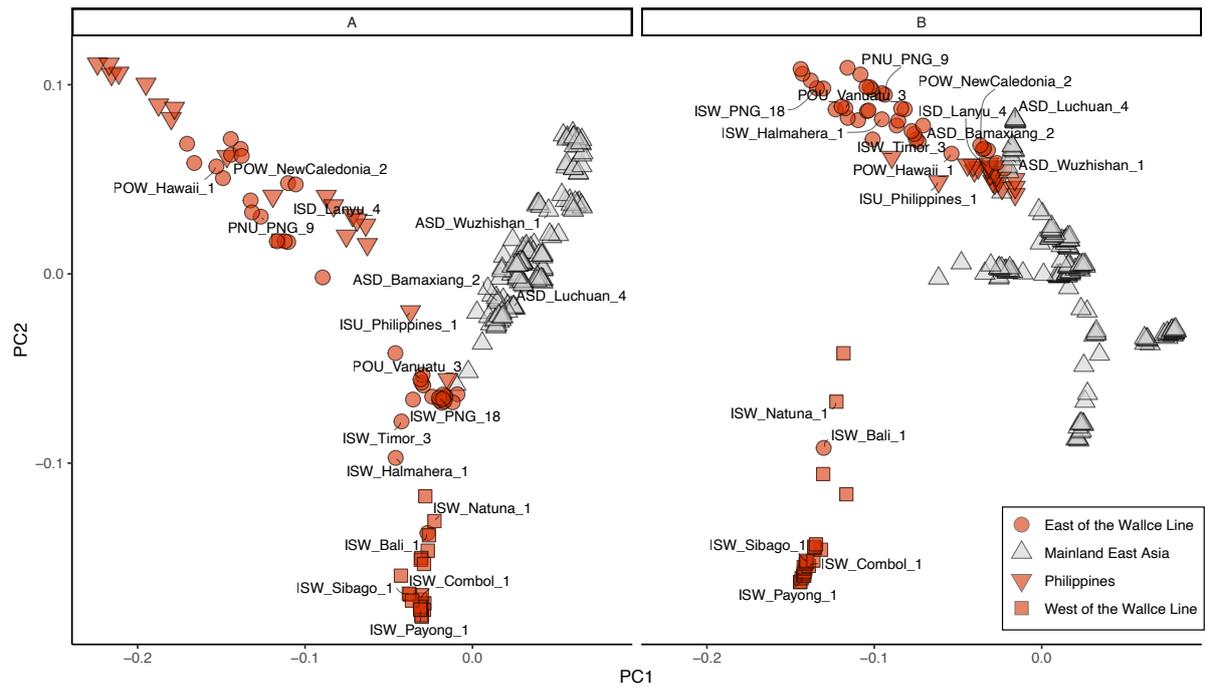


Fig. S10
 Principal component analysis using Asian pigs (both domestic and wild) whole genome (**A**) and deconvoluted genomes (**B**), i.e. without ISEA endemic-*Sus* and European *S. scrofa* ancestry.

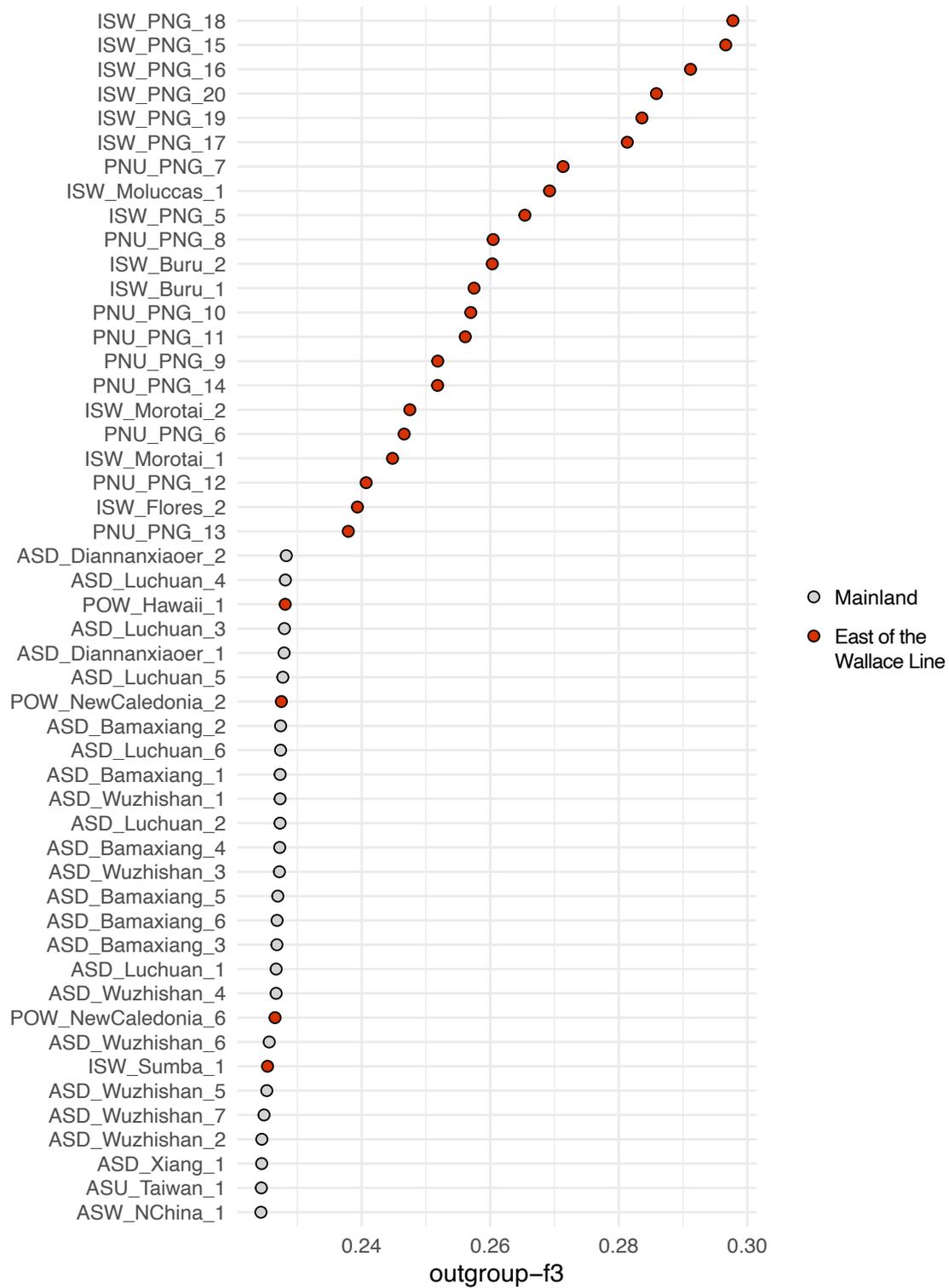


Fig. S11.

The top 50 outgroup-f3 of the form (Pygmy hog [outgroup],X,POU_Vanuatu_3), where X represents any genome in our dataset (y-axis). While this statistic was computed across all individual genomes in our dataset, we have plotted only the top 50 for clarity.

POU_Vanuatu_3 represents a 2,500-year-old ancient Vanuatu genome excavated from a Lapita archaeological complex.

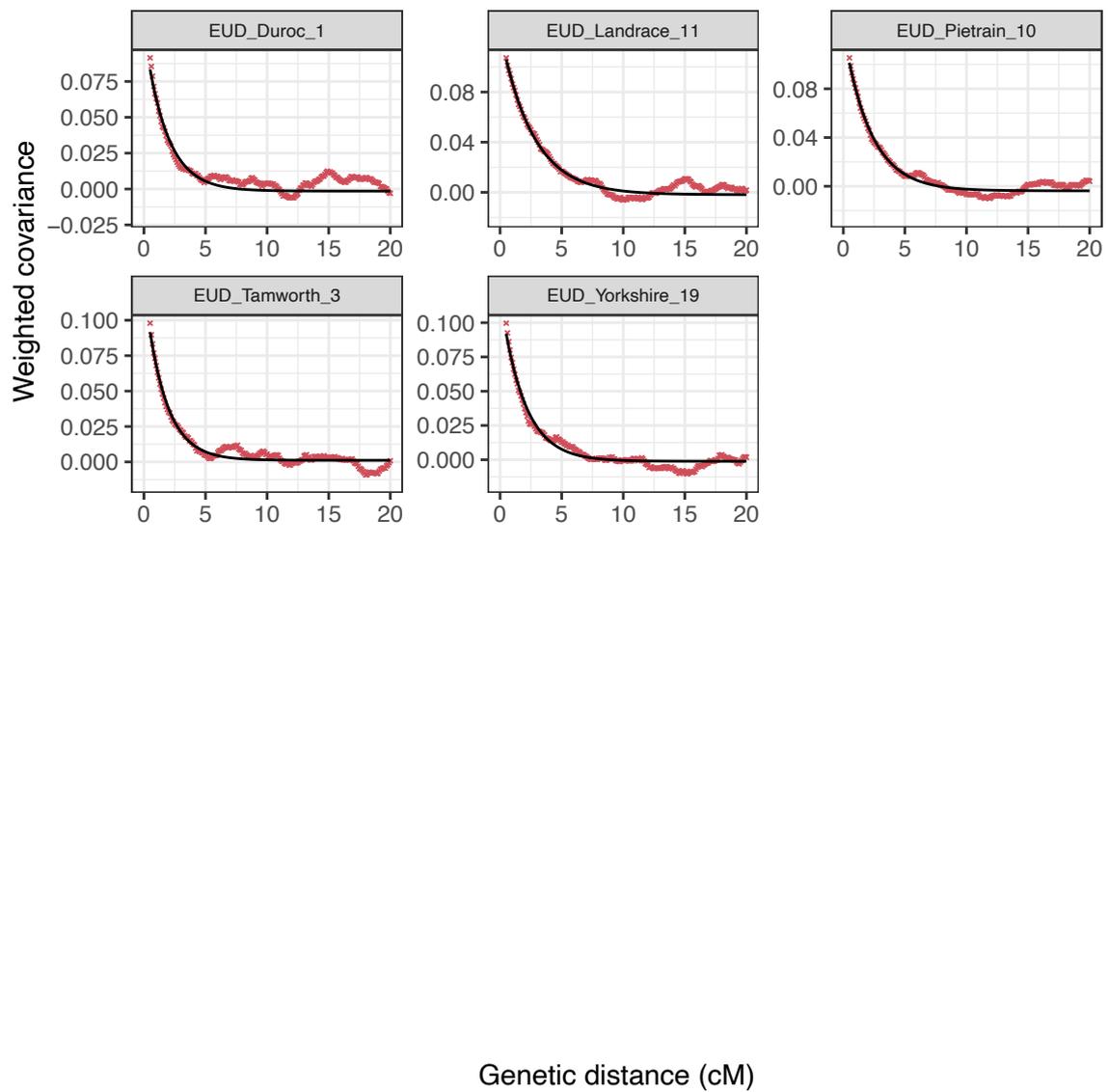


Fig. S12. Example of exponential function fitted by DATES to ancestry covariance in European pigs which are known to possess Asian ancestry.

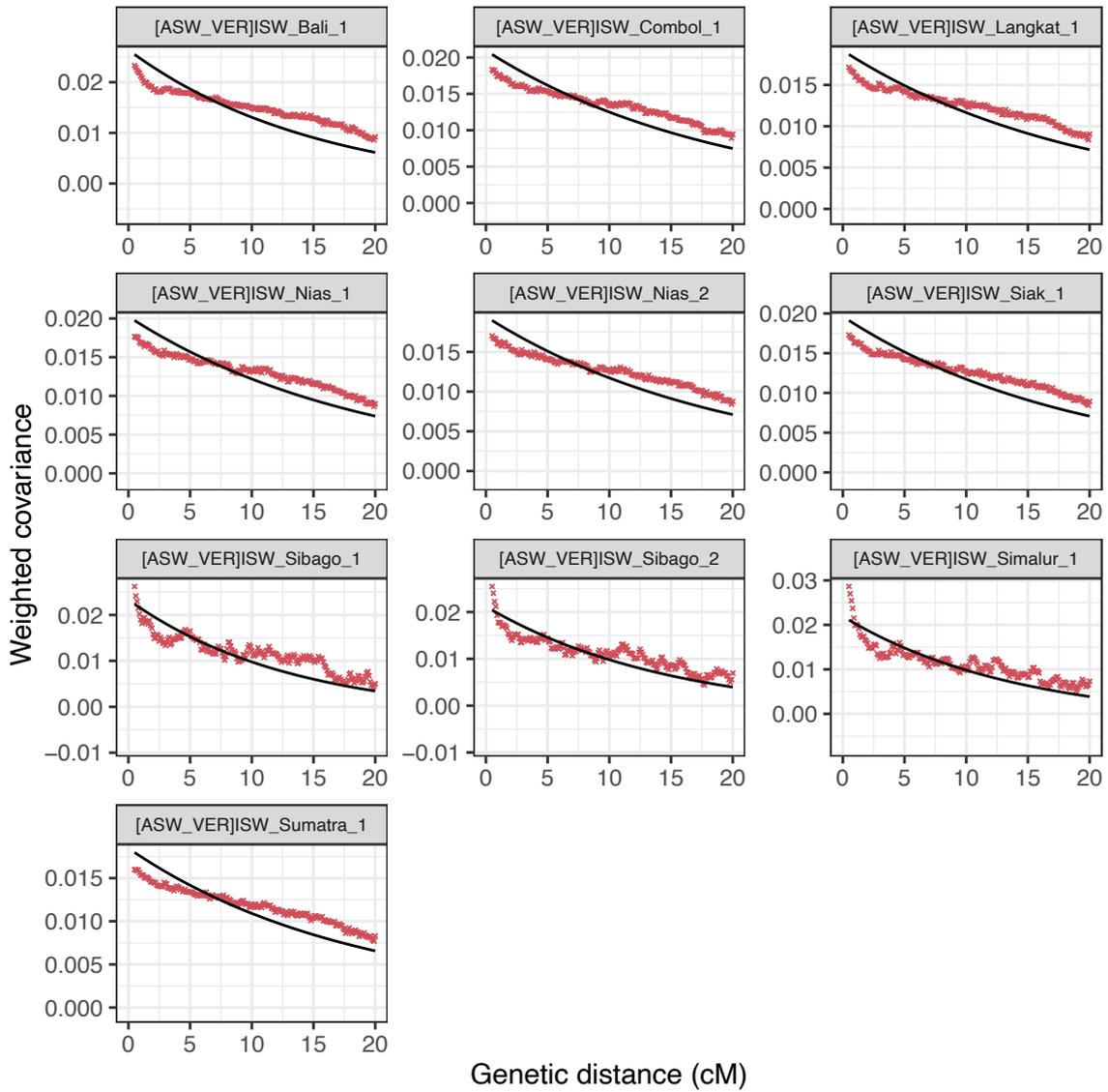


Fig. S13. Exponential function fitted by DATES to ancestry covariance in Asian ancestry which are known to possess *S. verrucosus* ancestry.

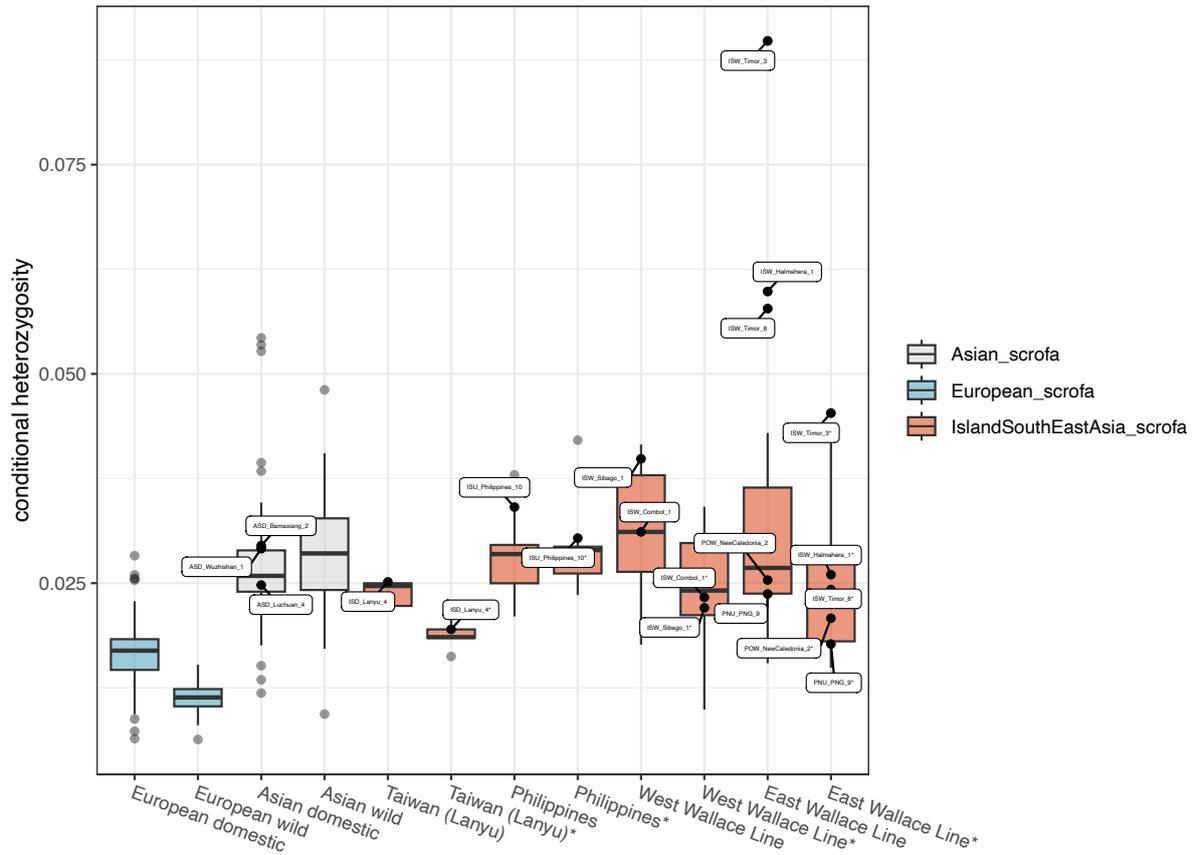


Fig. S14.

(Conditional) heterozygosity estimated by sampling two reads sites found heterozygous in a high coverage *S. cebifrons*. Individuals were grouped based on their location / population. Individuals / populations (on the x-axis) with a * indicate that conditional heterozygosity was computed only at sites that overlap with Asian *S. scrofa* ancestry as per our deconvolution procedure (see e.g. Fig. 3B)

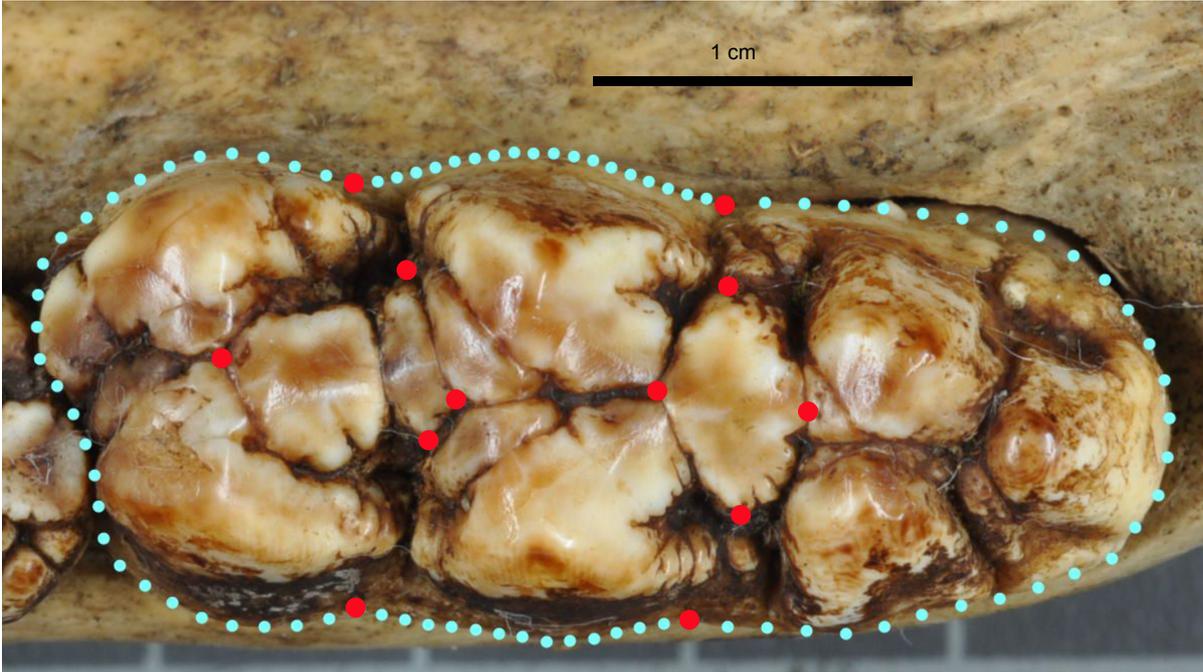
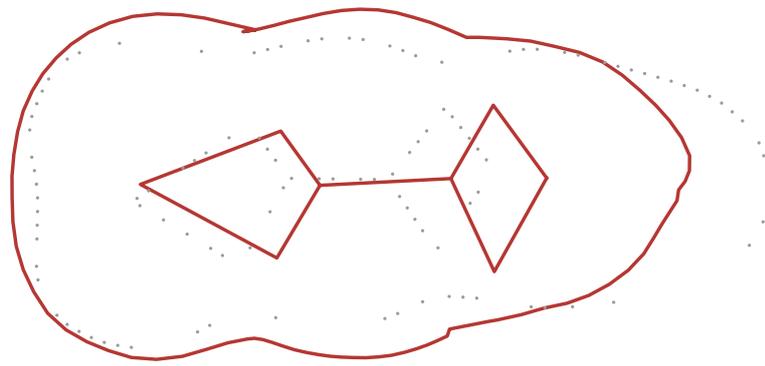


Fig. S15.
Position of the 12 2D-landmarks and 87 sliding semi-landmarks on a third lower molar (specimen nhm-69518).



Ancient Wallacean and Pacific specimens (N=31)
Ancient mainland Asia *Sus scrofa* (N=153)

Fig. S16.

Molar shape differences between Wallacean and Pacific specimens (N=31) and those from mainland Asia (N=147). Visualisation of the third lower molar shape variation along the discriminant axis between the two groups.

Data S1. (separate file)

Information about samples newly sequenced data and publicly available data used in this study including provenance, sampling date, age and sequencing statistics.

Data S2. (separate file)

Information about the mitochondrial data used in Fig. 2A.

Data S3. (separate file)

Metadata and contextual information for the specimens morphometrically analyzed in this study. The table includes the name of the institution hosting the specimen, individual identifier (Individual.ID), taxonomic classification (Genus, species, subspecies), DNA references, and geographical provenance (Country, Site Name, Site Code, Latitude, Longitude). Chronological and cultural attributions are indicated by Chrono, Period, and calibrated dates (Dating BC). Additional contextual data include sex, excavation details (Contact, Feature, Square, Date), inventory numbers (Inventory, Element), notes, the identifications in Fig. 1C and the final morphometric identifications (Supplementary Material).