University of Kent

# Kent Academic Repository

**Przelomska, Natalia, Campana, Michael, James, Helen, Kistler, Logan, McInerney, Nancy, Pérez-Escobar, Oscar, Hagemann, Molly, Groombridge, Jim J. and Fleischer, Robert (2025)** ***Population structure and inter-species admixture within a likely extinct yet formerly widespread Hawaiian honeycreeper.*** **Biology Letters, 21 (10). ISSN 1744-9561.**

## Downloaded from

https://kar.kent.ac.uk/111277/ The University of Kent's Academic Repository KAR

## The version of record is available from

https://doi.org/10.1098/rsbl.2025.0265

## This document version

Publisher pdf

## DOI for this version

## Licence for this version

CC BY (Attribution)

## Additional information

## Versions of research works

### Versions of Record

If this version is the version of record, it is the same as the published version available on the publisher's web site. Cite as the published version.

### Author Accepted Manuscripts

If this document is identified as the Author Accepted Manuscript it is the version after peer review but before type setting, copy editing or publisher branding. Cite as Surname, Initial. (Year) 'Title of article'. To be published in ***Title of Journal*** , Volume and issue numbers [peer-reviewed accepted version]. Available at: DOI or URL (Accessed: date).

## Enquiries

If you have questions about this document contact ResearchSupport@kent.ac.uk. Please include the URL of the record in KAR. If you believe that your, or a third party's rights have been compromised through this document please see our Take Down policy (available from https://www.kent.ac.uk/guides/kar-the-kent-academic-repository#policies).

BIOLOGY LETTERS

royalsocietypublishing.org/journal/rsbl

CC BY

Research

**Cite this article:** Przelomska NAS, Campana MG, James HF, Kistler L, Rotzel McInerney N, Pérez-Escobar OA, Hagemann M, Groombridge JJ, Fleischer RC. 2025 Population structure and inter-species admixture within a likely extinct yet formerly widespread Hawaiian honeycreeper. *Biol. Lett.* **21**: 20250265.
https://doi.org/10.1098/rsbl.2025.0265

Received: 18 May 2025
Accepted: 5 September 2025

**Subject Category:**
Population genetics

**Subject Areas:**
taxonomy and systematics, evolution, bioinformatics

**Keywords:**
admixture, Hawaiian honeycreepers, island biogeography, morphology, museum specimens, population genomics

**Author for correspondence:**
Natalia A. S. Przelomska
e-mail: natalia.przelomska@port.ac.uk

Supplementary material is available online at https://doi.org/10.6084/m9.figshare.c.8054551.

THE ROYAL SOCIETY PUBLISHING

# Population structure and inter-species admixture within a likely extinct yet formerly widespread Hawaiian honeycreeper

Natalia A. S. Przelomska[1,2,4], Michael G. Campana[1], Helen F. James[3], Logan Kistler[2], Nancy Rotzel McInerney[1], Oscar A. Pérez-Escobar[5], Molly Hagemann[6], Jim J. Groombridge[7] and Robert C. Fleischer[1]

[1]Center for Conservation Genomics, Smithsonian's National Zoo and Conservation Biology Institute, Smithsonian Institution, Washington, DC, USA
[2]Department of Anthropology, and [3]Department of Vertebrate Zoology, National Museum of Natural History, Smithsonian Institution, Washington, DC, USA
[4]School of the Environment and Life Sciences, University of Portsmouth, Portsmouth, UK
[5]Royal Botanic Gardens, Kew, Richmond, UK
[6]Vertebrate Zoology Department, Bernice Pauahi Bishop Museum, Honolulu, HI, USA
[7]Durrell Institute of Conservation and Ecology, School of Natural Sciences, University of Kent, Canterbury, UK

NASP, 0000-0001-9207-4565; MGC, 0000-0003-0461-6462; HFJ, 0000-0002-2495-6133; LK, 0000-0002-5730-5986; NRM, 0000-0002-6519-7671; OAP-E, 0000-0001-9166-2410; MH, 0000-0002-8475-754X; JJG, 0000-0002-6941-8187; RCF, 0000-0002-2792-7055

The Hawaiian honeycreepers simultaneously represent one of the most spectacular avian adaptive radiations and are one of the most endangered avian groups. This clade's few geographically widespread species can serve as a model to understand population-level processes shaping differentiation and characterizing decline. One such species is the likely extinct ʻōʻū (*Psittirostra psittacea*), a parrot-like beaked honeycreeper with a frugivorous feeding ecology. We compiled morphological and hybridization-captured ancient DNA datasets for the ʻōʻū from museum specimens from across the Hawaiian archipelago. We find (i) genomic differentiation among ʻōʻū from Kauaʻi, Lānaʻi, and the remaining Hawaiian Islands and (ii) a larger phenotype on Kauaʻi and smaller Maui Nui morphological phenotypes. While the differentiated population on Kauaʻi is likely a result of Kauaʻi's geographical isolation, the divergent population on Lānaʻi is harder to explain by biogeography alone. Thus, we investigated whether the unexpected divergence of Lānaʻi ʻōʻū could be attributed to inter-species admixture with the geographically overlapping, now extinct 'parrot-billed' Lānaʻi hookbill (*Dysmorodrepanis munroi*) or a critically endangered Maui endemic, the kiwikiu (*Pseudonestor xanthophrys*). We detect significant admixture between the Lānaʻi ʻōʻū population and the Lānaʻi hookbill, possibly explaining the observed population structure and associating interspecific breeding with populations on the precipice of extinction.

## 1. Introduction

The Hawaiian Islands are the world's most isolated oceanic archipelago and, as such, are a prominent model system for studies in ecology and evolution. This 'living laboratory' has greatly enhanced our understanding of adaptive radiations [1–4] and extinction, phenomena exemplified by the Hawaiian honeycreepers (Fringillidae: Drepanidinae) [5–7]. Hawaiian honeycreepers

© 2025 The Authors. Published by the Royal Society under the terms of the Creative Commons Attribution License http://creativecommons.org/licenses/by/4.0/, which permits unrestricted use, provided the original author and source are credited.

Downloaded from http://royalsocietypublishing.org/rsbl/article-pdf/doi/10.1098/rsbl.2025.0265/2830604/rsbl.2025.0265.pdf
by guest
on 15 December 2025

exhibit high variation in osteological morphology, feeding niche, plumage and song [8,9]. Unfortunately, about two thirds of known Hawaiian honeycreeper species have gone extinct since humans' arrival to the archipelago around AD 1000–1200 [10], including at least 22 species extinct since European arrival in Hawaii and another 18 only known from subfossils [11–14]. The International Union for Conservation of Nature classifies 14 of 17 extant drepanidine species as critically endangered, endangered or vulnerable [15].

Most drepanidine lineages exhibit morphological differentiation and little or no gene flow across the Hawaiian Islands, resulting in the designation of island-specific species and subspecies [5,11]. However, at least three species occurred on all major islands (Kauaʻi, Oʻahu, Molokaʻi, Maui, Lānaʻi and Hawaiʻi), with no currently recognized taxonomic splitting: iʻiwi (*Vestiaria coccinea*), ʻapapane (*Himatione sanguinea*) and ʻōʻū (*Psittirostra psittacea*) [5,16]. The iʻiwi and the ʻapapane remain extant on all major islands (the exception being the extirpation of iʻiwi from Lānaʻi), while the ʻōʻū is probably extinct [17,18]. Feeding ecology is an important driver in the radiation and can be associated with dispersal at the species level and intraspecific connectivity of populations. The iʻiwi and the ʻapapane are primarily nectivorous, whereas the ʻōʻū has a diverse diet but is primarily frugivorous. Thus far, rigorous analyses addressing intraspecific morphological and genetic differentiation have not yet been conducted for any of these three honeycreepers, limiting our understanding of how population-level processes shape the diversity and characterize the decline of widespread species in this radiation.

The ʻōʻū was a heavy-bodied Hawaiian honeycreeper with olive-green plumage and a dappled green (female) or yellow (male) head. It had a thick, parrot-like beak [9], a phenotype shared with only two other drepanidine species: the kiwikiu (*Pseudonestor xanthoprys*) of Maui and Molokaʻi and the Lānaʻi hookbill (*Dysmorodrepanis munroi*), known from Lānaʻi only. Before its disappearance, the ʻōʻū's behaviour was recorded in both traditional ecological knowledge [19] and in observations collected by naturalists and ornithological collectors. The ʻōʻū was observed in mid-elevation mesic to wet ʻōhiʻa forests, primarily feeding on ʻieʻie (*Freycinetia arborea*; Pandanaceae) vine fruits, ʻōhiʻa (*Metrosideros polymorpha*; Myrtaceae) nectar and other native fruits such as ʻōhā (*Clermontia* spp.: Campanulaceae) [8,20]. A strong flier, it was observed to travel in small groups between feeding grounds, depending on the seasonal availability of fruits [9]. Naturalists collected a number of these birds, and some noted minor differences in plumage or size across the archipelago [21–23], differences that are not currently regarded as sufficient for designating subspecies.

Naturalists frequently recorded the ʻōʻū in the 1800s (e.g. [9]). A 1976–1981 population survey estimated a census size of 400 ± 300 individuals on Hawaiʻi and approximately nine individuals on Kauaʻi [24]. The ʻōʻū's last confirmed sighting on Hawaiʻi was in 1987 after a lava flow from Mauna Loa destroyed much of the available ʻōʻū habitat [17]. The final ʻōʻū stronghold was the Alakaʻi swamp on Kauaʻi, where the last accepted sighting was in February 1989 [25]. Sound recordings suggest the Kauaʻi population may have persisted for a couple more years, but no more records were reported following Hurricane Iniki in 1992.

Here, we use morphological and DNA sequence data derived from museum specimens to investigate population structure, diversity and inbreeding within the likely extinct, formerly widespread ʻōʻū. Furthermore, given our unexpected discovery of genomically distinct ʻōʻū individuals from Lānaʻi, we investigated the potential for inter-species admixture across the 'parrot-billed' honeycreepers.

## 2. Materials and methods

### (a) Morphological dataset and analysis

We measured wing chord, culmen length, culmen width, tarsus length and tail length from 89 museum collection study skins in adult plumage (electronic supplementary material, table S1). We excluded culmen width due to a high proportion of missing data (26%). We used ANOVA to assess sexual dimorphism and inter-island morphological variation. To explore variation that could have been obscured by differing island sample sizes, we also applied a permutation approach, running ANOVA on down-sampled island groups. We then applied linear discriminant analysis (LDA) in the R [26] package MASS 7.3-60 [27] to examine whether detected differences in the phenotypic data could be specifically attributed to sex and inter-island differentiation, respectively (for further details, see electronic supplementary material).

### (b) DNA sampling and extraction

A total of 50 toe pad, skin or bone samples representing 49 *Psittirostra psittacea* individuals were obtained from specimens in collections of the American Museum of Natural History (AMNH), the British Museum (BM), the Bernice Pauahi Bishop Museum (BPBM), the Academy of Natural Sciences Drexel University, Philadelphia (ANSP), Muséum National d'Histoire Naturelle, Paris (MNHN), Statens Naturhistoriske Museum, Copenhagen (ZMK), Royal Ontario Museum (ROM), the Museum of Vertebrate Zoology at Berkeley (MVZ) and the Museum of Zoology, University of Cambridge (UMZC) (electronic supplementary material, table S2). AMNH 176717 was sampled twice. Samples were chosen to reflect as broad a distribution across the Hawaiian Islands as possible. For most samples, we used a sterile scalpel to shave an approximately 2 × 2 mm piece of toe pad from each specimen. For a subset of samples, a small piece of dried skin from the abdominal incision site or a small piece of bone was removed from the specimen.

Eighteen samples were extracted in an ancient DNA (aDNA) laboratory at the University of Durham. The remaining samples were extracted in the aDNA laboratory of the Centre for Conservation Genomics, Smithsonian's National Zoo and Conservation Biology Institute (CCG-NZCBI), Washington, DC, USA. We practised stringent aDNA protocols to limit sample contamination and extracted samples according to a previously published protocol [28] (electronic supplementary material).

Downloaded from http://royalsocietypublishing.org/rsbl/article-pdf/doi/10.1098/rsbl.2025.0265/2830604/rsbl.2025.0265.pdf
by guest
on 15 December 2025

Given unexpected differentiation of three ʻōʻū individuals from Lānaʻi (see Results), we investigated the possibility of admixture with two geographically overlapping, 'parrot-billed' Hawaiian honeycreeper species: the kiwikiu and the Lānaʻi hookbill. Hybridization between these species has been suggested previously based on the Lānaʻi hookbill's morphology and rarity [29]. The kiwikiu sample was derived from a modern tissue sample salvaged from the kiwikiu captive breeding programme (BPBM 185328). We obtained a toepad sample from the singular Lānaʻi hookbill specimen (BPBM 4792), an individual collected in 1913. The Lānaʻi hookbill sample was extracted in the CCG-NZCBI aDNA laboratory as described above.

## (c) DNA library preparation and high-throughput sequencing

The initial 18 samples were prepared using KAPA LTP library preparation kits and indexed with iNext primers [30], followed by hybridization-captured using a custom 40 000 drepanidine single nucleotide polymorphism (SNP) bait set (myBaits®: Daicel Arbour Biosciences, Ann Arbour, MI, USA) [31,32]. After evaluation of results, a more efficient library preparation strategy was adopted—the blunt end single tube (BEST) method [33] for the remaining samples. Nine samples were prepared using both KAPA and BEST. The kiwikiu sample was built into KAPA libraries as above. We built a single-stranded library from the Lānaʻi hookbill DNA using the SRSLY® Pico kit (Claret Bioscience, Scotts Valley, CA, USA) (for further details, see electronic supplementary material).

## (d) DNA read processing and single nucleotide polymorphism calling

Adapter sequences and low-quality bases were trimmed from each ʻōʻū sample's reads, and overlapping reads were merged using AdapterRemoval 2.1.7 [34] with the following parameters: --trimqualities --minquality 20 --minalignmentlength 25 --minlength 25 --collapse. Trimmed and merged sequences were then aligned to the draft reference sequence available for another Hawaiian honeycreeper species—the Hawaiʻi ʻamakihi (*Chlorodrepanis virens*; GenBank: GCA_003286495.1 [31]) using BWA 0.7.17 *aln* [35] with the seed disabled [36]. Polymerase chain reaction duplicates were removed using SAMtools 1.9 [37,38]. We estimated cytosine deamination rates and authenticated aDNA damage profiles using MapDamage 2.0.8 [39]. Based on the damage profiles, we masked the terminal two nucleotides of the BEST libraries and the terminal three nucleotides of the KAPA libraries' sequences. For individuals with multiple libraries, we merged the masked alignments using SAMtools. We calculated the number of unique mapped reads and the number of on-target unique mapped reads using SAMtools 1.18 and BEDtools 2.31.0 [40]. We retained 28 ʻōʻū individuals for which we recovered at least 100 000 on-target unique reads.

For the inter-species admixture investigation, we used the ʻakikiki (*Oreomystis bairdi*; SRA: SRR8746532 [41]) as a closely related outgroup to the 'parrot-billed' honeycreepers [42]. The kiwikiu, ʻakikiki, and Lānaʻi hookbill samples were processed following the same pipeline as the ʻōʻū samples except that they were processed with more recent versions of the pipeline software (AdapterRemoval 2.3.2, SAMtools 1.13 and MapDamage 2.2.1) and the modern kiwikiu and ʻakikiki samples were not masked for DNA damage. Based on the Lānaʻi hookbill's MapDamage profile, we masked its sequences' three terminal nucleotides.

For the 28 retained ʻōʻū individuals, we excluded sequences with mapping qualities <25 and called variants using BCFtools 1.18 [38] *mpileup* and *call* (multi-allelic caller model). Using VCFtools 0.1.16 [43], we included only localized, biallelic, autosomal SNPs with no more than 5% missing data per site and minimum minor allele frequencies of 5%. To explore the impact of missing data and allelic drop-out on our population genomic inferences, we filtered data for minimum depths of 1×, 3×, 5× and 10× per site per individual. For the analyses of possible inter-species admixture, we repeated the variant-discovery and filtration pipeline adding the kiwikiu, ʻakikiki and Lānaʻi hookbill samples.

## (e) Population structure, genetic diversity and inbreeding

For each ʻōʻū dataset, we assessed population structure through principal component analysis (PCA) using PCAngsd [44] and *K*-means clustering using ADMIXTURE [45] and NGSadmix [46] (electronic supplementary material). We repeated these analyses accounting for linkage disequilibrium (LD) and putative kin (electronic supplementary material). Further, we repeated a subset of the PCAs using BEAGLE-format likelihoods and did not observe any differences to the results, supporting that these are robust at the genotype depths analysed. Based on the above analyses, we assigned likely populations of origin for two unprovenanced individuals (ANSP 3357 and ANSP 3359). To explore potential inter-island differences and population genomic health, we computed observed and expected homozygosities and per-individual inbreeding coefficients for the 5× and 10× minimum depth datasets using PLINK 2.00a6 [47].

## (f) Inter-species admixture

Based on the population structure results, we defined three ʻōʻū populations: a Kauaʻi population, a Lānaʻi population and a population consisting of individuals from the islands of Hawaiʻi, Maui, Molokaʻi and Oʻahu (hereafter 'HMMO' population). For the inter-species admixture analyses, ANSP 3357 and ANSP 3359 were assigned to the Kauaʻi and HMMO populations, respectively. We calculated *D*- and *f*-branch statistics using Dsuite [48] and explored admixture graphs automatically using

Downloaded from http://royalsocietypublishing.org/rsbl/article-pdf/doi/10.1098/rsbl.2025.0265/2830604/rsbl.2025.0265.pdf
by guest
on 15 December 2025

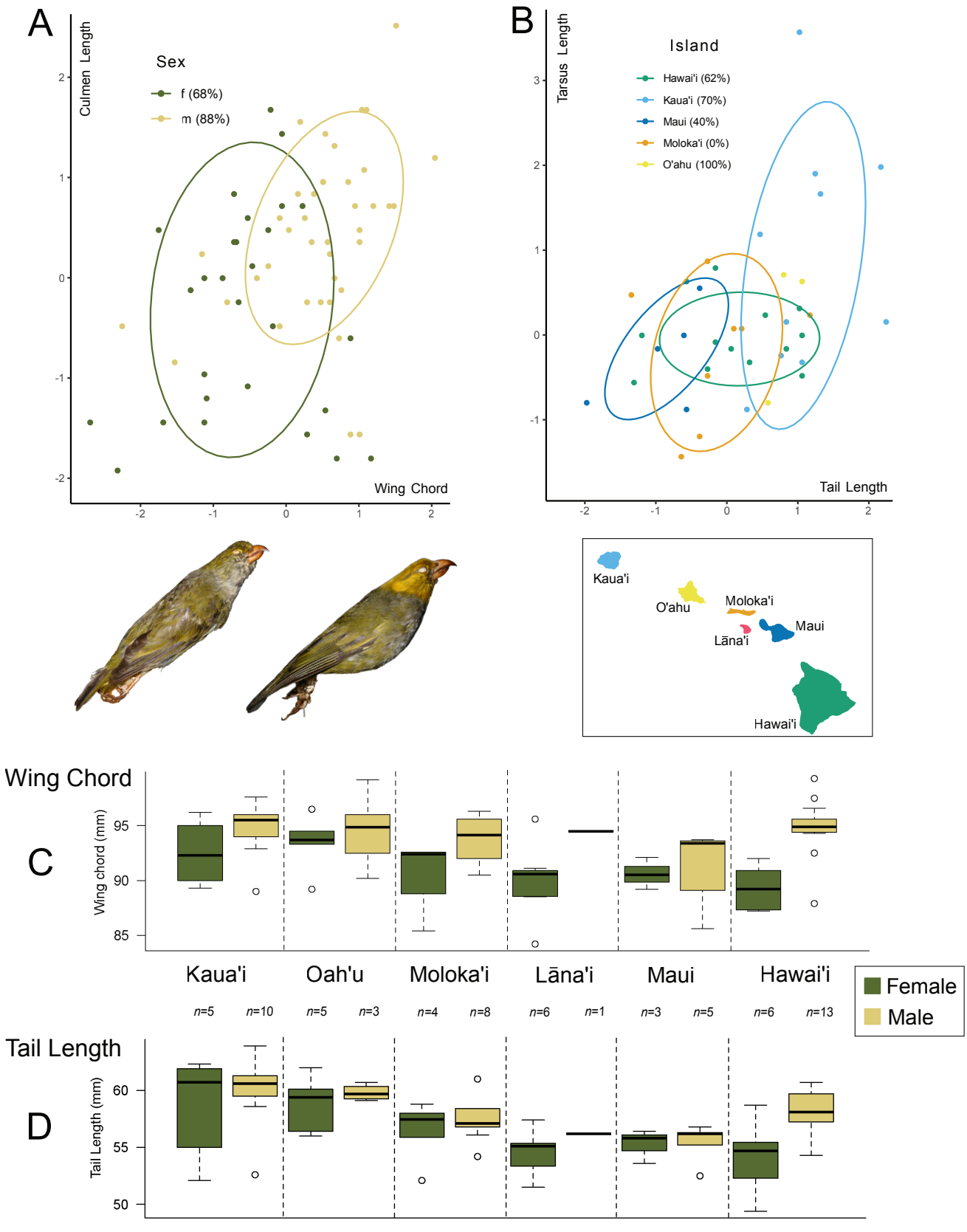


**Figure 1.** Morphometrics based on ʻōʻū specimen measurements split by sex and island group. (A) Culmen length against wing chord differentiating sexes. Ellipses represent 75% confidence intervals. Percentages in the legend represent posterior probabilities of assignment to this group based on LDA on all four analysed traits. (B) Tarsus length against tail length differentiating among islands (male ʻōʻū only). Ellipses represent 68% confidence intervals. Percentages in the legend represent posterior probabilities of assignment to this group based on LDA on all four analysed traits. (C) Boxplots summarizing the wing chord dataset. (D) Boxplots summarizing the tail length dataset. Inset photographs in A (© University Museum of Zoology, University of Cambridge): female ʻōʻū (left) and male ʻōʻū (right).

AdmixTools 2.0.7 [49], allowing up to two admixture edges. Since some ADMIXTURE and NGSadmix analyses clustered individuals from Hawaiʻi, we repeated the Dsuite and AdmixTools analyses, separating individuals from Hawaiʻi from the remaining HMMO individuals.

Downloaded from http://royalsocietypublishing.org/rsbl/article-pdf/doi/10.1098/rsbl.2025.0265/2830604/rsbl.2025.0265.pdf
by guest
on 15 December 2025

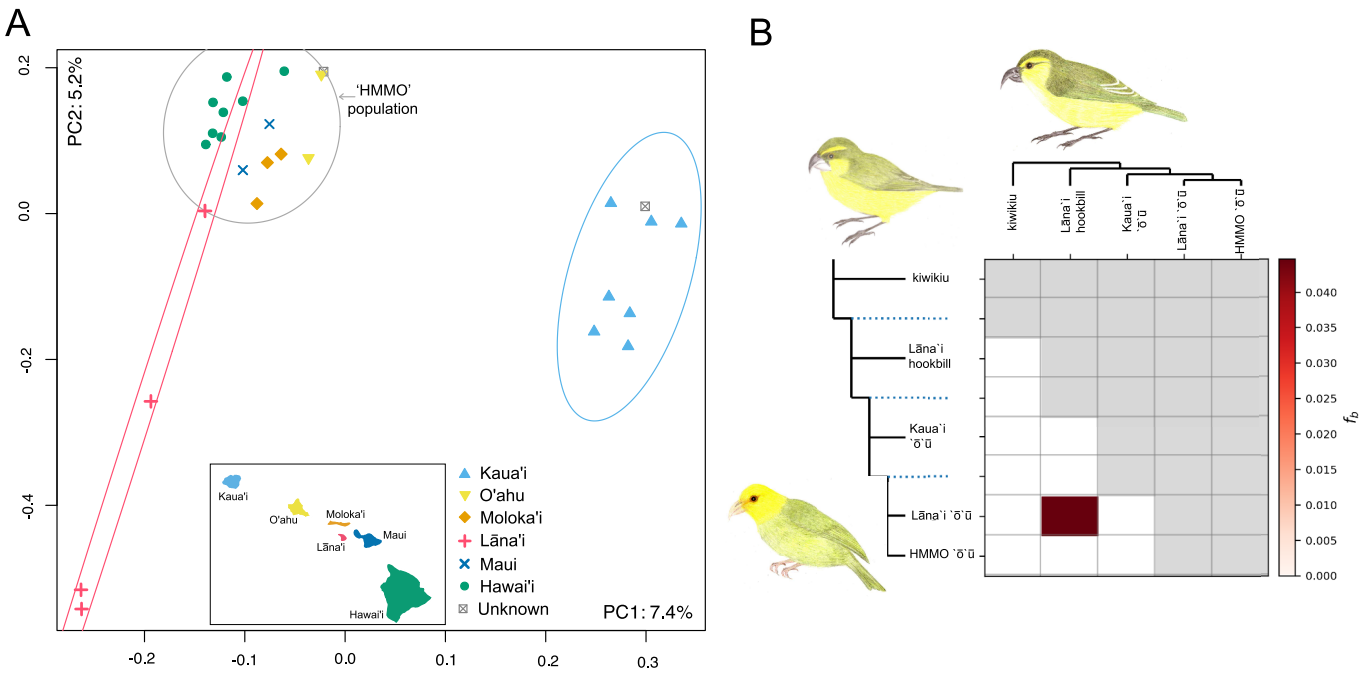


**Figure 2.** (A) Principal component analysis of the 3× minimum depth ʻōʻū dataset showing separation of Kaua'i from other island samples on PC1 and separation of most Lāna'i samples on PC2. Ninety-five per cent data ellipses are drawn for Kaua'i (light blue), Lāna'i (red) and other islands (grey), with a map of the Hawaiian Islands. (B) *f*-branch co-ancestry between Lāna'i ʻōʻū and Lāna'i hookbill in the unpruned 3× minimum depth dataset. Inset illustrations (clockwise from bottom left): male ʻōʻū, kiwikiu, Lāna'i hookbill (Angela Przelomski).

# 3. Results

## (a) Morphological differentiation

The ANOVA on the data split by sex revealed significantly smaller measurements for female birds compared to male birds for the following traits: wing chord, culmen length and tail length ($p < 0.003$), but not for tarsus length (figure 1A,C,D, electronic supplementary material, table S3 and figure S3). The LDA for classification to the correct sex based on phenotypic data resulted in successful predictions for 68% of female birds and 88% of male birds, where wing chord had the highest discriminatory value, followed by culmen length. The dimorphism justified our decision to separate sexes in the tests for inter-island differentiation. Significant inter-island differentiation emerged from the ANOVA tests for tail length and wing chord among male birds ($p < 0.019$) and no traits among the female birds. For wing chord, Maui birds are smaller than those from Hawaiʻi and Kauaʻi and for tail length, Maui, Molokaʻi and Hawaiʻi specimens are smaller than those from Kauaʻi and Maui specimens are smaller than those from Oʻahu (Tukey's honestly significant difference (HSD) test adjusted $p < 0.013$ for all significant instances) (figure 1B,C,D, electronic supplementary material). Although not a replacement for larger sampling sizes, our down-sampled permutations of the data served to demonstrate robustness of the above and indicated smaller culmen length in both females (20% iterations where $n = 3$ for island groups) and males (30% iterations where $n = 3$ for island groups) on Oʻahu. For further details, see electronic supplementary material results, table S4, and figure S3. The LDA for classification to island group based on phenotype for males resulted in two discriminant functions, accounting for 72.2% and 17.5% of the variation, respectively. Tail length had the highest discriminatory value, followed by tarsus length.

## (b) DNA extraction and sequencing

For the 49 sequenced ʻōʻū individuals, we obtained between 2081 and 5 945 326 unique mapped endogenous reads (mean ± s.d.: 1 322 208 ± 1 460 084), of which 270 to 972 936 fell within the bait-targeted regions (mean ± s.d.: 277 931 ± 323 021) (electronic supplementary material, table S5). For the kiwikiu, Lānaʻi hookbill and ʻakikiki samples, we obtained 1 571 614, 6 136 567, and 5 366 968 unique mapped reads, respectively. Of these, 210 252 kiwikiu, 1 546 263 Lānaʻi hookbill, and 1 105 830 ʻakikiki reads mapped to the targeted regions.

We retained 44 273, 9596, 3606 and 737 biallelic SNPs after quality filtering in the 1×, 3×, 5× and 10× minimum depth ʻōʻū datasets, respectively. LD pruning reduced these datasets to 20 221, 4487, 1778 and 430 sites, respectively. For the inter-species admixture analyses, we retained 27 443, 6633, 2673 and 546 SNPs (13 164, 3231, 1350 and 334 after LD pruning) for the 1×, 3×, 5× and 10×minimum depth datasets, respectively. After filtering for depth, residual per-individual missingness was low across all datasets (maximum 18.1%; electronic supplementary material, tables S6–S9).

Downloaded from http://royalsocietypublishing.org/rsbl/article-pdf/doi/10.1098/rsbl.2025.0265/2830604/rsbl.2025.0265.pdf
by guest
on 15 December 2025

### (c) Population genomic structure

The first two to three axes of the PCA (figure 2A, electronic supplementary material, figures S6–S29) indicate clear population structure. PC1 separates Kauaʻi individuals from the remaining ʻōʻū. PC2 and/or PC3 separates out some Lānaʻi individuals. Samples from the other islands form a more compact cluster (HMMO population). Within the HMMO population, we observe possible substructure between the Oʻahu, Maui, Molokaʻi and Hawaiʻi individuals. The unprovenanced ʻōʻū individuals cluster with the Kauaʻi (ANSP 3357) and HMMO (ANSP 3359) individuals. The two-population ADMIXTURE and NGSadmix models separate Kauaʻi from all other samples, while the three-population and higher models also frequently distinguish the Lānaʻi and Hawaiʻi populations (electronic supplementary material figures S30-77).

### (d) Genetic diversity and inbreeding

All ʻōʻū individuals were outbred, with no obvious patterning of homozygosity or inbreeding values across islands (electronic supplementary material, tables S10–S11).

### (e) Inter-species admixture

Dsuite *D*, $f_4$-ratio and *f*-branch statistics found strong evidence of co-ancestry between the Lānaʻi hookbill and Lānaʻi ʻōʻū population (figure 2B, electronic supplementary material, figures S78–S84, table S12). AdmixTools modelling agreed with the Dsuite results (electronic supplementary material, figures S85-108). Models without admixture edges had poor fits (strongly rejecting the null), while allowing admixture significantly improved model fit (electronic supplementary material, table S13). In 29 of 32 (91%) models including one or two admixture edges, the Lānaʻi hookbill was modelled as a mixture of Lānaʻi ʻōʻū-like ancestry and an ancestry that was basal to ʻōʻū (electronic supplementary material). The inferred second admixture edge was not replicable between datasets.

## 4. Discussion

The likely extinct ʻōʻū was known for its migratory behaviour [9], driven by the phenology of fruiting plants, which suggests sustained intraspecific gene flow in spite of its widespread distribution across the Hawaiian Islands. Nonetheless, our population-level analysis of the ʻōʻū revealed geographical structure with a genomically and morphologically divergent population on Kauaʻi as well as a genomically distinct population on Lānaʻi and morphologically smaller individuals from Maui Nui. Kauaʻi is the oldest of the main Hawaiian Islands (emerging approx. 5 Ma and existing in isolation for at least 1 Ma) and the most geographically separated [50], which may be the primary cause of the genomic and morphological differentiation [51]. For instance, Campillo [52] also noted greater divergence of Kauaʻi versus other populations of ʻapapane. The divergence of the Kauaʻi ʻōʻū population is consistent with expectations from biogeography and may reflect incipient reproductive isolation and the initiation of speciation within *Psittirostra* as has been observed in other drepanidine genera [32].

The Lānaʻi differentiation pattern was unexpected as this island is geographically close to Maui, a larger island for which the ʻōʻū fell within the HMMO cluster. Furthermore, the Maui Nui island group of Maui, Lānaʻi, Molokaʻi and Kahoʻolawe comprised a single island until approximately 200 ka with intermittent land connections between these islands (except Kahoʻolawe) existing until the end of the last glacial maximum (approximately 20 ka) [53]. This suggests a Holocene origin of the Lānaʻi population divergence, not supporting the pattern of differentiation observed. Our results document admixture between the Lānaʻi ʻōʻū population and the Lānaʻi hookbill, which could explain this unexpected population structure.

Admixture appears to be rare in Hawaiian honeycreepers [54]. However, admixture in closely related avian species has been documented during population declines (e.g. [55]). The Lānaʻi hookbill was only sighted four times between 1913 and 1918 and collected only once, indicating its extreme rarity by the early twentieth century [56]. Given its decline to extinction at the time, hybridization between the hookbill and the closely related Lānaʻi ʻōʻū is likely. AdmixTools modelled gene flow from the Lānaʻi ʻōʻū into the Lānaʻi hookbill. However, the opposite geneflow direction (from Lānaʻi hookbill into Lānaʻi ʻōʻū) is required if the Lānaʻi ʻōʻū's population divergence resulted from Lānaʻi hookbill introgression. Nevertheless, we cannot rule out bidirectional introgression from our data. Further, we cannot currently identify the source of non-ʻōʻū gene flow into the Lānaʻi hookbill, as we did not exhaustively analyse potential admixing species here. Taxonomically comprehensive, genome-wide analyses will be necessary to establish whether this introgression derives from a ghost lineage. Our findings of admixture into the Lānaʻi hookbill also revive the debate around its taxonomic status (unique species or hybrid) [23], which similarly could be resolved by more in-depth genomic analyses.

Our study documents gene flow between two species of Hawaiian honeycreepers during and prior to their Late Holocene declines, suggesting that interspecific admixture can be promoted in regions undergoing an extinction event. Furthermore, our population-level analysis of the ʻōʻū adds to the evidence that Hawaiian honeycreeper species tend to harbour remarkably high levels of genetic diversity, regardless of inbreeding status, which is generally reflective of their historically large population sizes [57]. The demise of this once widespread species signifies the loss of Hawaiian honeycreeper evolutionary potential and, more specifically, of an element of functional diversity: the ʻōʻū was more strongly frugivorous than any other Hawaiian honeycreeper species whose diet is known.

Downloaded from http://royalsocietypublishing.org/rsbl/article-pdf/doi/10.1098/rsbl.2025.0265/2830604/rsbl.2025.0265.pdf
by guest
on 15 December 2025

Ethics. All specimens sampled for this study are from museum collections, with respective loans for their use (see Methods, Acknowledgements and ESM2 supplementary tables S1 and S2).

Data accessibility. Raw genetic sequence data have been deposited in NCBI (Bioproject PRJNA1244941). Raw morphological measurement data is available in electronic supplementary material, table 1 (in ESM file 2).

Supplementary material is available online [58].

Declaration of AI use. We have not used AI-assisted technologies in creating this article.

Authors' contributions. N.A.S.P.: conceptualization, formal analysis, investigation, methodology, visualization, writing—original draft, writing—review and editing; M.G.C.: data curation, formal analysis, methodology, visualization, writing—original draft, writing—review and editing; H.F.J.: conceptualization, formal analysis, investigation, methodology, resources, writing—review and editing; L.K.: formal analysis, resources, supervision, writing—review and editing; N.R.McI.: investigation, writing—review and editing; O.A.P-E.: formal analysis, visualization, writing—review and editing; M.H.: resources, writing—review and editing; J.J.G.: resources, writing—review and editing; R.C.F.: conceptualization, funding acquisition, investigation, methodology, supervision, writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare we have no competing interests.

Funding. Funding was provided by a National Science Foundation EEID grant number DEB-1717498, the Beeson Fund, a Smithsonian Institution Buck Fellowship awarded to N.A.S.P. and Smithsonian Institution funds awarded to L.K. J.J.G. was supported by Research England's Expanding Excellence in England (E3) Fund, UK Research and Innovation.

Acknowledgements. We thank the curators of the museums from which samples were collected, especially Paul Sweet at AMNH, Jason Weckstein and Nate Rice at ANSP, and Michael Brooke at UMZC. We also thank Michael Brooke for providing photographs of specimens, where copyright remains with the UMZC. We acknowledge Storrs Olson's contribution to the morphological dataset. We thank Taylor Callicrate for the kiwikiu laboratory work and Lillian Parker and Hernan Morales Villegas for helpful discussions regarding DNA library preparation and genetic diversity respectively. We also thank Alexandre Antonelli for constructive comments on the manuscript. Laboratory work was conducted in, and with the support of both modern and ancient DNA laboratories of the Center for Conservation Genomics, Smithsonian National Zoo and Conservation Biology Institute (NZCBI), Department of Biological Sciences, University of Durham, and the Laboratories for Analytical Biology (LAB) facilities of the Smithsonian's National Museum of Natural History (NMNH). Computations were conducted on the Smithsonian High Performance Cluster (HPC), Smithsonian Institution: https://doi.org/10.25572/SIHPC.

# References

1. Baldwin BG, Sanderson MJ. 1998 Age and rate of diversification of the Hawaiian silversword alliance (Compositae). *Proc. Natl Acad. Sci. USA* **95**, 9402–9406. (doi:10.1073/pnas.95.16.9402)
2. Givnish TJ *et al*. 2009 Origin, adaptive radiation and diversification of the Hawaiian lobeliads (Asterales: Campanulaceae). *Proc. R. Soc. B* **276**, 407–416. (doi:10.1098/rspb.2008.1204)
3. Kaneshiro KY. 1988 Speciation in the Hawaiian *Drosophila*: sexual selection appears to play an important role. *Bioscience* **38**, 258–263. (doi:10.2307/1310849)
4. Fleischer RC, Campana MG, James HF. 2022 Hawaiian songbird radiations. *Curr. Biol*. **32**, R1070–R1072. (doi:10.1016/j.cub.2022.08.057)
5. Amadon D. 1950 The Hawaiian honeycreepers (Aves, Drepaniidae). In *Bulletin of the AMNH; v. 95, article 4*. New York, NY: American Museum of Natural History.
6. Berger AJ. 1970 The present status of the birds of Hawai'i. *Pac. Sci.* **24**, 29–42.
7. Freed LA, Conant S, Fleischer RC. 1987 Evolutionary ecology and radiation of Hawaiian passerine birds. *Trends Ecol. Evol*. **2**, 196–203. (doi:10.1016/0169-5347(87)90020-6)
8. James HF, Olson SL. 1991 Descriptions of thirty-two new species of birds from the Hawaiian Islands: part II. Passeriformes. *Ornithol. Monogr*. 1–88. (doi:10.2307/40166713)
9. Perkins RCL. 1903 Vertebrata (aves). In *Fauna Hawaiiensis; being the land-fauna of the Hawaiian Islands*. (ed. D Sharp), pp. 368–465, vol. **1**. Cambridge, UK: Cambridge University Press.
10. Kirch PV. 2011 When did the Polynesians settle Hawai'i? A review of 150 years of scholarly inquiry and a tentative answer. *Impact Prehist. Polyn. Hawaii. Ecosyst. Hawaii Archaeol* **12**, 3–26. https://hdl.handle.net/10524/74851
11. James HF. 2004 The osteology and phylogeny of the Hawaiian finch radiation (Fringillidae: Drepanidini), including extinct taxa. *Zool. J. Linn. Soc.* **141**, 207–255. (doi:10.1111/j.1096-3642.2004.00117.x)
12. Olson SL, James HF. 1984 The role of polynesians in the extinction of the Avifauna of the Hawaiian Islands. In *Quaternary extinctions: a prehistoric revolution* (eds PL Martin, RG Klein), pp. 768–780. Tucson, AZ: University of Arizona Press. (doi:10.2307/j.ctv264f91j.44)
13. Athens JS, Toggle HD, Ward JV, Welch DJ. 2002 Avifaunal extinctions, vegetation change, and polynesian impacts in prehistoric Hawai'i. *Archaeol. Ocean*. **37**, 57–78. (doi:10.1002/j.1834-4453.2002.tb00507.x)
14. Boyer AG. 2008 Extinction patterns in the avifauna of the Hawaiian islands. *Divers. Distrib*. **14**, 509–517. (doi:10.1111/j.1472-4642.2007.00459.x)
15. International Union for the Conservation of Nature. 2025 *The IUCN red list of threatened species*. Version 2025-v1. See https://www.iucnredlist.org/.
16. Fancy SG, Ralph CJ. 1997 Apapane (*Himatione sanguinea*). In *The birds of North America, no. 296* (eds A Poole, F Gill). Ithaca, NY: Cornell Lab of Ornithology. (doi:10.2173/bna.296)
17. BirdLife International. 2018 *Psittirostra psittacea* (amended version of 2016 assessment). *The IUCN red list of threatened species. E.T22720734A126791352*. See https://www.iucnredlist.org/species/22720734/126791352.
18. United States Fish and Wildlife Service. 2022 'Ō'ū (*Psittirostra psittacea*) 5-year review summary and evaluation. Honolulu, HI: Pacific Islands Fish and Wildlife Office. See https://ecos.fws.gov/docs/five_year_review/doc2541.pdf.
19. Gomes NJ. 2020 Reclaiming native Hawaiian knowledge represented in bird taxonomies. *Ethnobiol. Lett*. **11**, 30–43. (doi:10.14237/ebl.11.2.2020.1682)
20. Snetsinger TJ, Reynolds MH, Herrmann CM. 1998 'O'u (*Psittirostra psittacea*). In *The birds of North America, no. 335* (eds A Poole, F Gill). Ithaca, NY: Cornell Lab of Ornithology. (doi:10.2173/bna.ou.02)
21. Stejneger L. 1887 Notes on *Psittirostra psittacea* from Kauai, Hawaiian Islands. *Proc. US. Natl. Mus.* **10**, 389–390. (doi:10.5479/si.00963801.640.389)
22. Bangs O. 1911 Two new birds from the island of Molokai. *Proc. Biol. Soc Wash*. **24**, 29–30.
23. Rothschild W. 1905 Renaming of O'ahu' O'u to *Psittirostra psittacea deppei*. *Bull. Brit. Ornith. Club* **15**, 45.
24. Ellis S, Kuehler C, Lacy R, Hughes K, Seal US, Captive Breeding Specialist Group. 1992 *Hawaiian forest birds conservation assessment and management plan*. Gland, Switzerland: IUCN – The World Conservation Union.
25. Pyle RL. 1989 Hawaiian islands region. *Am. Birds.* **43**, 369–371.
26. R Core Team. 2025 R: a language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing. See http://www.R-project.org/.

Downloaded from http://royalsocietypublishing.org/rsbl/article-pdf/doi/10.1098/rsbl.2025.0265/2830604/rsbl.2025.0265.pdf
by guest
on 15 December 2025

27. Venables WN, Ripley BD. 2002 Random and mixed effects. In *Modern applied statistics with S* (eds WN Venables, BD Ripley), pp. 271–300. New York, NY: Springer. (doi:10.1007/978-0-387-21706-2_10)
28. Fleischer RC, Olson SL, James HF, Cooper AC. 2000 Identification of the extinct Hawaiian eagle (*Haliaeetus*) by mtDNA sequence analysis. *Auk* **117**, 1051–1056. (doi:10.1093/auk/117.4.1051)
29. James HF, Zusi RI, Olson SL. 1989 *Dysmorodrepanis munroi* (Fringillidae: Drepanidini), a valid genus and species of Hawaiian finch. *Wilson Bull.* **101**, 159–179.
30. Glenn TC *et al*. 2019 Adapterama I: universal stubs and primers for 384 unique dual-indexed or 147,456 combinatorially-indexed Illumina libraries (iTru & iNext). *PeerJ* **7**, e7755. (doi:10.7717/peerj.7755)
31. Callicrate T, Dikow R, Thomas JW, Mullikin JC, Jarvis ED, Fleischer RC, NISC Comparative Sequencing Program. 2014 Genomic resources for the endangered Hawaiian honeycreepers. *BMC Genom*. **15**, 1098. (doi:10.1186/1471-2164-15-1098)
32. Cassin-Sackett L, Callicrate TE, Fleischer RC. 2019 Parallel evolution of gene classes, but not genes: evidence from Hawai'ian honeycreeper populations exposed to avian malaria. *Mol. Ecol*. **28**, 568–583. (doi:10.1111/mec.14891)
33. Carøe C, Gopalakrishnan S, Vinner L, Mak SST, Sinding MHS, Samaniego JA, Wales N, Sicheritz-Pontén T, Gilbert MTP. 2018 Single-tube library preparation for degraded DNA. *Methods Ecol. Evol*. **9**, 410–419. (doi:10.1111/2041-210x.12871)
34. Schubert M, Lindgreen S, Orlando L. 2016 AdapterRemoval v2: rapid adapter trimming, identification, and read merging. *BMC Res. Notes* **9**, 88. (doi:10.1186/s13104-016-1900-2)
35. Li H, Durbin R. 2009 Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics* **25**, 1754–1760. (doi:10.1093/bioinformatics/btp324)
36. Schubert M, Ginolhac A, Lindgreen S, Thompson JF, AL-Rasheid KA, Willerslev E, Krogh A, Orlando L. 2012 Improving ancient DNA read mapping against modern reference genomes. *BMC Genom*. **13**, 178. (doi:10.1186/1471-2164-13-178)
37. Li H *et al*. 2009 The sequence Alignment/Map format and SAMtools. *Bioinformatics* **25**, 2078–2079. (doi:10.1093/bioinformatics/btp352)
38. Danecek P *et al*. 2021 Twelve years of SAMtools and BCFtools. *Gigascience* **10**, giab008. (doi:10.1093/gigascience/giab008)
39. Jónsson H, Ginolhac A, Schubert M, Johnson PLF, Orlando L. 2013 mapDamage2.0: fast approximate Bayesian estimates of ancient DNA damage parameters. *Bioinformatics* **29**, 1682–1684. (doi:10.1093/bioinformatics/btt193)
40. Quinlan AR, Hall IM. 2010 BEDTools: a flexible suite of utilities for comparing genomic features. *Bioinformatics* **26**, 841–842. (doi:10.1093/bioinformatics/btq033)
41. Cassin-Sackett L *et al*. 2021 Genetic structure and population history in two critically endangered Kaua'i honeycreepers. *Conserv. Genet*. **22**, 601–614. (doi:10.1007/s10592-021-01382-x)
42. Lerner HRL, Meyer M, James HF, Hofreiter M, Fleischer RC. 2011 Multilocus resolution of phylogeny and timescale in the extant adaptive radiation of Hawaiian honeycreepers. *Curr. Biol*. **21**, 1838–1844. (doi:10.1016/j.cub.2011.09.039)
43. Danecek P *et al*. 2011 The variant call format and VCFtools. *Bioinformatics* **27**, 2156–2158. (doi:10.1093/bioinformatics/btr330)
44. Meisner J, Albrechtsen A. 2018 Inferring population structure and admixture proportions in low-depth NGS data. *Genetics* **210**, 719–731. (doi:10.1534/genetics.118.301336)
45. Alexander DH, Novembre J, Lange K. 2009 Fast model-based estimation of ancestry in unrelated individuals. *Genome Res*. **19**, 1655–1664. (doi:10.1101/gr.094052.109)
46. Skotte L, Korneliussen TS, Albrechtsen A. 2013 Estimating individual admixture proportions from next generation sequencing data. *Genetics* **195**, 693–702. (doi:10.1534/genetics.113.154138)
47. Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. 2015 Second-generation PLINK: rising to the challenge of larger and richer datasets. *GigaScience* **4**, 7. (doi:10.1186/s13742-015-0047-8)
48. Malinsky M, Matschiner M, Svardal H. 2021 Dsuite—fast D-statistics and related admixture evidence from VCF files. *Mol. Ecol. Resour*. **21**, 584–595. (doi:10.1111/1755-0998.13265)
49. Maier R, Flegontov P, Flegontova O, Işıldak U, Changmai P, Reich D. 2023 On the limits of fitting complex models of population history to f-statistics. *eLife* **12**, e85492. (doi:10.7554/elife.85492)
50. Carson HL, Clague DA. 1995 Geology and biogeography. In *Hawaiian biogeography: evolution on a hot spot archipelago* (eds WL Wagner, VA Funk), pp. 14–29. Washington, DC and London, UK: Smithsonian Institution Press.
51. Fleischer RC, Mcintosh CE, Tarr CL. 1998 Evolution on a volcanic conveyor belt: using phylogeographic reconstructions and K–Ar-based ages of the Hawaiian Islands to estimate molecular evolutionary rates. *Mol. Ecol*. **7**, 533–545. (doi:10.1046/j.1365-294x.1998.00364.x)
52. Campillo LC. 2022 Patterns and processes shaping avian diversity in the Hawaiian islands. PhD Thesis, University of Hawai'i at Mānoa, Honolulu, HI.
53. Price JP, Elliott-Fisk D. 2004 Topographic history of the Maui Nui complex, Hawai'i and its implications for biogeography. *Pac. Sci.* **58**, 27–45. (doi:10.1353/psc.2004.0008)
54. Knowlton JL, Flaspohler DJ, Rotzel Mcinerney NC, Fleischer RC. 2014 First record of hybridization in the Hawaiian honeycreepers: 'I'iwi (*Vestiaria coccinea*) × 'Apapane (*Himatione sanguinea*). *Wilson J. Ornithol*. **126**, 562–568. (doi:10.1676/13-054.1)
55. Kersten O *et al*. 2023 Hybridization of Atlantic puffins in the Arctic coincides with 20th-century climate change. *Sci. Adv*. **9**, eadh1407. (doi:10.1126/sciadv.adh1407)
56. Perkins RCL. 1919 XXIII.— On a new genus and species of bird of the family Drepanididæ from the Hawaiian islands. *Ann. Mag. Nat. Hist.* **3**, 250–252. (doi:10.1080/00222931908673819)
57. Kyriazis CC *et al*. 2025 Population genomics of recovery and extinction in Hawaiian honeycreepers. *Curr. Biol.* **35**, 2697–2708. (doi:10.1016/j.cub.2025.04.078)
58. Przelomska N, Campana MG, James H, Kistler L, Mclnerney NR, Pérez-Escobar O *et al*. 2025 Supplementary material from "Population structure and inter-species admixture within a likely extinct yet formerly widespread Hawaiian honeycreeper. Figshare. (doi:10.6084/m9.figshare.c.8054551)


Downloaded from http://royalsocietypublishing.org/rsbl/article-pdf/doi/10.1098/rsbl.2025.0265/2830604/rsbl.2025.0265.pdf
by guest
on 15 December 2025