



Kent Academic Repository

Akdur-Öztürk, Eylem, Al-Adilee, Yaseen Majid, Edwards, William, Gentekaki, Eleni, Tsaousis, Anastasios D. and Dogruman-Al, Funda (2025) *Blastocystis across humans, animals and the environment in rural Türkiye, and relationships with the human intestinal microbiome*. *Frontiers in Microbiology*, 16 .

Downloaded from

<https://kar.kent.ac.uk/111989/> The University of Kent's Academic Repository KAR

The version of record is available from

<https://doi.org/10.3389/fmicb.2025.1665966>

This document version

Publisher pdf

DOI for this version

Licence for this version

CC BY (Attribution)

Additional information

Versions of research works

Versions of Record

If this version is the version of record, it is the same as the published version available on the publisher's web site. Cite as the published version.

Author Accepted Manuscripts

If this document is identified as the Author Accepted Manuscript it is the version after peer review but before type setting, copy editing or publisher branding. Cite as Surname, Initial. (Year) 'Title of article'. To be published in **Title of Journal**, Volume and issue numbers [peer-reviewed accepted version]. Available at: DOI or URL (Accessed: date).

Enquiries

If you have questions about this document contact ResearchSupport@kent.ac.uk. Please include the URL of the record in KAR. If you believe that your, or a third party's rights have been compromised through this document please see our [Take Down policy](https://www.kent.ac.uk/guides/kar-the-kent-academic-repository#policies) (available from <https://www.kent.ac.uk/guides/kar-the-kent-academic-repository#policies>).



OPEN ACCESS

EDITED BY

Lei Deng,
Chinese Academy of Agricultural Sciences,
China

REVIEWED BY

Francesco Asnicar,
University of Trento, Italy
Adedolapo Aminat Rauff-Adedotun,
North-Eastern University, Nigeria

*CORRESPONDENCE

Eleni Gentekaki
✉ gentekaki.e@unic.ac.cy
Anastasios D. Tsaousis
✉ tsaousis.anastasios@gmail.com
Funda Dogruman-Al
✉ alfunda@gazi.edu.tr

RECEIVED 14 July 2025

ACCEPTED 30 September 2025

PUBLISHED 20 October 2025

CITATION

Akdur-Öztürk E, Al-Adilee YMS, Edwards W,
Gentekaki E, Tsaousis AD and
Dogruman-Al F (2025) *Blastocystis* across
humans, animals and the environment in
rural Türkiye, and relationships with the
human intestinal microbiome.
Front. Microbiol. 16:1665966.
doi: 10.3389/fmicb.2025.1665966

COPYRIGHT

© 2025 Akdur-Öztürk, Al-Adilee, Edwards,
Gentekaki, Tsaousis and Dogruman-Al. This is
an open-access article distributed under the
terms of the [Creative Commons Attribution
License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or
reproduction in other forums is permitted,
provided the original author(s) and the
copyright owner(s) are credited and that the
original publication in this journal is cited, in
accordance with accepted academic
practice. No use, distribution or reproduction
is permitted which does not comply with
these terms.

Blastocystis across humans, animals and the environment in rural Türkiye, and relationships with the human intestinal microbiome

Eylem Akdur-Öztürk¹, Yaseen Majid Salman Al-Adilee^{2,3},
William Edwards², Eleni Gentekaki^{4*}, Anastasios D. Tsaousis^{2*}
and Funda Dogruman-Al^{5*}

¹Department of Medical Parasitology, Faculty of Medicine, Çukurova University, Adana, Türkiye, ²School of Natural Sciences, University of Kent, Canterbury, United Kingdom, ³Department of Medical Microbiology, College of Medicine, Ninevah University, Mosul, Iraq, ⁴Department of Veterinary Medicine, University of Nicosia School of Veterinary Medicine, Nicosia, Cyprus, ⁵Division of Parasitology, Department of Microbiology, Gazi University, Ankara, Türkiye

Blastocystis is a globally prevalent intestinal protist commonly found in humans and animals, yet its role in health and disease remains ambiguous. This is a cross-sectional study of *Blastocystis* in rural Türkiye, examining 124 human, 305 livestock (cattle, sheep, goats), and 40 environmental samples using culture/microscopy, qPCR, and sequencing. We further explored associations between *Blastocystis* and population parameters, along with gut microbiota profiles. Using a combination of sequencing and microscopy, the overall prevalence was high, at 76.6% in humans, 71%–78% in livestock, and 38% in environmental samples. Subtypes ST1–ST4 were detected in humans, with ST3 being most frequent. Livestock harbored ST10 predominantly, with goats showing high carriage of ST24. Several subtypes (e.g., ST25, ST26) were recorded in livestock for the first time in Türkiye. Body mass index (BMI) was significantly associated with *Blastocystis* colonization, with lean individuals having higher carriage. Contrary to other studies, individuals with ST4 exhibited reduced bacterial diversity and altered microbial composition, suggesting subtype-specific interactions. By combining parasitology, microbiome, and environmental analysis, this study offers an overview of *Blastocystis* diversity and distribution in rural Türkiye. This work provides a foundation for future integrative research approaches to explore the ecological role of *Blastocystis* and its subtypes, potential health implications, and interactions with other microbes in rural and global contexts.

KEYWORDS

Blastocystis, gut microbiome, transmission dynamics, rural, microbial eukaryotes, Türkiye

Introduction

Blastocystis is a common intestinal protist found in humans and various animals, including mammals, birds, and reptiles. It is one of the two stramenopiles known to inhabit the human gut (Hublin et al., 2021; Nguyen et al., 2023). Despite its widespread presence, its role in health and disease remains unclear, making it a subject of ongoing research (Centers for Disease Control and Prevention [CDC], 2025).

The organism exists in multiple forms—vacuolar, granular, amoeboid, cystic, and less commonly avacuolar and multivacuolar—transmitting through the fecal-oral route via cysts shed in feces (Hublin et al., 2021; Tan, 2008). *Blastocystis* is estimated to colonize nearly one billion people globally (Stensvold and Clark, 2020) with prevalence varying from 5%–20% in developed regions to over 30% in developing areas (Tan, 2008; Khorshidvand et al., 2021). Studies of human populations across diverse geographic regions suggest that *Blastocystis* colonization is associated with distinct gut microbial profiles and higher levels of microbial diversity. It has also been associated with increases in the abundance of beneficial bacterial taxa such as *Ruminococcaceae* and *Prevotella* (Audebert et al., 2016; Beghini et al., 2017; Tito et al., 2019). This is in contrast to the reduced microbial diversity typically observed in individuals with gastrointestinal diseases such as inflammatory bowel disease. Moreover, *Blastocystis* has been reported to be associated with healthy dietary patterns, and lower rates of obesity, cardiometabolic risk, and mortality (Piperni et al., 2024). These findings suggest that *Blastocystis* may be indicative of healthy gut microbiota though the underlying mechanisms of how this might be achieved remain unknown (Audebert et al., 2016; Beghini et al., 2017; Nieves-Ramírez et al., 2018; Kodio et al., 2019; Tito et al., 2019; Alzate et al., 2020; Castañeda et al., 2020; Even et al., 2021).

Based on the diversity of SSU rRNA, at least 44 *Blastocystis* subtypes (STs) have been identified. Among these, 16 STs, including ST1–ST10, ST12, ST14, ST16, ST23, ST35, and ST41, have been identified in humans, with ST1–ST4 being the most frequently reported (McCain et al., 2023; Koehler et al., 2024; Santin et al., 2024). In Türkiye, *Blastocystis* prevalence ranged from 2.1 to 51% across studies, with ST3 (47.9%) as the dominant subtype (Malatyali et al., 2023). Studies on livestock (i.e., cattle, sheep, water buffaloes, and chickens), and companion animals (i.e., dogs, cats, and horses) together with those on environmental sources indicate carriage rates of 3.65% to over 60%, but transmission dynamics remain poorly understood (Onder et al., 2021; Tavour and Önder, 2022).

Recent data suggest that there is interplay of body mass index (BMI) and *Blastocystis* colonization, with several studies reporting higher *Blastocystis* presence in lean individuals (Beghini et al., 2017; Asnicar et al., 2021; Malatyali et al., 2021; Matovelle et al., 2022), while one study found a higher *Blastocystis* prevalence in obese individuals, though the obese sample size was small (Jinatham et al., 2021). Other studies reported prevalence exceeding 40% in obese populations but the absence of lean controls limits interpretation (Caudet et al., 2022a,b).

Zoonotic transmission studies suggest that certain subtypes may spread between humans and animals, but it remains unclear whether these same strains establish colonization or are merely transient (Tsaousis et al., 2025; Abdo et al., 2021; Ruang-areerate

et al., 2021; Rudzińska et al., 2022). Recent studies have reported molecular detection of *Blastocystis* in soil (Jinatham et al., 2021; Blackburn et al., 2024), with certain subtypes shared between human and environmental samples (Jinatham et al., 2021). These findings suggest soil as a transmission route, adding another layer of complexity to its transmission. Hence, an integrative approach that considers humans, animals, and the environment, is essential for shedding light to the organism's epidemiology (Tsaousis et al., 2024). The first study to investigate *Blastocystis* occurrence in this context took place in a rural community in Thailand (Jinatham et al., 2021). Nevertheless studies from diverse regions are essential to assess potential geographic or rural/urban differences. To address this gap, we conducted a study in a rural area of Türkiye. We aimed to explore diversity, distribution, and possible transmission dynamics of *Blastocystis* in humans, livestock (cattle, sheep, goats), and environmental samples from Seyhan Dam Lake in Kırıklı village using microscopy and molecular methods. Moreover, we investigated the relationship between *Blastocystis* colonization and human gut microbiota composition.

Materials and methods

This is a cross-sectional study conducted in Kırıklı village located in the Seyhan Dam Lake basin, Adana, between October and November 2023. Human, animal (cattle, sheep, and goats), and environmental (water and mud) samples were collected and analyzed using microscopy and molecular methods.

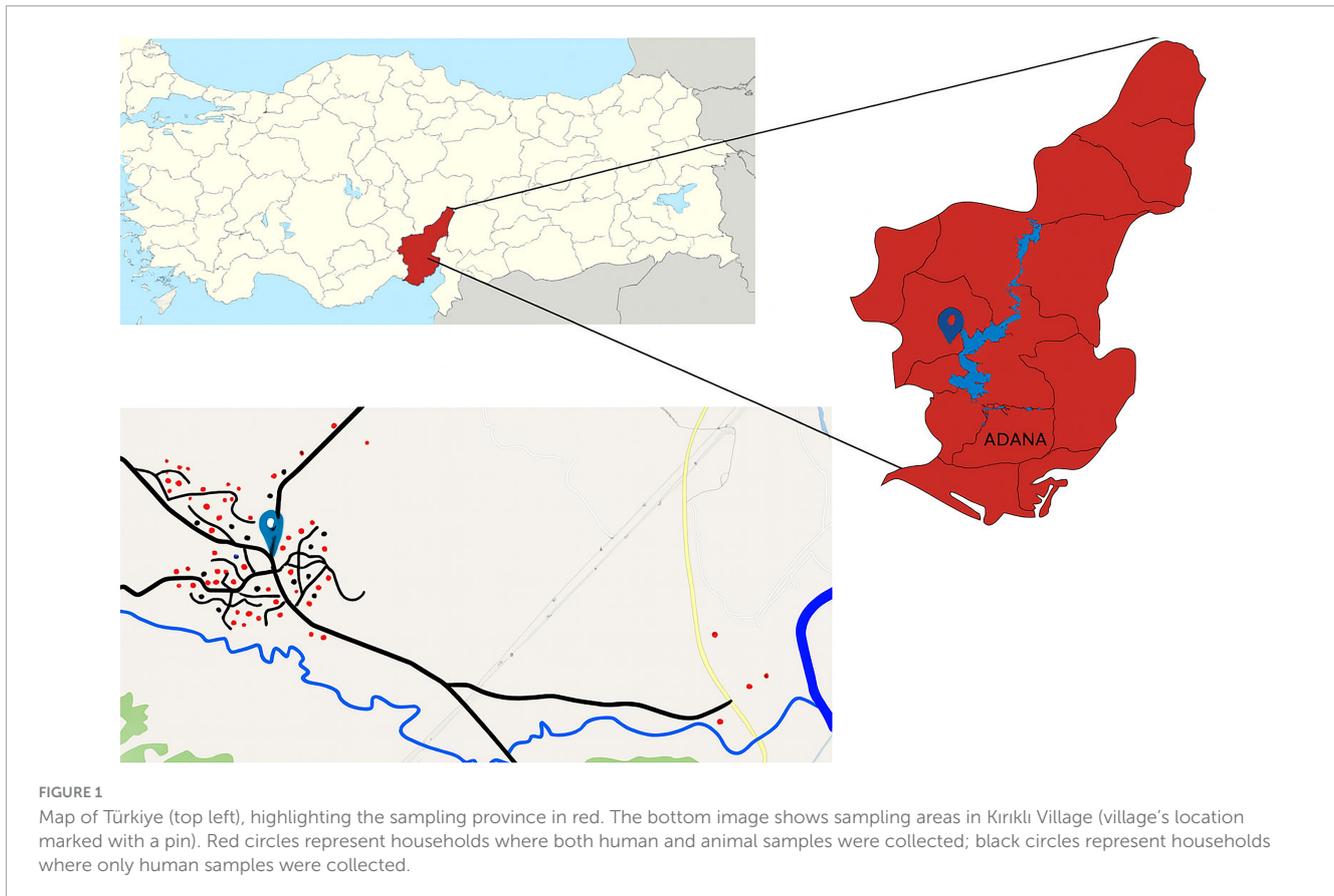
Ethics statement and research permissions

The ethics committee of Çukurova University approved this study for sample collection (approval number 49/135) and microbiome analysis (approval number 39/147). Ethical rules were according to the Declaration of Helsinki. All participants were informed about the nature of the project. Signed consent was obtained from the participants and the parents of the child participants.

For the environmental samples, research permission was obtained from the Adana Governorate and Karaisalı District Directorate of Agriculture and Forestry for the use of dam lake materials (approval number E-12757666-140.03).

Study area

This study was conducted in Kırıklı, a village with a population of 582, located in Karaisalı district of Adana province, Türkiye (37°10'N, 35°14'E), 35 km away from the city center (Figure 1). The village is located within the Mediterranean climate zone, characterized by hot, dry summers and mild, rainy winters. The settlement, whose residents derive their living from farming and animal husbandry, is surrounded by agricultural lands, forest patches, grazing areas and tributaries of the Seyhan Dam Lake. Seasonal animal movements are common in the areas surrounding the village. The Seyhan Dam was constructed around 70 years



ago and is located 12 km from the Kırıklı village (Supplementary Figure 1). Seyhan Dam Lake passes through the border of the village in the study area and is used to irrigate agricultural areas and animals. The water level recedes during November and December. Thus, the area becomes accessible for recreational activities, such as picnic and camping. It is also used as pasture for grazing animals, where they defecate. The area fills with water again in April and May due to snowmelt. This region was selected as a model site for the One Health approach due to its geographical location, climate, proximity to the Seyhan Lake Dam (used for livestock watering and field irrigation) substantial animal populations, rich pastures, and a population primarily engaged in agriculture and animal husbandry. Furthermore, the area has the potential to represent the human-animal-environment cycle of intestinal parasites.

Sample collection

Human, animal (cattle, sheep and goats) and environmental samples were collected and preserved in a 1:2 ratio DNA/RNA Shield™ (ZymoResearch, Freiburg, Germany) until further analysis.

Human fecal samples

The “purposive sampling method,” commonly utilized in biological research and modified accordingly, was used (Işıklar, 2018) to determine the number of human samples for this study. All participants appeared to be healthy, with no cases of

diarrhea or bloody stool. Samples were randomly collected from 124 participants across 64 households. Of these, 90 participants belonged to households actively engaged in animal husbandry, while the other 34 participants were from households without animals and reported not owning any animals for at least 6 months. These two groups were evaluated based on whether they were engaged in animal husbandry or not. There were no cases of diarrhea or bloody stool. Each participant was provided with a labeled, sterile fecal collection container.

Animal fecal samples

A total of 305 fecal samples were collected from 89 cattle, 151 sheep, and 65 goats belonging to the individuals participating in the study. The number of samples for each animal was calculated using the “proportional stratified sampling method” (Işıklar, 2018). Therefore, the number of animals sampled per household varied according to the household’s animal population. To minimize environmental contamination, samples were collected by direct observation of the animals’ defecation during morning feeding, grooming, or rest, so that only a single sample was collected from each animal. Individual samples were collected in pre-labeled containers.

Environmental samples

Twenty-four dam lake water and 16 mud samples were collected directly from the closest parts of the Seyhan Dam lake, where human activities (picnics, camping, fishing, etc.) and animal grazing occurred and from areas irrigated for livestock and agricultural use. Water samples were collected from the dam

lake, its tributaries, and from standing water in 1-liter sterile containers. These were left to stand on a flat surface at room temperature overnight (Jinatham et al., 2021), and the supernatant was drained until 50 ml of sediment was left with a manual drainage system. Then, the sediment was transferred to 15 ml tubes and centrifuged at 500 g for 10 min, the supernatant was drained, and the sediment remaining at the bottom was preserved in a 1:2 ratio DNA/RNA shield until further analysis. Mud samples were taken from the mentioned locations and directly preserved in a 1:2 ratio DNA/RNA shield until further analysis.

Screening of *Blastocystis*

Culture

Approximately 200 mg were taken from all fecal samples and inoculated into 2 ml of Jones' medium containing 10% horse serum (Stensvold et al., 2007; Sarzhanov et al., 2021). Similarly, sediment obtained from water and mud samples processed as described in the Environmental Samples section was also inoculated into the medium. After 48–72 h of incubation at 37 °C, culture samples were examined using a light microscope (Supplementary Figure 2) to determine whether *Blastocystis* was present.

Genomic DNA Extraction

DNA extraction was performed by taking 200 µl of a thoroughly vortexed DNA/RNA shield-sample mixture, and the PureLink™ Microbiome DNA Purification Kit (Thermo Fisher Scientific, Carlsbad, CA, USA) was used according to the manufacturer's protocol.

qPCR (Real-time PCR)

Blastocystis was identified using specific primer sequences (BL18SPPF1_5'-AGTAGTCATACGCTCGTCTCAAA-3' and BL18SR2PP 5'-TCTTCGTTACCCGTTACTGC-3') to amplify a conserved region of SSU rRNA gene (330 bp) by qPCR (Poirier et al., 2011). A negative control (nuclease-free water) and a positive control (genomic DNA of *Blastocystis*) were used in every qPCR run. The reaction mixtures (10 µl) contained 5 µl of Luna Taq Universal (New England Biolabs, Ipswich, MA, USA), 0.5 µl of a 10 µM primer pair, and 2 µl of template DNA. qPCR protocol included; pre-denaturation step: 95 °C for 5 min; followed by 49 cycles of denaturation step: 95 °C for 5 s, annealing step, 68 °C for 10 s, extension step: 72 °C for 15 s and final extension at 72 °C for 10 min. Reactions were set up in 96-well plates in a CFX96 Touch Real-Time PCR Detection System (Bio-Rad, United States).

Polymerase chain reaction and phylogenetic analysis

Nested PCR was performed on samples positive for *Blastocystis* by either culturing (microscopy) or qPCR methods. The primer oligonucleotide sequences of nested PCR are detailed in Supplementary Table 1. First and second-round PCR protocols were set up as follows: pre-denaturation step: 95 °C for 5 min; 30 cycles of: denaturation step: 94 °C for 1 min; annealing step: 59 °C and 50 °C (first and second-round PCR, respectively) for 1 min; extension step: 72 °C for 1 min and a final extension step: 72 °C for 10 min. DNA sequencing was performed by using internal primers for each second-round PCR-positive sample (Cologne, Germany).

The sequences obtained by Sanger sequencing were manually inspected and used as queries to perform BLAST searches for comparisons with reference gene sequences in the National Center for Biotechnology Information (NCBI). For the phylogenetic analysis, a dataset spanning *Blastocystis* diversity, as well as newly derived sequences, was constructed. To avoid redundancy, groups of highly similar sequences (% divergence <98%) were collapsed and only one or two representatives were included in the dataset. Sequences were aligned using MUSCLE v5. Ambiguous positions were removed using Trimal v1.4. Maximum likelihood analysis was performed using IQTREE (Nguyen et al., 2015).

Microbiome sequencing

High-throughput amplicon sequencing was outsourced to Novogene, following a modified version of the protocol described by Caporaso et al., 2011. One nanogram of extracted DNA was used, fragmented, and adapted for paired-end sequencing. The 16S rRNA gene was amplified using the primer pair 515F (GTGCCAGCMGCCGCGGTAA) and 907R (CCGTCAATTCCTTTGAGTTT), which targets the V3-V4 hypervariable region. Sequencing was performed on the Illumina NovaSeq platform.

Raw sequencing reads were processed using the Lotus2 pipeline (Özkurt et al., 2022). The workflow included several key steps: chimera detection and removal were conducted using Minimap2 (Li, 2018), which was also employed to identify and exclude off-target human DNA reads by performing a BLAST search against the Genome Reference Consortium Human Build 38.p14 (0 contaminated samples detected). The trimmed reads were then clustered into Amplicon Sequence Variants (ASVs) with a maximum of one nucleotide difference, using the Divisive Amplicon Denoising Algorithm 2 (DADA2) (Callahan et al., 2016). ASVs were taxonomically classified through BLAST searches against the GreenGenes2 (GG2) database (DeSantis et al., 2006).

Statistical analysis

The data obtained in this study was statistically analyzed using IBM Statistical Package for the Social Sciences (SPSS) version 29 software. A global chi-square test was used to assess whether the distributions of gender and age groups (0–18 years, 19–39 years, 40–59 years, and 60 years and above) differed significantly ($p < 0.05$). For microbiome analysis, the Shapiro-Wilk test was used to assess the normality of data distribution. Normally distributed data were analyzed using ANOVA, followed by the Tukey's HSD test for pairwise comparisons. For non-normally distributed data, the Kruskal-Wallis test was applied, followed by the Dunn test (with Bonferroni adjustment) for multiple comparisons.

Statistical analyses and data visualization were performed using R Studio 4.2.3. To account for variations in sequencing depth, data were first rarefied to 60,000 reads based on the species accumulation curve (Supplementary Figure 1). Rarefaction resulted in the exclusion of one sample. The relative abundance of each genus was then calculated for each sample, and a heatmap was generated to visualize the results. Alpha diversity was assessed using diversity indices, including Shannon and Simpson, and richness estimators including Chao1 and observed taxa, implemented in the Phyloseq

package. These indices were compared between *Blastocystis*-positive and *Blastocystis*-negative samples. To visualize microbiome composition, compositional bar plots were generated using the Microbiome package, including only taxa representing more than 1% of total reads. Principal Coordinate Analysis (PCoA) based on Bray-Curtis dissimilarity was used to visualize differences in microbial community structure between *Blastocystis*-positive and *Blastocystis*-negative samples. Samples were plotted using Bray-Curtis dissimilarity matrices. PERMANOVA (Anderson, 2017) was used to test for statistical significance of group differences. Linear Discriminant Analysis Effect Size (LEfSe) (Segata et al., 2011) was applied to identify potential biomarkers discriminating *Blastocystis*-positive from *Blastocystis*-negative samples based on relative abundance profiles.

Results

Demographic characteristics of the human participants

Of the 124 participants in the study, 58 (46.8%) were female, and 66 (53.2%) were male, with an average age of 44.7 (range between 6 and 82). Of the participants, 90 out of 124 (72.6%) are actively involved in animal husbandry, and 34 (27.4%) stated being involved in caring for any animals for at least the last 6 months. Additionally, all participants declared that they drank tap water. Body mass index (BMI) was calculated for all participants who were 19 years of age or older ($n=109$) (Geifman and Rubin, 2011). Ninety-five (87.2%) of the individuals participating in the study had a BMI above 25 and were classified as overweight or obese. The remaining participants were classified as having normal weight (18.5–24.9). There were no individuals in the study who were extremely underweight (BMI < 18.5). The distribution of sociodemographic characteristics of the participants according to

the presence of *Blastocystis* is shown in Table 1. A more detailed table can be found in Supplementary Table 1.

Blastocystis occurrence

In this study, all sample types were investigated for the presence of *Blastocystis* using culture and qPCR. A sample was considered positive when *Blastocystis* was seen microscopically (Supplementary Figure 2) in culture or when a sequence was obtained (qPCR product or PCR product). Hence, samples positive only for microscopy were included in the prevalence calculation but not subtyping. In all cases, the vacuolar form was predominantly observed microscopically and occasionally granular and amoeboid forms. In human samples, culture positivity was 70.1% (87/124). Regarding qPCR, 111/124 samples yielded a band of the expected size, however of these only 36 samples were successfully sequenced and subtyped. 28 samples were positive by both microscopy and molecular methods. Combining the two approaches, the overall positivity rate in humans in this study was 76.6% (95/124). Positivity in the 0–18 age group was 66.7% (10/15), 80.8% (21/26) in the 19–39 group, 76.3% (45/59) in the 40–59 group and 75% (18/24) in the over 60 group. Statistical analysis showed a significant association between BMI and *Blastocystis* positivity ($p < 0.05$). Though *Blastocystis* was more prevalent among individuals with normal BMI no statistical test was performed at the subgroup level. Global chi square tests revealed no statistically significant relationships between *Blastocystis* positivity and other demographic variables such as gender, occupation, and age groups ($p > 0.05$).

In animals, 66% (200/304) were culture positive. More specifically, 65.2% (58/89) of cattle were culture positive, 66.2% (100/151) of sheep, and 64.6% (42/65) of goats. Using qPCR, 100% of the samples showed a band of the expected size. Of these, 77 were successfully sequenced and subtyped (24 cattle, 35 sheep, and 17 goats). Combining the two approaches, the positivity rate was as 71% (63/89) in cattle, 73% (110/151) in sheep, and 78% (50/64) in goats.

The environmental samples showed 37.5% (15/40) positivity solely through culture. Eleven of these were water samples, while four were mud. None of the environmental samples yielded *Blastocystis* sequences.

Blastocystis subtypes

In total, 112 sequences were subtyped from humans ($n = 36$), cattle ($n = 24$), sheep ($n = 35$) and goat ($n = 17$). In multiple instances the chromatograms showed evidence of mixed infections, as indicated by multiple peaks. To assign subtype the criteria below were followed: reasonably good quality sequence of over 300 bp and over 98% similarity in GenBank. Subtypes were identified using a combination of phylogeny and blast against GenBank and the curated database pubmlst¹ and phylogeny (Supplementary Figure 3). In the phylogenetic analysis all subtypes were monophyletic and the new sequences placed within known subtype clades. For 18 sequences, the methodologies did not agree. In pubmlst, they were identified as belonging to one of the subtypes previously comprising ST10 (i.e., ST23, ST42-ST44), perhaps due to the short length of the sequences. In the phylogenetic tree, these

TABLE 1 Distribution of sociodemographic characteristics of participants according to the presence of *Blastocystis*.

Characteristics		Positive (n%)	Negative (n%)	Total (%)	p value
Gender	Female	40 (69.0%)	18 (31.0%)	58 (46.8%)	$p \approx 0.095$ $\chi^2 \approx 2.7842$
	Male	54 (81.8%)	12 (18.2%)	66 (53.2%)	
Age group	0–18	10 (66.7%)	5 (33.3%)	15 (12.1%)	$p \approx 0.79$ $\chi^2 = 1.048$
	19–39	21 (80.8%)	5 (19.2%)	26 (21.0%)	
	40–59	45 (76.3%)	14 (23.7%)	59 (47.6%)	
	60 and above	18 (75.0%)	6 (25.0%)	24 (19.4%)	
BMI*	18.5–24.9	14 (100.0%)	0 (0.0%)	14 (12.8%)	$p \approx 0.032$ $\chi^2 = 6.892$
	25.0–29.9	28 (66.7%)	14 (33.3%)	42 (38.5%)	
	≥30.0	42 (79.2%)	11 (20.8%)	53 (48.6%)	
Animal husbandry	Yes	65 (72.2%)	25 (27.8%)	90 (72.6%)	$p \approx 0.129$ $\chi^2 = 2.297$
	No	29 (85.3%)	5 (14.7%)	34 (27.4%)	

*Body mass index (BMI) is calculated for ages 19 and above. Bold values indicate statistical significance ($p < 0.05$).

1 https://pubmlst.org/bigbdb?db=pubmlst_blastocystis_seqdef&page=sequenceQuery

TABLE 2 Distribution of *Blastocystis* subtypes among human and animal hosts.

Host	ST1 n (%)	ST2 n (%)	ST3 n (%)	ST4 n (%)	ST5 n (%)	ST10 n (%)	ST21 n (%)	ST24 n (%)	ST25 n (%)	ST26 n (%)	ST42 n (%)	ST43 n (%)	ST44 n (%)	Total n
Human	11 31%	11 31%	12 33%	2 6%										36
Cattle						8 33.3%			2 8.3%	3 12.5%	5 20.8%	1 4.1%	5 20.8%	24
Sheep				1 2.9%	2 5.7%	17 48.6%		5 14.3%		4 11.4%		3 8.6%	3 8.6%	35
Goat						5 29.4%	2 11.8%	7 41.2%		1 5.9%		1 5.9%	1 5.9%	17
Total	11	11	12	3	2	30	2	12	2	8	5	5	9	112

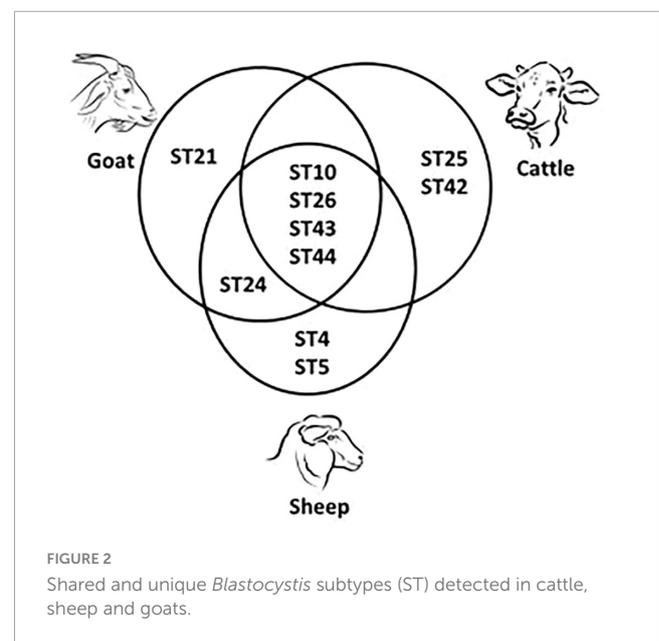
placed within the clade comprising ST10 in the past, but jumped within the clade depending on taxon sampling. Of these, 13 were from cattle, three from sheep and two from goats. These sequences were not assigned a subtype, only the clade with which they cluster.

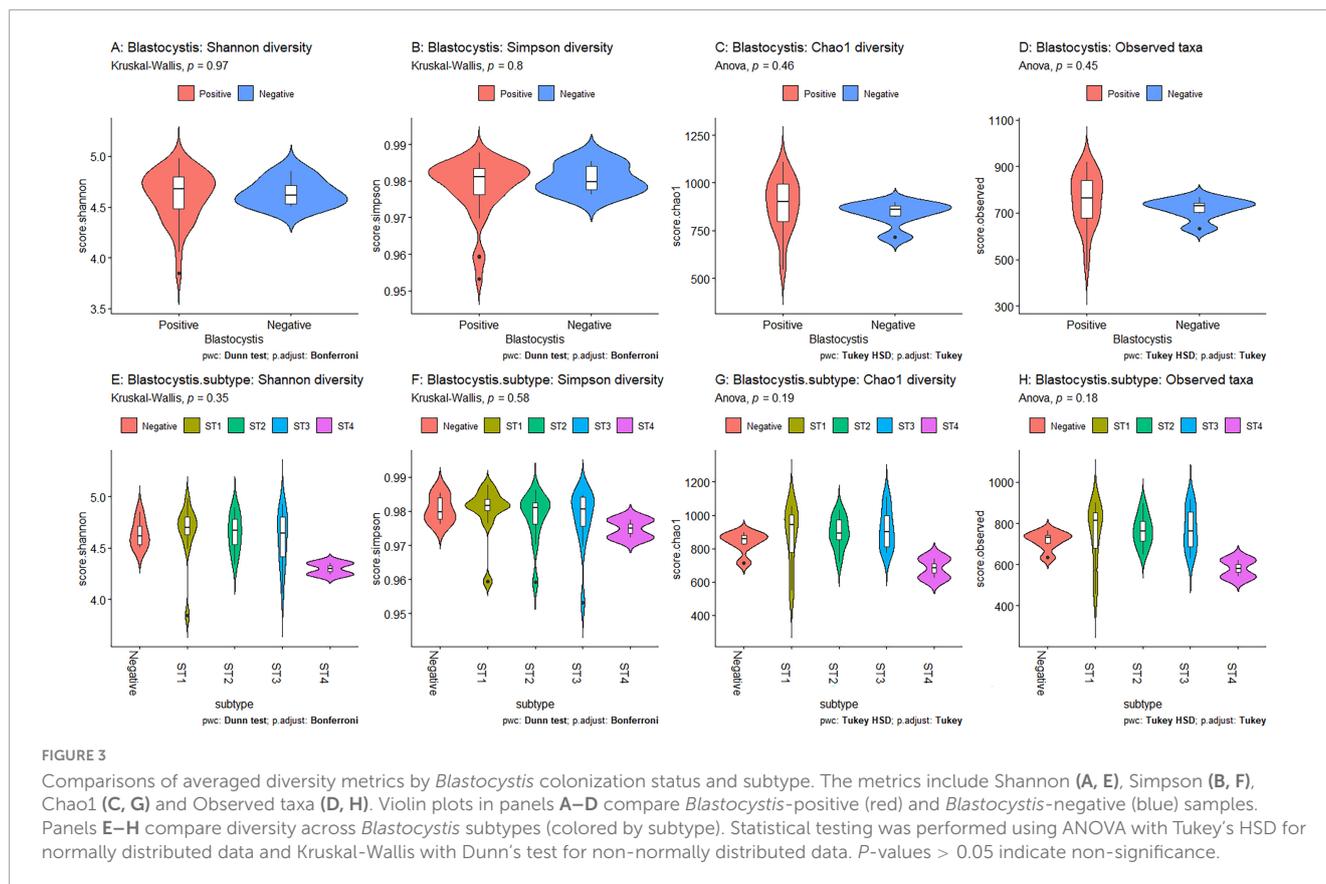
In total, 13 subtypes were identified: ST1, ST2, ST3, ST4, ST5, ST10, ST21, ST24, ST25, ST26, ST42, ST43, and ST44. In humans, four subtypes were detected, namely ST1, ST2, ST3, and ST4. The most abundant subtype was ST3 (33%, 12/36), followed by ST1, and ST2 (each at 31%, 11/36) and ST4 (6%, 2/36). In animals, ten subtypes were detected: ST4, ST5, ST10, ST21, ST24, ST25, ST26, ST42, ST43, and ST44. In cattle, ST10 ($n = 8$), ST25 ($n = 2$), ST26 ($n = 3$), ST42 ($n = 5$), ST43 ($n = 1$), and ST44 ($n = 5$) were identified. In sheep, ST4 ($n = 1$), ST5 ($n = 2$), ST10 ($n = 17$), ST24 ($n = 5$), ST26 ($n = 4$), ST43 ($n = 3$), and ST44 ($n = 3$) were detected. Goat samples were positive for ST10 ($n = 5$), ST21 ($n = 2$), ST24 ($n = 7$), ST26 ($n = 1$), ST43 ($n = 1$), and ST44 ($n = 1$). Table 2 showed the distribution of *Blastocystis* subtypes among human and animal host. ST10 was the most abundant in cattle and sheep, while in goats ST24 was. ST10, ST26, ST43, and ST44 were shared by all three ruminant species (Figure 2). ST25 and ST42 were detected only in cattle, while ST21 was detected only in goats. ST24 was detected in goats and sheep, but not in cattle.

At the household level, subtype analysis was performed on more than one individual from five households, and no common subtype (ST) was identified among individuals within the same household. However, common subtypes were reported among livestock within the same household.

Microbiome analysis

Using Shannon (Figure 3A), Simpson (Figure 3B), Chao1 (Figure 3C) and observed taxa (Figure 3D), there were no significant differences in bacterial alpha diversity between *Blastocystis* positive and *Blastocystis* negative samples, as confirmed by ANOVA/Kruskal-Wallis tests. The same metrics were then



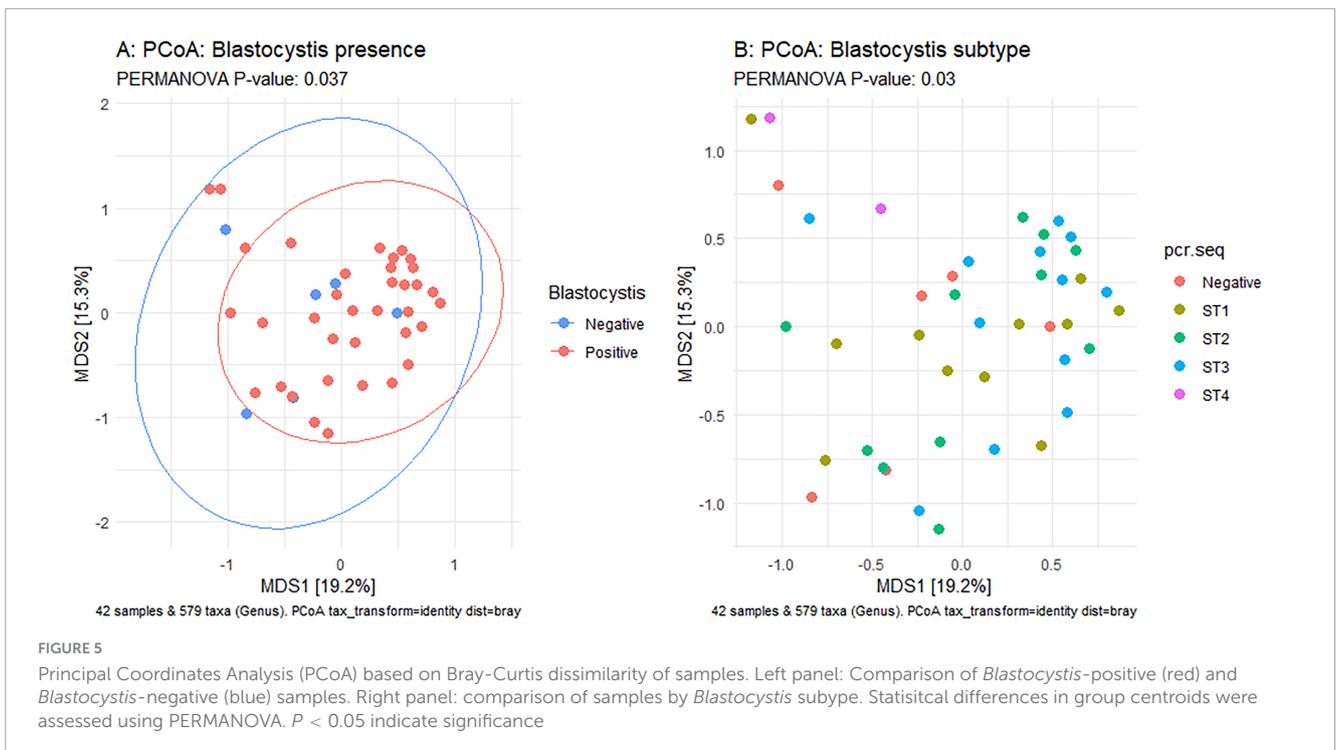
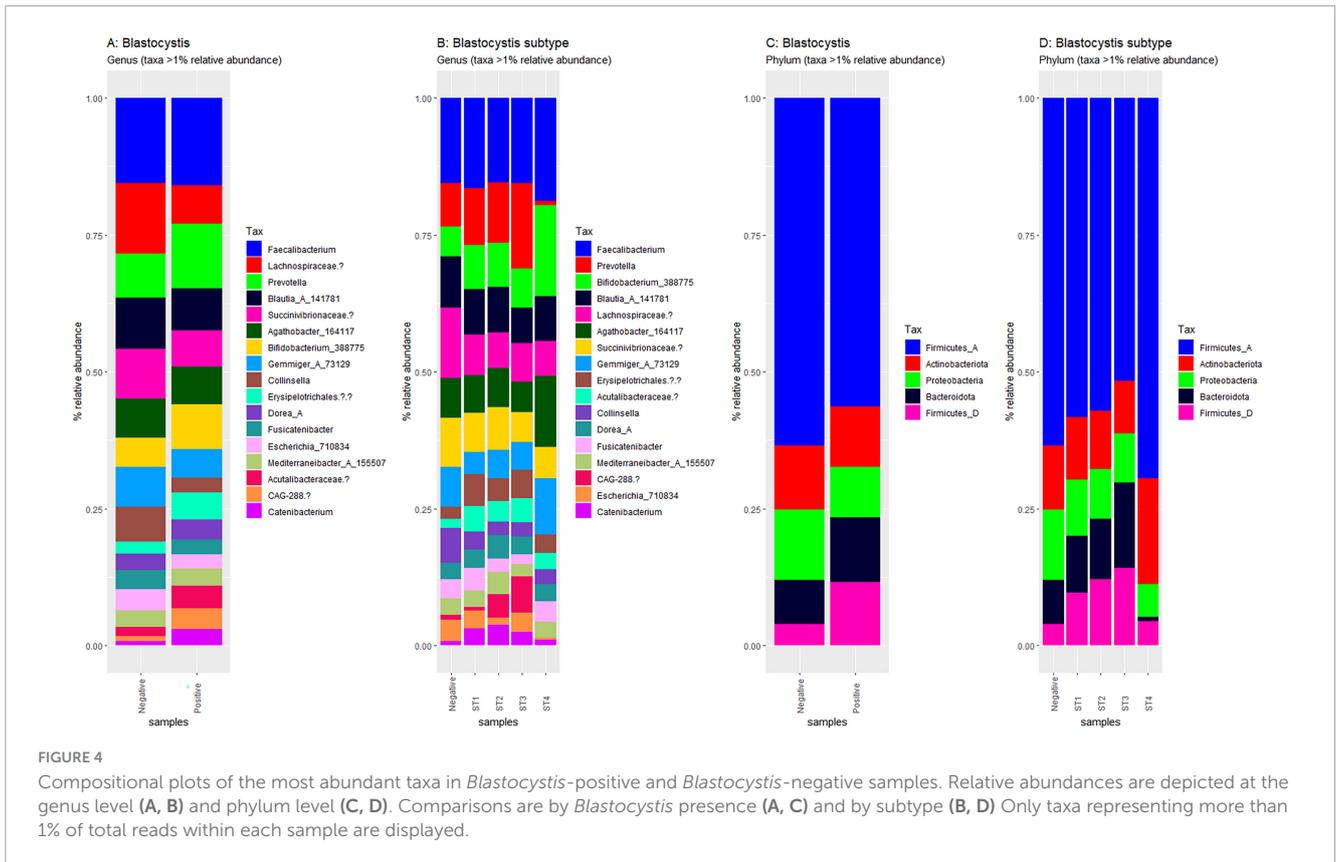


compared at the subtype level (ST1–ST4) (Figure 3E–H). ST1 had the highest average diversity score average across all metrics relative to the *Blastocystis* negative samples, while ST4 samples showed a trend toward lower diversity and richness, but these results were not significant. Comparison of the composition of samples grouped by *Blastocystis* presence is shown in Figure 4 (individual samples are shown in Supplementary Figure 2). At the genus level, *Blastocystis* positive samples had a higher relative abundance of *Prevotella* and *Bifidobacterium* and relatively decreased *Lachnospiraceae* (Figure 4A). When samples were grouped by subtype, ST4 had a notable reduction in *Prevotella* and a marked increase in *Bifidobacterium* along with an increase in *Agathobacter* (Figure 4B). At the phylum level, *Blastocystis* positive samples showed an increase in *Bacteroidota* and a decrease in *Proteobacteria* (Figure 4C). When the samples were grouped by *Blastocystis* subtype, ST4 showed markedly lower relative abundance of *Bacteroidota*, and an increase in *Actinobacteriota* (Figure 4D). To assess differences in overall bacterial community composition principal coordinate analysis (PCoA) was used based on Bray-Curtis dissimilarities (Figure 5). PERMANOVA testing indicated that the community structure differed significantly between *Blastocystis* positive and *Blastocystis* negative groups, although the effect size was small ($p = 0.037$, $R^2 = 0.046$), (Figure 5A). When samples were grouped by *Blastocystis* subtype (Figure 5B), PERMANOVA testing also revealed significant differences in community composition ($p = 0.03$, $R^2 = 0.136$), indicating a stronger effect size compared to colonization status alone. Pairwise Adonis was used to assess differences in community composition between groups. A significant difference was observed

between ST3 and *Blastocystis* negative samples (Supplementary Table 2). To identify taxa associated with *Blastocystis* colonization status, Linear discriminant analysis Effect Size (LEfSE) was used. Linear discriminant analysis (LDA) scores with an absolute value of 2 were considered indicative of discriminative features. Taxa were aggregated to the genus and family levels (Figure 6). Those with high LDA scores contributed most strongly to distinguishing *Blastocystis* positive from *Blastocystis* negative samples. In total, 38 genera were identified as discriminatory for *Blastocystis* negative samples, while 13 genera were discriminatory for *Blastocystis* positive samples. At the family level, these numbers were 5 and 13, respectively.

Discussion

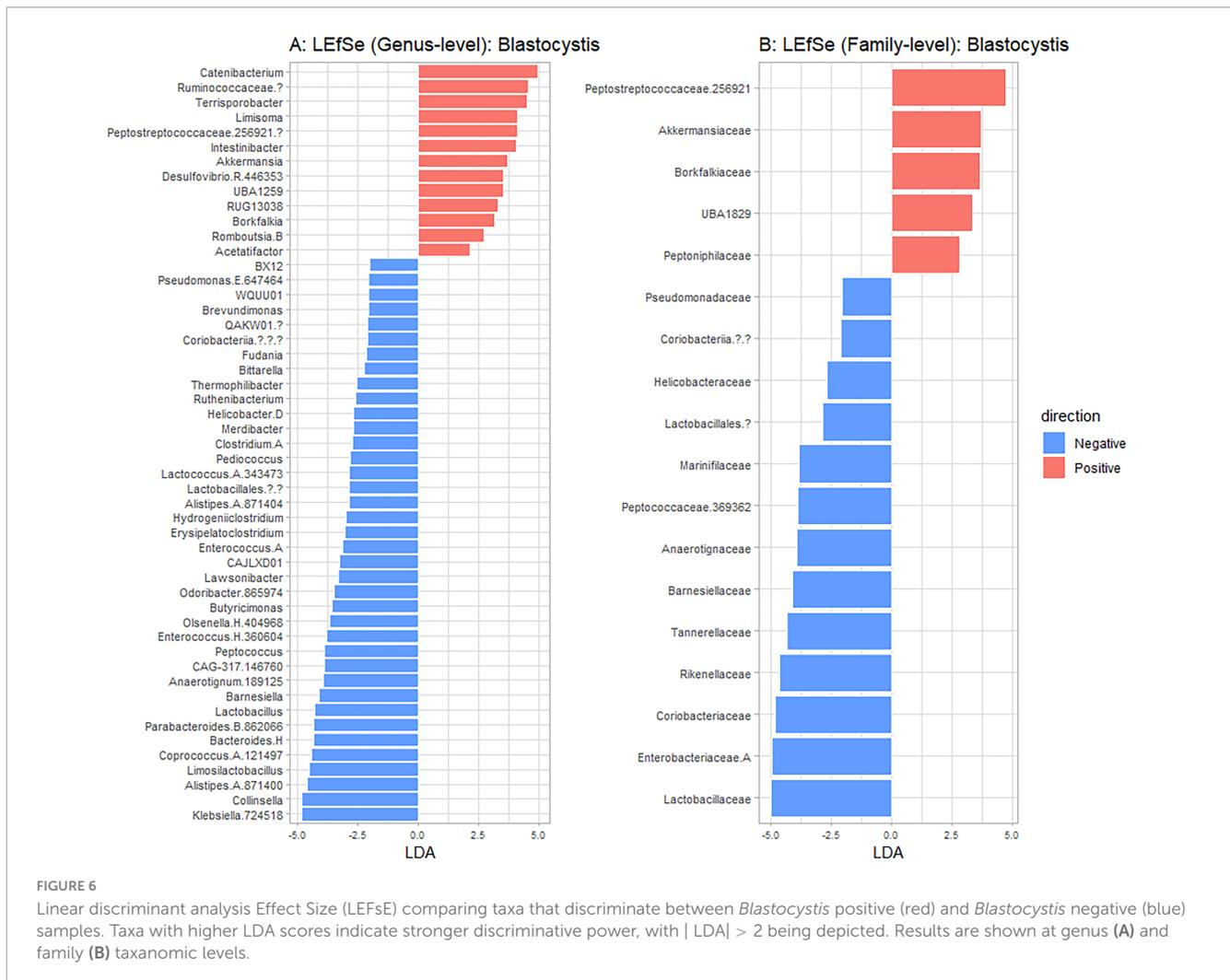
The current study presents the first integrative investigation of *Blastocystis* in rural Türkiye, examining human, livestock, and environmental samples to assess its prevalence, subtype distribution, and gut microbiome composition. Anthropometric data from human participants revealed a significant association between BMI and *Blastocystis* colonization in agreement with previous studies (Beghini et al., 2017; Asnicar et al., 2021; Malatyali et al., 2021; Matovelle et al., 2022), with lean individuals having the highest *Blastocystis* positivity rate. A few studies have found a high occurrence of *Blastocystis* in obese individuals (Jinatham et al., 2021; Caudet et al., 2022a,b). Nonetheless, the organism’s presence has been associated with bacterial taxa linked to improved cardiometabolic health profiles (Piperni et al., 2024) and in the



obese with lower incidence of metabolic syndrome (Caudet et al., 2022a,b). Moreover, *Blastocystis* colonization was not associated with gut inflammation as indicated by lower fecal calprotectin levels (Nieves-Ramírez et al., 2018). Collectively, these findings support hypotheses of a beneficial role for *Blastocystis* within the

gut ecosystem and warrants further exploration to understand its contributions and underlying mechanisms (Deng and Tan, 2025; Stensvold, 2025).

Our results indicate a high occurrence of *Blastocystis* across all studied host species, consistent with findings from other



rural communities (El Safadi et al., 2014; Jinatham et al., 2021). However, the true incidence is likely underestimated as several samples negative by microscopy were clearly *Blastocystis* positive, but had to be excluded due to poor sequencing quality. Moreover, several samples positive only by microscopy also failed amplification or sequencing. This was notably the case with the animal and environmental samples. A main issue is the lack of standardization of *Blastocystis* qPCR from environmental samples. Alternative explanations that could account for this, include potential amplification inhibition and mixed infections. Optimizing environmental DNA methodologies are essential and could significantly improve our understanding of ecological reservoirs and transmission pathways.

Overall, 13 subtypes were identified indicating high genetic diversity in the sampled area and associated hosts. Humans predominantly carried subtypes ST1, ST2, and ST3, aligning with global human-associated subtype distributions (Alfellani et al., 2013; Jiménez et al., 2022). Livestock, all of which were ruminants (cattle, sheep, and goats), carried subtypes typical of these animals (Maloney et al., 2019; Rauff-Adedotun et al., 2023; Stensvold et al., 2023; Heydarian et al., 2024; Naguib et al., 2024; Santin et al., 2024). When looking at the community and household level there was barely any sharing of subtypes between humans and their

animals. This finding does not seem to match several other studies, which provide evidence that supports the zoonotic transmission of *Blastocystis*. Notably, however, the overall proportion of confirmed zoonotic cases remains low; in nearly all studies, only a small subset of humans shared identical subtypes with animals (Yoshikawa et al., 2009; Rudzińska et al., 2022; Salehi et al., 2022; McCain et al., 2023; Šejnohová et al., 2024). Hence, even though zoonotic transmission is possible, it appears to be infrequent in the studied populations. An alternative explanation might be environmental exposure. *Blastocystis* was previously detected in environmental samples, and recent studies focusing on subtyping have revealed a wide range of subtypes in such samples (Noradilah et al., 2016; Jinatham et al., 2021; Sanhueza Teneo et al., 2025). Herein, we detected *Blastocystis* in water and/or soil samples; however, sequencing failure prevented subtype identification. Collectively, these findings point toward the environment having a role in driving *Blastocystis* transmission. Nonetheless, this route remains comparatively less studied and warrants further investigation.

Blastocystis colonization has been associated with distinct gut microbial profiles characterized by increased microbial diversity and bacterial species richness (Audebert et al., 2016; Nieves-Ramírez et al., 2018; Kodio et al., 2019; Even et al., 2021; Piperni et al., 2024; Castañeda et al., 2025). Most of these studies

have considered the presence or absence of *Blastocystis*, with comparatively fewer exploring differences across subtypes (Beghini et al., 2017; Tito et al., 2019; Antonetti et al., 2024). Our analyses reveal subtypes specific structuring, indicating that individuals harboring different subtypes have distinct microbial communities consistent with previous findings. In this study, ST4 stood out as it showed a trend toward lower microbial diversity and richness in contrast to earlier studies (Beghini et al., 2017; Tito et al., 2019). Notably, in these studies, nearly all ST4-positive samples (except for two from Asia) originated from westernized populations, mainly from Europe. This suggests that ST4-microbiota associations may vary across populations or environments. Alternatively, the difference could be due to the small number of individuals with ST4 herein. Regardless, this finding highlights the need to investigate diverse populations focusing on *Blastocystis* subtype-level differences.

This study has certain limitations. First, the cross-sectional design precludes assessment of temporal relationships (Tsaousis et al., 2025). While a significant association between BMI and *Blastocystis* carriage was revealed, the test did not establish whether differences among the three BMI groups underlie that finding as other potential confounding factors (e.g., diet, co-existing diseases) were not considered.

Conclusion

This study not only bridges a knowledge gap by characterizing *Blastocystis* subtype diversity in humans and livestock within a rural Turkish setting but also spearheads a model for integrated One Health investigations in the region. By combining classical parasitological, and microbiome analyses, we demonstrate the feasibility and value of this type of framework. These findings lay the groundwork for future longitudinal and comparative research both within Türkiye and in similar rural communities globally. Additional multidisciplinary collaborative studies will be instrumental in redefining our understanding of intestinal protists—not solely as pathogens, but as complex microbial players in host health and environmental ecosystems (Tsaousis et al., 2024).

Data availability statement

The *Blastocystis* sequences presented in this study can be found in NCBI under the accession numbers PX444604-PX444713. The raw microbiome data presented in this study can be found under the accession number PRJNA1291457 in the same database.

Ethics statement

The studies involving humans were approved by Research Ethics Committee of Çukurova University Faculty of Medicine, Adana, Türkiye. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

EA-Ö: Conceptualization, Formal analysis, Funding acquisition, Investigation, Writing – original draft. YA-A: Data curation, Investigation, Methodology, Writing – original draft. WE: Data curation, Formal analysis, Methodology, Writing – original draft. EG: Investigation, Supervision, Validation, Writing – review & editing. AT: Conceptualization, Data curation, Funding acquisition, Project administration, Resources, Supervision, Validation, Visualization, Writing – review & editing. FD-A: Conceptualization, Funding acquisition, Project administration, Resources, Visualization, Writing – original draft.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This study was supported within the scope of the Scientific and Technological Research Council of Türkiye (TÜBİTAK) 1002 Quick Support Program (project no: 123S965), CA21105 - *Blastocystis* under One Health (OneHealthBlastocystis) and South Coast Biosciences Doctoral Training Partnership SoCoBio DTP BBSRC BB/T008768/1 to W.J.S.E.

Acknowledgments

We would like to thank the village veterinarians Nadir Öztürk and Hacer Güneş for their help in collecting animal materials, the village doctor Mustafa Yaşar Yılmaz and the village nurse Raşide Çam for their help in collecting human materials, and all the Kırıklı villagers for their support.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The handling editor LD declared a past co-authorship with the author AT.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmicb.2025.1665966/full#supplementary-material>

SUPPLEMENTARY FIGURE 1

Field photographs of the sampling area.

SUPPLEMENTARY FIGURE 2

Blastocystis cells observed in culture.

SUPPLEMENTARY FIGURE 3

Maximum likelihood phylogeny of *Blastocystis* sequences.

SUPPLEMENTARY FIGURE 4

Rarefaction curves of 16S sequencing samples. The total number of sequencing reads is shown on the X-axis, and the taxa detected at the corresponding sequencing are displayed on the Y-axis.

SUPPLEMENTARY FIGURE 5

Compositional bar plots showing the most abundant taxa in *Blastocystis*-positive and negative samples. Each bar represents an individual sample. Taxa were aggregated to Genus (A,B) and phylum (C,D) level. Samples have been grouped by *Blastocystis* colonization status (A,C) or *Blastocystis* subtype (B,D). Only taxa representing more than 1% of the total reads within each sample are displayed.

References

- Abdo, S. M., El-Adawy, H., Farag, H. F., El-Taweel, H. A., Elhadad, H., and El-Badry, A. A. M. (2021). Detection and molecular identification of *Blastocystis* isolates from humans and cattle in northern Egypt. *J. Parasitic Dis.* 45, 738–745. doi: 10.1007/s12639-021-01354-5
- Alfellani, M. A., Taner-Mulla, D., Jacob, A. S., Imeede, C. A., Yoshikawa, H., Stensvold, C. R., et al. (2013). Genetic diversity of *Blastocystis* in livestock and zoo animals. *Protist* 164, 497–509. doi: 10.1016/j.protis.2013.05.003
- Alzate, J. F., Toro-Londoño, M., Cabarcas, F., Garcia-Montoya, G., and Galvan-Diaz, A. (2020). Contrasting microbiota profiles observed in children carrying either *Blastocystis* spp. or the commensal amoeba *Entamoeba coli* or *Endolimax nana*. *Sci. Rep.* 10:15354. doi: 10.1038/s41598-020-72286-y
- Anderson, M. J. (2017). “Permutational multivariate analysis of variance (PERMANOVA),” in *Wiley StatsRef: Statistics Reference Online*, eds N. Balakrishnan, T. Colton, and B. Everitt (Hoboken, NJ: Wiley), 1–15. doi: 10.1002/9781118445112.stat07841
- Antonetti, L., Berrilli, F., Di Cristanziano, V., Farowski, F., Daeumer, M., Eberhardt, K. A., et al. (2024). Investigation of gut microbiota composition in humans carrying *Blastocystis* subtypes 1 and 2 and *Entamoeba hartmanni*. *Gut Pathog.* 16:72. doi: 10.1186/s13099-024-00661-5
- Asnicar, F., Berry, S. E., Valdes, A. M., Nguyen, L. H., Piccinno, G., Drew, D. A., et al. (2021). Microbiome connections with host metabolism and habitual diet from 1,098 deeply phenotyped individuals. *Nat. Med.* 27, 321–332. doi: 10.1038/s41591-020-01183-8
- Audebert, C., Even, G., Cian, A., *Blastocystis* Investigation, Group, Loywick, A., et al. (2016). Colonization with the enteric protozoa *Blastocystis* is associated with increased diversity of human gut bacterial microbiota. *Sci. Rep.* 6:25255. doi: 10.1038/srep25255
- Beghini, F., Pasolli, E., Truong, T. D., Putignani, L., Cacciò, S. M., and Segata, N. (2017). Large-scale comparative metagenomics of *Blastocystis*, a common member of the human gut microbiome. *ISME J.* 11, 2848–2863. doi: 10.1038/ismej.2017.139
- Blackburn, C. C., Yan, S. M., McCormick, D., Herrera, L. N., Iordanov, R. B., Bailey, M. D., et al. (2024). Parasitic contamination of soil in the Southern United States. *Am. J. Trop. Med. Hyg.* 111, 506–514. doi: 10.4269/ajtmh.24-0075
- Callahan, B. J., McMurdie, P. J., Rosen, M. J., Han, A. W., Johnson, A. J. A., and Holmes, S. P. (2016). DADA2: High-resolution sample inference from Illumina amplicon data. *Nat. Methods* 13, 581–583. doi: 10.1038/nmeth.3869
- Caporaso, J. G., Lauber, C. L., Walters, W. A., Berg-Lyons, D., Lozupone, C. A., Turnbaugh, P. J., et al. (2011). Global patterns of 16S rRNA diversity at a depth of millions of sequences per sample. *Proc. Natl. Acad. Sci. U S A.* 108, 4516–4522. doi: 10.1073/pnas.1000080107
- Castañeda, S., Muñoz, M., Villamizar, X., Hernández, P. C., Vásquez, L. R., Tito, R. Y., et al. (2020). Microbiota characterization in *Blastocystis*-colonized and *Blastocystis*-free school-age children from Colombia. *Parasit Vectors* 13:521. doi: 10.1186/s13071-020-04392-9
- Castañeda, S., Tomiak, J., Andersen, L. O. B., Acosta, C. P., Vasquez-A, L. R., Stensvold, C. R., et al. (2025). Impact of *Blastocystis* carriage and colonization intensity on gut microbiota composition in a non-westernized rural population from Colombia. *PLoS Negl. Trop. Dis.* 19:e0013111. doi: 10.1371/journal.pntd.0013111
- Caudet, J., Trelis, M., Cifre, S., Soriano, J. M., and Merino-Torres, J. F. (2022a). Presence and significance of intestinal unicellular parasites in a morbidly obese population. *Int. J. Obes.* 46, 220–227. doi: 10.1038/s41366-021-00980-6
- Caudet, J., Trelis, M., Cifre, S., Soriano, J. M., Rico, H., and Merino-Torres, J. F. (2022b). Interplay between intestinal bacterial communities and unicellular parasites in a morbidly obese population: A neglected trinomial. *Nutrients* 14:3211. doi: 10.3390/nu14153211
- Centers for Disease Control and Prevention [CDC]. (2025). *Blastocystis*. Available online at: https://www.cdc.gov/blastocystis/about/?CDC_AAref_Val=https://www.cdc.gov/parasites/blastocystis/index.html (accessed October 7, 2025).
- Deng, L., and Tan, K. S. W. (2025). Unveiling the hidden ally: *Blastocystis* links healthier diets and cardiometabolic benefits. *Signal Transduct. Target Ther.* 10:77. doi: 10.1038/s41392-025-02146-6
- DeSantis, T. Z., Hugenholtz, P., Larsen, N., Rojas, M., Brodie, E. L., Keller, K., et al. (2006). Greengenes, a chimera-checked 16S rRNA gene database and workbench compatible with ARB. *Appl. Environ. Microbiol.* 72, 5069–5072. doi: 10.1128/AEM.03006-05
- El Safadi, D., Gaayeb, L., Meloni, D., Cian, A., Poirier, P., Wawrzyniak, I., et al. (2014). Children of senegal river basin show the highest prevalence of *Blastocystis* sp. ever observed worldwide. *BMC Infect. Dis.* 14:164. doi: 10.1186/1471-2334-14-164
- Even, G., Lokmer, A., Rodrigues, J., Audebert, C., Viscogliosi, E., Ségurel, L., et al. (2021). Changes in the human gut microbiota associated with colonization by *Blastocystis* sp. and *Entamoeba* spp. in non-industrialized populations. *Front. Cell. Infect. Microbiol.* 11:533528. doi: 10.3389/fcimb.2021.533528
- Geifman, N., and Rubin, E. (2011). Towards an age-phenome knowledge-base. *BMC Bioinformatics* 12:229. doi: 10.1186/1471-2105-12-229
- Heydarian, M., Manouchehri Naeini, K., Kheiri, S., and Abdizadeh, R. (2024). Prevalence and subtyping of *Blastocystis* sp. in ruminants in Southwestern. *Iran. Sci. Rep.* 14:20254. doi: 10.1038/s41598-024-70907-4
- Hublin, J. S. Y., Maloney, J. G., and Santin, M. (2021). *Blastocystis* in domesticated and wild mammals and birds. *Res. Vet. Sci.* 135, 260–282. doi: 10.1016/j.rvsc.2020.09.031
- İşıklar, E. (2018). *Özmen ahmet and Şeniş berat fethi [Özmen ahmet and Şeniş berat fethi]*. Eskişehir: Anadolu Üniversitesi. Turkish.
- Jiménez, P., Muñoz, M., and Ramírez, J. D. (2022). An update on the distribution of *Blastocystis* subtypes in the Americas. *Heliyon* 8:e12592. doi: 10.1016/j.heliyon.2022.e12592
- Jinatham, V., Maxamhud, S., Popleuchai, S., Tsaousis, A. D., and Gentekaki, E. (2021). *Blastocystis* one health approach in a rural community of Northern Thailand: Prevalence, subtypes and novel transmission routes. *Front. Microbiol.* 12:746340. doi: 10.3389/fmicb.2021.746340
- Khorshidvand, Z., Khazaei, S., Amiri, M. R., Taherkhani, H., and Mirzaei, A. (2021). Worldwide prevalence of emerging parasite *Blastocystis* in immunocompromised patients: A systematic review and meta-analysis. *Microb. Pathog.* 152:104615. doi: 10.1016/j.micpath.2020.104615

- Kodio, A., Coulibaly, D., Koné, A. K., Konaté, S., Doumbo, S., Guindo, A., et al. (2019). Blastocystis colonization is associated with increased diversity and altered gut bacterial communities in healthy malian children. *Microorganisms* 7:649. doi: 10.3390/microorganisms7120649
- Koehler, A. V., Herath, H. M. P. D., Hall, R. S., Wilcox, S., and Gasser, R. B. (2024). Marked genetic diversity within Blastocystis in Australian wildlife revealed using a next generation sequencing–phylogenetic approach. *Int. J. Parasitol. Parasites Wildl* 23:100902. doi: 10.1016/j.ijppaw.2023.100902
- Li, H. (2018). Minimap2: Pairwise alignment for nucleotide sequences. *Bioinformatics* 34, 3094–3100. doi: 10.1093/bioinformatics/bty191
- Malatyali, E., Ertabaklar, H., and Ertuğ, S. (2023). Subtype distribution of blastocystis in Türkiye. *Türkiye Parazitoloji Dergisi*. 47, 184–189. doi: 10.4274/tpd.galenos.2023.79188
- Malatyali, E., Tileklioglu, E., Yildiz, I., Unsal, O., Demirag, S., Hatice, E., et al. (2021). Investigation of the relationship between obesity and Blastocystis infection in an adult population in Aydin, Turkey. *Ann. Parasitol.* 67, 249–255. doi: 10.17420/ap6702.336
- Maloney, J. G., Lombard, J. E., Urie, N. J., Shivley, C. B., and Santin, M. (2019). Zoonotic and genetically diverse subtypes of Blastocystis in US pre-weaned dairy heifer calves. *Parasitol. Res.* 118, 575–582. doi: 10.1007/s00436-018-6149-3
- Matovelle, C., Tejedor, M. T., Monteagudo, L. V., Beltrán, A., and Quilez, J. (2022). Prevalence and associated factors of Blastocystis sp. infection in patients with gastrointestinal symptoms in Spain: A case-control study. *Trop. Med. Infect. Dis.* 7:226. doi: 10.3390/tropicalmed7090226
- McCain, A., Grunec, L., Popluechai, S., Tsaousis, A. D., and Gentekaki, E. (2023). Circulation and colonisation of Blastocystis subtypes in schoolchildren of various ethnicities in rural northern Thailand. *Epidemiol. Infect.* 151:e77. doi: 10.1017/S0950268823000596
- Naguib, D., Gantois, N., Desramaut, J., Dominguez, R. G., Arafat, N., Atwa, S. M., et al. (2024). Large-scale molecular epidemiological survey of Blastocystis sp. among herbivores in Egypt and assessment of potential zoonotic risk. *Microorganisms* 12:1286. doi: 10.3390/microorganisms12071286
- Nguyen, L. D. N., Gantois, N., Hoang, T. T., Do, B. T., Desramaut, J., Naguib, D., et al. (2023). First epidemiological survey on the prevalence and subtypes distribution of the enteric parasite Blastocystis sp. in Vietnam. *Microorganisms* 11:731. doi: 10.3390/microorganisms11030731
- Nguyen, L. T., Schmidt, H. A., von Haeseler, A., and Minh, B. Q. (2015). IQ-TREE: A fast and effective stochastic algorithm for estimating maximum likelihood phylogenies. *Mol. Biol. Evol.* 32, 268–274. doi: 10.1093/molbev/msu300
- Nieves-Ramirez, M. E., Partida-Rodríguez, O., Laforest-Lapointe, I., Reynolds, L. A., Brown, E. M., Valdez-Salazar, A., et al. (2018). Asymptomatic intestinal colonization with protist *Blastocystis* is strongly associated with distinct microbiome ecological patterns. *mSystems* 3:e00007-18. doi: 10.1128/mSystems.00007-18
- Noradilah, S. A., Lee, I. L., Anuar, T. S., Salleh, F. M., Manap, S. N. A. A., Mohtar, N. S. H. M., et al. (2016). Occurrence of Blastocystis sp. in water catchments at Malay villages and Aboriginal settlement during wet and dry seasons in Peninsular Malaysia. *PeerJ* 2016:e2541. doi: 10.7717/peerj.2541
- Onder, Z., Yildirim, A., Pekmezci, D., Duzlu, O., Pekmezci, G. Z., Ciloglu, A., et al. (2021). Molecular identification and subtype distribution of Blastocystis sp. in farm and pet animals in Turkey. *Acta Trop.* 220:105939. doi: 10.1016/j.actatropica.2021.105939
- Özkurt, E., Fritscher, J., Soranzo, N., Ng, D. Y. K., Davey, R. P., Bahram, M., et al. (2022). LotuS2: An ultrafast and highly accurate tool for amplicon sequencing analysis. *Microbiome* 10:176. doi: 10.1186/s40168-022-01365-1
- Piperni, E., Nguyen, L. H., Manghi, P., Kim, H., Pasolli, E., Andreu-Sánchez, S., et al. (2024). Intestinal Blastocystis is linked to healthier diets and more favorable cardiometabolic outcomes in 56,989 individuals from 32 countries. *Cell* 187, 4554–4570.e18. doi: 10.1016/j.cell.2024.06.018
- Poirier, P., Wawrzyniak, I., Albert, A., El Alaoui, H., Delbac, F., and Livrelli, V. (2011). Development and evaluation of a real-time PCR assay for detection and quantification of Blastocystis parasites in human stool samples: Prospective study of patients with hematological malignancies. *J. Clin. Microbiol.* 49, 975–983. doi: 10.1128/JCM.01392-10
- Rauff-Adedotun, A. A., Lee, I. L., Abd Talib, N., Shaari, N., Yahaya, Z. S., and Meor Termizi, F. H. (2023). Prevalence, potential risk factors and genetic diversity of Blastocystis in ruminant livestock animals from Penang, Malaysia. *Parasitol. Res.* 122, 2193–2205. doi: 10.1007/s00436-023-07920-5
- Ruang-areerate, T., Piyaraj, P., Suwannahitatorn, P., Ruang-areerate, P., Thita, T., Naaglor, T., et al. (2021). Zoonotic transmission of blastocystis subtype I among people in Eastern communities of thailand: Organic fertilizer from pig feces as a potential source. *Microbiol. Spectr.* 9, 362–321. doi: 10.1128/spectrum.00362-21
- Rudzińska, M., Kowalewska, B., Kurpas, M., and Szostakowska, B. (2022). Rare occurrence of blastocystis in pet animals and their owners in the pomeranian voivodeship in poland in the light of literature data. *J. Clin. Med.* 11:2975. doi: 10.3390/jcm11112975
- Salehi, R., Rostami, A., Mirjalali, H., Stensvold, C. R., and Haghghi, A. (2022). Genetic characterization of Blastocystis from poultry, livestock animals and humans in the southwest region of Iran—Zoonotic implications. *Transbound. Emerg. Dis.* 69, 1178–1185. doi: 10.1111/tbed.14078
- Sanhueza Teneo, D., Chesnais, C. B., Manzano, J., Moll, M. P., Téllez, A., and Valenzuela-Nieto, G. (2025). Waterborne transmission driving the prevalence of Blastocystis sp. in Los Ríos Region, Southern Chile. *Microorganisms* 13:1549. doi: 10.3390/microorganisms13071549
- Santin, M., Figueiredo, A., Molokin, A., George, N. S., Köster, P. C., Dashti, A., et al. (2024). Division of blastocystis ST10 into three new subtypes: ST42-ST44. *J. Eukaryotic Microbiol.* 71:e12998. doi: 10.1111/jeu.12998
- Sarzhonov, F., Dogruman-Al, F., Santin, M., Maloney, J. G., Gureser, A. S., Karasartova, D., et al. (2021). Investigation of neglected protists blastocystis sp. And Dientamoeba fragilis in immunocompetent and immunodeficient diarrheal patients using both conventional and molecular methods. *PLoS Negl. Trop. Dis.* 15:e0009779. doi: 10.1371/JOURNAL.PNTD.0009779
- Segata, N., Izard, J., Waldron, L., Gevers, D., Miropolsky, L., Garrett, W. S., et al. (2011). Metagenomic biomarker discovery and explanation. *Genome Biol.* 12:R60. doi: 10.1186/gb-2011-12-6-r60
- Šejnohová, A., Koutenská, M., Jirků, M., Brožová, K., Pavlíčková, Z., Kadlecová, O., et al. (2024). A cross-sectional survey of Blastocystis sp. and Dientamoeba fragilis in non-human primates and their caregivers in Czech zoos. *One Health* 19:100862. doi: 10.1016/j.onehlt.2024.100862
- Stensvold, C. R. (2025). Blastocystis in stool: Friend, foe or both? *J. Travel Med.* 32:taaf011. doi: 10.1093/jtm/taaf011
- Stensvold, C. R., and Clark, C. G. (2020). Pre-empting pandora's box: Blastocystis subtypes revisited. *Trends Parasitol.* 36, 229–232. doi: 10.1016/j.pt.2019.12.009
- Stensvold, C. R., Arendrup, M. C., Jespersgaard, C., Mølbak, K., and Nielsen, H. V. (2007). Detecting Blastocystis using parasitologic and DNA-based methods: A comparative study. *Diagn. Microbiol. Infect. Dis.* 59, 303–307. doi: 10.1016/j.diagmicrobio.2007.06.003
- Stensvold, C. R., Berg, R. P. K. D., Maloney, J. G., Molokin, A., and Santin, M. (2023). Molecular characterization of Blastocystis and entamoeba of muskoxen and sheep in Greenland. *Int. J. Parasitol.* 53, 673–685. doi: 10.1016/j.ijpara.2023.05.005
- Tan, K. S. W. (2008). New insights on classification, identification, and clinical relevance of Blastocystis spp. *Clin. Microbiol. Rev.* 21, 639–665. doi: 10.1128/cMR.00022-08
- Tavur, A., and Önder, Z. (2022). Molecular prevalence and phylogenetic characterization of blastocystis in cattle in Kayseri Province, Turkey. *Kocatepe Vet. J.* 15, 1–6. doi: 10.30607/kvj.996557
- Tito, R. Y., Chaffron, S., Caenepeel, C., Lima-Mendez, G., Wang, J., Vieira-Silva, S., et al. (2019). Population-level analysis of Blastocystis subtype prevalence and variation in the human gut microbiota. *Gut* 68, 1180–1189. doi: 10.1136/gutjnl-2018-316106
- Tsaousis, A. D., Gentekaki, E., and Stensvold, C. R. (2024). Advancing research on Blastocystis through a one health approach. *Open Res. Eur.* 4:145. doi: 10.12688/openreseurope.18046.1
- Tsaousis, A. D., Shaw, D., Jirku, K., Carmena, D., and Gentekaki, E. (2025). Rethinking Blastocystis: Ubiquity and cyclical abundance in the human gut. *Trends Parasitol.* doi: 10.1016/j.pt.2025.08.009 Online ahead of print.
- Yoshikawa, H., Wu, Z., Pandey, K., Pandey, B. D., Sherchand, J. B., Yanagi, T., et al. (2009). Molecular characterization of Blastocystis isolates from children and rhesus monkeys in Kathmandu, Nepal. *Vet. Parasitol.* 160, 295–300. doi: 10.1016/j.vetpar.2008.11.029