

Endostructural morphology in hominoid mandibular third premolars: Discrete traits at the enamel-dentine junction

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2 enamel-dentine junction

3

#### 4 **Abstract**

5 The mandibular third premolar ( $P_3$ ) exhibits substantial differences in size and shape among  
6 hominoid taxa, and displays a number of discrete traits that have proven to be useful in studies  
7 of hominin taxonomy and phylogeny. Discrete traits at the enamel-dentine junction (EDJ) can  
8 be accurately assessed on moderately worn specimens, and often appear sharper than at the  
9 outer-enamel surface (OES). Here we use microtomography to image the  $P_3$  EDJ of a broad  
10 sample of extant apes, extinct hominins and modern humans ( $n = 100$ ). We present typologies  
11 for three important premolar discrete traits at the EDJ (transverse crest, marginal ridge and  
12 buccal grooves), and score trait frequencies within our sample. We find that the transverse  
13 crest is variable in extant apes, while the majority of hominins display a transverse crest  
14 which runs directly between the two major premolar cusps. Some Neanderthals display a  
15 unique form in which the transverse crest fails to reach the protoconid. We find that mesial  
16 marginal ridge discontinuity is common in *Australopithecus anamensis* and *Australopithecus*  
17 *afarensis* while continuous marginal ridges largely characterize *Australopithecus africanus*  
18 and *Paranthropus*. Interrupted mesial and distal marginal ridges are again seen in *Homo*  
19 *sapiens* and Neanderthals. Premolar buccal grooves, previously identified at the OES as  
20 important for hominin systematics, are again found to show a number of taxon-specific  
21 patterns at the EDJ, including a clear difference between *Australopithecus* and *Paranthropus*  
22 specimens. However, their appearance may be dependent on the morphology of other parts of  
23 the crown such as the protoconid crest, and the presence of accessory dentine horns. Finally,  
24 we discuss rare variations in the form of dentine horns that underlie premolar cusps, and their  
25 potential homology to similar morphologies in other tooth positions.

## 27 **1. Introduction**

28 Teeth are an important component of the fossil record; as highly mineralized and compact  
29 tissues, they are well preserved and, therefore, common in fossil deposits. They are also a rich  
30 source of information regarding taxonomy, diet, and environment, among other factors  
31 (Walker et al., 1978; Sponheimer and Lee-Thorp, 1999; Richards et al., 2001; Lee-Thorp et  
32 al., 2003; Grine et al., 2012). Unlike bones, the external morphology of the tooth crown is not  
33 remodeled throughout life; once fully developed, it is only modified by external factors such  
34 as breakage or wear. Further, aspects of tooth morphology show high levels of heritability  
35 (Townsend and Martin, 1992; Dempsey and Townsend, 2001; Hlusko and Mahaney, 2003)  
36 and dental traits are considered particularly useful in studies of taxonomy and phylogeny  
37 (Wood and Abbott, 1983, Wood et al., 1983; Suwa et al., 1996; Irish and Guatelli-Steinberg,  
38 2003).

39 Many previous studies of the hominin dentition have focussed on molars, since they are the  
40 most morphologically complex teeth, while premolars are considered transitional teeth  
41 between the simple, single-cusped canines and the more complex, multi-cusped molars.  
42 Premolars are extremely variable, however, and can show a variety of morphologies even  
43 among hominoids (e.g., Davies et al., 2019). This is especially true of the mandibular third  
44 premolar ( $P_3$ ), which forms part of the catarrhine canine honing complex (e.g., Walker, 1984;  
45 Greenfield and Washburn, 1992; Delezene, 2015). As a result of substantial reduction in  
46 maxillary canine height, hominins are the only catarrhine clade in which the function of  
47 canine honing has been lost (e.g., Greenfield, 1990; Brunet et al., 2002; Haile-Selassie et al.,  
48 2004; Suwa et al., 2009). The loss of canine honing may have removed a functional constraint  
49 on  $P_3$  morphology that permitted the evolution of novel  $P_3$  forms among hominins.

50 Intraspecific P<sub>3</sub> variability was noted by Kraus and Furr (1953), who outlined 17 traits in  
51 modern humans that relate to the development and position of the major cusps, the form of  
52 major occlusal ridges, and the presence of features such as buccal grooves. Wood and  
53 Uytterschaut (1987) built upon these definitions to study mandibular premolar variation  
54 among Plio-Pleistocene hominins, noting for example that cusp number is effective in  
55 distinguishing between *Paranthropus* and other hominins, while the position of the lingual  
56 cusp can be used to distinguish eastern African *Homo*. Further, Suwa (1988) identified a  
57 number of P<sub>3</sub> features that distinguish *Paranthropus* from other hominins and Suwa et al.  
58 (1996) outlined derived P<sub>3</sub> features of early *Homo* and *Australopithecus africanus*, relative to  
59 *Australopithecus afarensis*, as well as those unique to early *Homo*. P<sub>3</sub> traits have also been  
60 utilized in studies of *Ardipithecus* (Haile-Selassie et al., 2004; Suwa et al., 2009),  
61 *Australopithecus anamensis*, *A. afarensis* (Leonard and Hegmon, 1987; Ward et al., 2001;  
62 Kimbel et al., 2006; Deleuzene and Kimbel, 2011) and *Homo naledi* (Irish et al., 2018).

63 Nonmetric features are typically assessed at the outer-enamel surface (OES); however,  
64 advances in microtomography allow the assessment of internal tooth structures. Of particular  
65 interest in studying discrete dental traits is the enamel-dentine junction (EDJ). The EDJ  
66 preserves the form of the basement layer of the inner-enamel epithelium, the morphology of  
67 which is determined during odontogenesis. Subsequently, enamel and dentine are deposited  
68 on either side of this epithelial layer, meaning that important OES dental features such as  
69 cusps and crests typically originate at the EDJ (Skinner et al., 2008 and references therein).  
70 The advantages of studying the morphology of the EDJ are twofold. Firstly, tooth wear can  
71 prevent assessment of discrete dental traits (Burnett et al., 2013); however, the EDJ is  
72 preserved in specimens with moderate wear, allowing assessment of specimens that would  
73 otherwise be excluded from analyses. Secondly, hominin teeth often have thick enamel  
74 (Beynon and Wood, 1986; Macho and Thackeray, 1992; Smith et al., 2012; Skinner et al.,

75 2015), which may make the exact form and layout of crown features difficult to assess. In  
76 contrast, features at the EDJ appear much sharper, allowing for more accurate and precise trait  
77 assessment. As a result, although the OES and EDJ have a high level of correspondence  
78 (Nager, 1960; Skinner et al., 2010; Ortiz et al., 2012; Morita et al., 2014; Guy et al., 2015),  
79 trait-scoring systems developed for the OES may not always be applicable when studying the  
80 EDJ.

81 Various studies have investigated the expression of discrete molar traits at the EDJ (e.g.,  
82 Korenhof, 1960; Corruccini, 1987; Skinner et al., 2008; Anemone et al., 2012; Zanolli and  
83 Mazurier, 2013; Martín-Torres et al., 2014; Martínez de Pinillos et al., 2014; Zanolli, 2015;  
84 Martin et al., 2017; Liao et al., 2019), which help to understand the range of variation in the  
85 form of these traits. Studying the EDJ also allows us to better understand the development of  
86 dental traits so that we can accurately identify traits and their covariation with one another, or  
87 with other aspects of crown morphology, both of which can be important when using traits to  
88 make taxonomic arguments. Fewer studies have explored discrete premolar traits at the EDJ,  
89 although a number of studies have done so alongside molars (e.g., Braga et al., 2010; Liu et  
90 al., 2013; Zanolli et al., 2018). Sakai (1967) studied the modern human P<sub>3</sub> using destructive  
91 techniques to view the EDJ and explore the expression of discrete traits such as buccal  
92 grooves and an indentation on the mesial marginal ridge termed a ‘trigonid notch’. The form  
93 of these traits was compared to the morphology of the OES, and discussed with reference to  
94 what was known about the expression of these traits at the OES of fossil hominins. Further,  
95 Krenn et al. (2019) studied modern human mandibular premolars at the EDJ and OES, scoring  
96 the variation in seven different qualitative traits including the form of the marginal ridges, the  
97 extension of the transverse crest, and the number of accessory crests present on the crown,  
98 finding inter-population differences in a number of traits. Despite advances in  
99 microtomography allowing the non-destructive imaging of internal tooth structures in fossils,

100 no study has yet explored the EDJ expression of P<sub>3</sub> discrete traits across the hominin clade.  
101 Here we examine the P<sub>3</sub> EDJ of a broad sample of extant apes, modern humans and extinct  
102 hominins, focussing on three important discrete traits that are variable within this sample. We  
103 present scoring systems for these traits, and discuss the concordance (or lack thereof) with  
104 traits previously scored at the OES. We score trait frequencies within our sample to identify  
105 features that may have taxonomic significance, and discuss the developmental basis of a  
106 number of these traits.

107 A second companion paper will use geometric morphometrics to quantitatively analyze  
108 hominoid P<sub>3</sub> EDJ morphology (Davies et al. 2019).

109

## 110 **2. Methods**

### 111 *2.1 Study sample*

112 The sample is summarized in Table 1 (a full list of specimens can be found in  
113 Supplementary Online Material [SOM] Table S1) and consists of 100 specimens, of which 93  
114 are assigned to species rank. Two specimens are assigned to *Homo* sp., and five are  
115 considered indeterminate. The recent *H. sapiens* sample is curated at the University of Leipzig  
116 Anatomical Collection (ULAC). Relatively little information is available on the provenance  
117 of this sample, but the available information is presented in SOM Table S2. The study sample  
118 is limited by the availability of microtomographic scans and the ability to extract the EDJ  
119 surface from those scans. The EDJ surface is more difficult to extract in some cases due to a  
120 lack of radiographic tissue contrast between enamel and dentine; this problem is more  
121 prevalent at eastern African sites (although the taphonomic processes that are likely causing  
122 this are not clear), and the sample sizes for taxa known mostly from these sites (e.g.,  
123 *Paranthropus boisei*, *A. anamensis*, *A. afarensis*, and early *Homo*) are more limited. Further,  
124 the physical preservation of the fossils is important, and specimens that are heavily cracked,

125 eroded or otherwise broken often have to be excluded from analyses when digital  
126 reconstruction is not possible. Scan resolution is also important to consider: larger traits such  
127 as main cusps and crests remain visible at the entire range of scan resolutions used; however,  
128 smaller features such as accessory cusps can be difficult to identify at lower resolutions,  
129 particularly in poorer quality scans. Scan resolution is limited by a number of factors  
130 including the shape and size of the specimen, as this limits how closely the specimen can be  
131 placed to the X-ray source, as well as the time available for scanning. Nonetheless, the sample  
132 does capture a broad swath of the hominin clade, including modern humans, and includes  
133 representatives of all extant apes as comparative outgroups. Given sample size constraints, it  
134 is very likely that this study does not capture the full extent of P<sub>3</sub> variation in each taxon;  
135 however, the trait scoring systems outlined can be expanded as necessary.

136

## 137 2.2 *Microtomography and segmentation*

138 Microtomographic scans of P<sub>3</sub> sample were obtained using either a SkyScan 1173 at 100–  
139 130 kV and 90–130  $\mu$ A, a BIR ACTIS 225/300 scanner at 130 kV and 100–120  $\mu$ A, or a  
140 Diondo d3 at 100–140kV and 100–140  $\mu$ A, and reconstructed as 8-bit TIFF stacks (isometric  
141 voxel resolutions ranging from 13–45  $\mu$ m). TIFF stacks were filtered using 3D median filter,  
142 followed by a mean of least variance filter (each with a kernel size of either one or three),  
143 implemented using MIA open source software (Wollny et al., 2013). Filtered image stacks  
144 were segmented using Avizo 6.3 (Visualization Sciences Group, 2010) to produce PLY  
145 format surface models of the EDJ. The EDJ of specimens with substantial cracks were  
146 realigned using Geomagic Studio 2014 (3D systems, Rock Hill) when possible.

147



148 2.3 *Scoring procedures*

149 The presence and form of four EDJ discrete traits were scored; the transverse crest,  
150 marginal ridge continuity, mesial buccal groove and distal buccal groove. While each of these  
151 traits has been previously described at the OES for hominoids, the extent of variation in these  
152 traits at the EDJ has not been previously described. These traits were chosen because initial  
153 investigations showed that they are variable within our sample, and are able to be scored in a  
154 way that is both precise and reproducible. Since traits were only scored at the EDJ, and not at  
155 the OES, discussion of the form of these traits refers to the EDJ, unless otherwise specified.

156 In addition to these four traits, unusual dentine horn morphologies were recorded when  
157 present and, although not formally scored, the presence of accessory cusps is noted.  
158 Intraobserver error was assessed through the rescoring of the entire sample by the primary  
159 observer (T.W.D.) more than one month after the initial scoring. Interobserver error was  
160 assessed through a second observer (M.M.S.) scoring a subset of 25 specimens for each trait.  
161 Inter- and intraobserver agreement was high, and errors appear to be random as they are not  
162 limited to specific taxa or trait types; results of the observer error tests can be found in SOM  
163 Table S3.

164 Transverse crest A number of authors have considered aspects of premolar transverse crest  
165 form at the OES (Suwa, 1990; Leonard and Hegmon, 1987; Bailey, 2002; Delezene and  
166 Kimbel, 2011, Irish et al., 2018). Some focussed on the prominence of the transverse crest  
167 (Suwa, 1990; Bailey, 2002; Irish et al., 2018), while others preferred to score the orientation  
168 of the transverse crest (Delezene and Kimbel, 2011). Leonard and Hegmon (1987) previously  
169 scored a number of *A. afarensis* P<sub>3</sub> specimens for ‘transverse ridge development’, using a  
170 five-point typology, which attempted to score both the form and prominence of the P<sub>3</sub>  
171 transverse crest. In a modern humans, Krenn et al. (2019) scored whether or not the transverse

172 crest is bifurcated, as well as whether it runs continuously between the two cusps or is  
173 interrupted.

174 When studying the hominoid transverse crest at the EDJ, it is apparent that the relationship  
175 between this crest and other crown structures, particularly the main premolar cusps, is highly  
176 variable. Therefore the typology used here focuses on the position of the transverse crest  
177 relative to other crown structures, and is based on the range of variation observed within the  
178 present sample. Unlike previous studies, the scoring system does not aim to characterize how  
179 strongly developed the transverse crest is (beyond presence/absence) because this is difficult  
180 to score objectively.

181 The scoring procedure consists of five discrete categories (although Types 1 and 1a are  
182 related) as follows (Fig. 1): Type 0 = transverse crest is absent, or only small incipient crests  
183 are present; Type 1 = transverse crest is present, and connects the protoconid to the metaconid  
184 (or equivalent point on the marginal ridge, when a clear metaconid is not present); Type 1a =  
185 transverse crest connects to the lingual margin of the tooth, but flattens to the level of the  
186 surrounding occlusal basin before connecting to the protoconid; Type 2 = transverse crest  
187 connects to the protoconid, but is either deflected distally or flattens to the level of the  
188 occlusal basin before connecting to the marginal ridge on the lingual side of the tooth; Type 3  
189 = transverse crest connects to the protoconid crest distal to the dentine horn tip and, as in  
190 Type 2, is either deflected distally or flattens to the level of the surrounding occlusal basin  
191 before connecting to the marginal ridge on the lingual side of the tooth.

192 In some specimens, particularly those with tall well-developed dentine horns, the  
193 transverse crest may appear not to reach the dentine horn tip or adjacent crest, being flattened  
194 to the level of the surrounding dentine such that the ridge is no longer visible. Here, the  
195 transverse crest was considered associated to the dentine horn provided that the crest is visible  
196 for at least two-thirds of the dentine horn height. Therefore, Type 1a is reserved for specimens

197 in which the transverse crest is flattened for more than one-third the height of the protoconid,  
198 as measured from the tip of the protoconid to the bottom of the occlusal basin. This distinction  
199 is illustrated in Figure 1 when comparing Neanderthal specimens KRP 51 (Type 1) with KRP  
200 D33 and KRP D114 (Type 1a). The distinction between Types 2 and 3, in the case of a partly  
201 flattened transverse crest, was made by judging whether the remaining portion of the crest is  
202 angled such that, were it to be present for the full length of the crown, it would make contact  
203 with the tip of the protoconid or with the adjacent crests. Specimens that have been digitally  
204 reconstructed due to tooth wear have been included in this analysis provided we could be  
205 confident of the form of the transverse crest, and provided the reconstructed portion makes up  
206 no more than 1/3 of the height of the dentine horn. The expression of any additional crests  
207 present in the occlusal basin of the tooth will also be discussed. It should be noted that  
208 although Type 1a is classed as a subtype of Type 1 (an explanation for this can be found in the  
209 discussion), we refer to these types separately, unless otherwise stated.

210 Marginal ridge continuity One difference between the P<sub>3</sub> of hominins and apes is that  
211 hominins typically have more strongly developed distal and mesial marginal ridges (Suwa,  
212 1990). However, even among hominins the prominence and form of the marginal ridge varies;  
213 this variation has been considered by a number of authors (Suwa, 1990; Ward et al., 2001). In  
214 particular, it has been noted that the mesial marginal ridge is often poorly developed in early  
215 hominins; such specimens are described as having an ‘open mesial fovea’ (Kimbel et al.,  
216 2006; Delezene and Kimbel, 2011).

217 Further, Sakai (1967) noted the presence of a ‘trigonid notch’ in the P<sub>3</sub> EDJ of modern  
218 humans, defined as a clear indentation in the mesial marginal ridge, and Krenn et al. (2019)  
219 scored modern human premolars for missing or reduced marginal ridges at the EDJ and OES.  
220 In our sample, specimens may exhibit a morphology resembling the trigonid notch described  
221 by Sakai (1967), or they may show poorly developed, or absent, marginal ridges on the mesial

222 or distal side of the crown. Since the difference between a ‘notched’ marginal ridge and one  
223 that is poorly developed may be very slight, the grading system used here instead scores  
224 marginal ridge continuity (or discontinuity), where a discontinuous marginal ridge is one  
225 which is flattened to the level of the occlusal basin for some portion of its length. This system  
226 therefore does not distinguish between absent, poorly developed and notched marginal ridges.  
227 Marginal ridge continuity was scored on the mesial and distal marginal ridges, according to  
228 the following system (Fig. 2): C = continuous marginal ridges; the distal marginal ridge runs  
229 from the distal protoconid crest to the metaconid (or equivalent point on the crown), and the  
230 mesial marginal ridge runs from this point, to the mesial protoconid crest; M = the mesial  
231 marginal ridge is discontinuous; D = the distal marginal ridge is discontinuous; MD = the  
232 mesial and distal marginal ridges are both discontinuous.

233 Marginal ridges may appear to flatten to the level of the occlusal basin at the base of the  
234 metaconid dentine horn, but this was only counted as discontinuous if the flattened section  
235 clearly extended further into the mesial or distal fovea than the base of the metaconid. This  
236 analysis was only completed on hominin specimens, excluding the extant apes, since the  
237 mesial and distal marginal ridges are generally poorly developed in extant apes.

238 Buccal grooves Kraus and Furr (1953:562), in their description of the morphology of the  
239 modern human P<sub>3</sub>, described the occasional presence of “a ridge and accompanying shallow  
240 vertical groove” on both the mesial and distal margins of the buccal face, but specified that  
241 these features are more often absent than present. Typically termed ‘buccal grooves’, this  
242 feature is argued to be particularly common in *A. africanus* (Robinson, 1956; Wood and  
243 Uytterschaut, 1987; Suwa, 1990). Suwa (1990) scored the degree of expression of buccal  
244 grooves in the modern human P<sub>3</sub> at the OES; distal buccal grooves were scored as ‘strong’,  
245 ‘moderate’ or ‘trace/lacking’, while a fourth category of ‘ill-defined but with a significant  
246 triangular depression’ was included for mesial buccal grooves. Similarly, Sakai (1967) scored

247 the presence of 'buccal ridges' at the EDJ in a sample of modern human P<sub>3</sub> using three  
248 categories: 'pronounced', 'weak' or 'no ridge'. Here specimens were scored using a similar  
249 system to these two studies, although the fourth category used by Suwa (1990) for mesial  
250 buccal grooves was not included such that specimens were scored using the same procedure  
251 for both mesial and distal EDJ buccal grooves (Fig. 3): 0 = absent; the EDJ buccal face shows  
252 no distinct grooves; there may be a slight vertical ridge on the mesial or distal margin of the  
253 buccal face, but it is not associated with a clear concavity; 1 = minor; a vertical ridge is  
254 present on the EDJ surface, and is associated with a small but distinct concavity; 2 = marked;  
255 the EDJ buccal surface shows a clear extended vertical ridge associated with a marked  
256 concavity.

257 It should be noted that buccal grooves are usually directly associated with either the  
258 protoconid crest or marginal ridges (often the intersection between the two); although, in  
259 some cases they are located closer to the cervix and may show little or no development higher  
260 up on the crown. We made no distinction between these types, and instead scored only how  
261 strongly developed the grooves are.

262 Cusp form and frequency The protoconid is universally present in the hominoid P<sub>3</sub> and, in the  
263 majority of cases, consists of a single raised, conic dentine horn. However, there are a limited  
264 number of cases in which the protoconid departs from this form, and these examples will be  
265 discussed. The presence of accessory cusps will be discussed briefly; however, premolar  
266 accessory cusps can often be very small at the EDJ, and for some specimens with a lower scan  
267 resolution and/or poor contrast between tissue types in the scan, the presence of accessory  
268 cusps at the EDJ is not always clear. The frequency of these cusps will therefore not be  
269 formally scored.

270

### 271 3. Results

#### 272 3.1 Transverse crest

273 The form of the transverse crest was assessed for 71 specimens using the typology outlined  
274 in Figure 2; the results are shown in Table 2 (full results can be found in SOM Table S1). The  
275 majority of these display a Type 1 transverse crest (65%; 46/71), however this type is rare  
276 among extant apes, where it is only seen in *Pongo*. *Hylobates* specimens uniformly have  
277 deflected transverse crests, with some connecting to the protoconid (Type 2) and others  
278 connecting to the distal protoconid crest (Type 3). The *Gorilla* P<sub>3</sub> also shows a deflected  
279 transverse crest, but connects to the protoconid (Type 2). All five *Pan* specimens display a  
280 deflected transverse crest (or one which otherwise fails to reach the lingual margin of the  
281 tooth); 4/5 of these are not angled towards the tip of the protoconid (Type 3). Type 3 is only  
282 present among the extant apes, and only one hominin specimen displays a Type 2 transverse  
283 crest. 78% (40/51) of hominin specimens display a transverse crest that runs from the  
284 protoconid to the metaconid (or the equivalent point on the marginal ridge; Type 1), and all  
285 hominin species represented here show a Type 1 transverse crest in at least one specimen.  
286 Type 1a, in which the transverse crest connects to the marginal ridge but does not reach the  
287 protoconid, is seen exclusively in Neanderthal specimens; 4/9 specimens display this type,  
288 while the remainder are Type 1.

289 A special case is the modern human specimen ULAC 790. This individual has poorly  
290 developed mesial and distal marginal ridges and no metaconid, making attribution to our  
291 typology difficult. In this specimen, the transverse crest extends lingually from the protoconid  
292 to meet a small ridge, which may or may not be a section of distal marginal ridge, but is very  
293 poorly developed (the main portion of the distal marginal ridge is situated on the distal edge  
294 of the crown, and does not connect to the transverse crest at any point).

295 A number of specimens display crests in addition to the main transverse crest (Fig. 4). For  
296 example, *A. africanus* and *P. robustus* frequently display small crests running distolingually  
297 towards the center of the distal fovea. These crests typically meet the transverse crest at the  
298 protoconid apex, although they may meet partway across the transverse crest and lingual to  
299 the protoconid (e.g., SK 100). Accessory crests are sometimes present on the lingual side of  
300 the tooth as well. For example, for DNH 46 a crest originates at the metaconid and runs  
301 towards the center of the distal fovea, while SK 62 displays a similar crest on the lingual side  
302 of the tooth that appears to originate at the distal marginal ridge. Moreover, Neanderthal  
303 specimens frequently display accessory crests, either mesial or distal to the main transverse  
304 crest, on the face of the tall protoconid crest (Fig. 4). These are variable in number, size and  
305 position, and are also seen, albeit less frequently, in modern humans.

306

### 307 3.2 *Marginal ridge continuity*

308 There were 69 hominin specimens for which the continuity of the marginal ridge was  
309 scored (Table 3; see full results in SOM Table S1). Of these, 70% (48/69) displayed a  
310 continuous marginal ridge (Fig. 2). With one exception, *A. anamensis*, all hominin species  
311 represented by three or more individuals have at least one specimen displaying a continuous  
312 marginal ridge. *Australopithecus anamensis* is unique among the hominin sample; all three  
313 specimens display a poorly developed mesial marginal ridge, scored as mesially  
314 discontinuous. Three *A. afarensis* specimens were scored, of which two (A.L. 266-1 and A.L.  
315 333w-1c) displayed a discontinuous mesial marginal ridge, and one (A.L. 333-10) displayed  
316 an entirely continuous marginal ridge. KNM-WT 8556 and W8-978 also display continuous  
317 marginal ridges.

318 *Australopithecus africanus* displays a range of marginal ridge forms, although the majority  
319 are continuous (nine continuous, two mesially discontinuous and one distally discontinuous).

320 STW 401 is the only example of an *Australopithecus* P<sub>3</sub> within this sample of a discontinuous  
321 distal marginal ridge. This specimen is also noteworthy because the form of this trait differs  
322 from that seen in later hominin specimens, and is different from that often seen on the mesial  
323 marginal ridge in *Australopithecus*. In the majority of cases discontinuities in the marginal  
324 ridge appear on either side of the metaconid; the marginal ridge lowers and flattens before  
325 reaching the dentine horn. In this case, however, there are two portions of the distal marginal  
326 ridge present that overlap one another, but do not meet. Continuous mesial and distal marginal  
327 ridges are seen in all specimens of *Paranthropus*, *H. naledi*, as well as the probable early  
328 *Homo* specimen SKX 21204 and are the most common form in *A. africanus*. In contrast, for  
329 *H. sapiens* (fossil and recent), half of the specimens in the sample display a discontinuous  
330 mesial and/or distal marginal ridge. None of the Neanderthal specimens in our sample show  
331 distal marginal ridge discontinuity, but 31% (4/13) show discontinuity in the mesial marginal  
332 ridge. The P<sub>3</sub> of the Mauer mandible also shows both mesial and distal discontinuity, while  
333 the Cave of Hearths P<sub>3</sub> shows mesial marginal ridge discontinuity. In some cases (e.g., ULAC  
334 171; Fig. 1b), the marginal ridge is mostly absent, with only small lingual deflections from the  
335 mesial and distal protoconid crests. Finally, this trait is not always consistent between  
336 antimeres—ULAC 58 displays a discontinuous mesial marginal ridge on the left P<sub>3</sub>, but not  
337 the right.

338

### 339 3.3 *Buccal grooves*

340 Buccal grooves were scored for 95 specimens (Table 4; see full results in SOM Table S1).  
341 Of these, 66% (63/95) show some level of buccal groove expression (mesial and/or distal).  
342 Buccal grooves are less common in the extant apes. In our sample, no extant ape specimen  
343 showed marked buccal grooves, mesial or distal; although all *Gorilla* and *Pan* specimens  
344 show minor distal buccal grooves (minor mesial buccal grooves are also present in half of



345 these specimens). Conversely, all 19 scorable *Australopithecus* specimens, as well as KNM-  
346 WT 8556 and W8-978, showed either minor or marked buccal grooves on both the distal and  
347 mesial sides. Further, in all *Australopithecus* specimens in which the mesial and distal buccal  
348 grooves are unequal, it is always the mesial buccal groove which is more strongly expressed.  
349 The opposite pattern is evident in *Paranthropus*, where the distal buccal grooves are generally  
350 better developed, and, in fact, mesial buccal grooves are absent in 86% (12/14) of  
351 *Paranthropus* specimens. In this respect, Omo specimen L427-7 is unusual for *Paranthropus*,  
352 as previously noted by Suwa (1990), as the P<sub>3</sub> displays marked mesial and distal buccal  
353 grooves. SXX 21204, a specimen attributed to early *Homo* (Grine, 1989), shows marked  
354 mesial and minor distal buccal grooves. *Homo naledi* specimens show minor or absent buccal  
355 grooves on both mesial and distal sides. Among modern humans and Neanderthals, buccal  
356 grooves are less common; no specimen showed marked mesial or distal buccal grooves.  
357 Neither the Mauer P<sub>3</sub> nor the Cave of Hearths P<sub>3</sub> show any buccal grooves, while 43% (6/14)  
358 Neanderthal and 86% (12/14) *H. sapiens* specimens exhibit no buccal grooves at all.  
359 Neanderthal specimens more often show minor buccal grooves on the mesial side (57%; 8/14)  
360 than on the distal side (21%; 3/14).

361

### 362 3.4 Protoconid form

363 The variation observed in the form of the protoconid is shown in Figure 5. The majority of  
364 specimens display a single, conic protoconid dentine horn tip, and as such, only the  
365 exceptions will be listed here (Fig. 5A). In Neanderthals and modern humans, the protoconid  
366 dentine horn tip may be less pronounced due to the presence of tall mesial and distal  
367 protoconid crests, and in one specimen, ULAC 790, there is no clearly differentiated  
368 protoconid tip, only a well-developed ridge (Fig. 5B). A small number of specimens display a  
369 protoconid with a longitudinally expanded tip (Fig. 5C). This feature is seen in two *H. naledi*

370 specimens, UW 101-144 and UW 101-889, and their probable antimeres (UW 101-506 and  
371 UW 101-377, respectively), as well as one *Pan* specimen, MPITC 11800. In some cases, such  
372 as UW 101-377, it is clear that the expanded dentine horn actually consists of two  
373 semidistinct tips. In other cases, the tip simply appears as a flattened ridge; however, since  
374 this structure is very small, it is possible that the resolution of the scans may be insufficient to  
375 discern two individual peaks. In one modern human, ULAC 58, the tip of the protoconid is  
376 transversely expanded (Fig. 5D). The protoconid crest meets the protoconid on the buccal side  
377 of the tip, while the transverse crest meets it on the lingual side of the tip, but these two points  
378 are not coincident, and are connected by a short ridge. This feature is present in both  
379 antimeres, but is not present in any other P<sub>3</sub> within our sample.

380

### 381 3.5 *Accessory cusps*

382 Poor tissue contrast in a number of specimens inhibits proper characterization of the  
383 frequency and detailed morphology of small accessory cusps. However, a number of general  
384 observations can be made. Many hominin specimens display small accessory cusps at  
385 multiple locations along the distal and mesial marginal ridges. There is commonly a small  
386 dentine horn at the distobuccal corner of the tooth, at the intersection between the distal  
387 marginal ridge and the distal protoconid crest. This cuspid can be seen in most hominin  
388 species, although it appears to be less common in modern humans and Neanderthals (issues of  
389 scan tissue contrast are less problematic in these specimens). It may be related to the presence  
390 of distal buccal grooves since the tip of the dentine horn is often contiguous with a raised  
391 ridge of dentine on the buccal face, while the concavity seen on the buccal face, immediately  
392 mesial to this ridge, is somewhat contiguous with the base of the dentine horn on the  
393 protoconid crest. A particularly pronounced example of this can be seen in STW 213 (Fig. 3).  
394 Accessory cusps are also found along the distal and mesial marginal ridges, and in some

395 cases, they can be nearly as large as the metaconid, as seen in L427-7. Other specimens, such  
396 as STW 151, display multiple accessory cusps, in this case along the distal marginal ridge.

397

## 398 4. Discussion

### 399 4.1 Transverse crest form

400 In the majority of hominins (78%; 40/51), the P<sub>3</sub> transverse crest extends lingually from  
401 the protoconid to meet the metaconid or equivalent point on the marginal ridge (Type 1; Fig.  
402 1). This is rare in extant apes. In *Hylobates* and *Gorilla*, the transverse crest does not typically  
403 reach the weakly developed marginal ridge, either ending mesial to the ridge, or deflecting  
404 distally. All *Pan* specimens studied here fit within Type 2 or 3, although the form of the  
405 transverse crest is quite variable in shape; some specimens have small associated accessory  
406 crests, while others are raised high above the crown basin or may have raised sections. Given  
407 the level of variability in even our small sample, further investigation is required. Such studies  
408 should also include other subspecies of *Pan troglodytes*, as well as *Pan paniscus*, as it is  
409 possible that the patterns observed here are specific to *Pan troglodytes verus*. *Pongo* is the  
410 exception among the apes; all specimens studied here display a Type 1 transverse crest,  
411 similar to the majority of hominins.

412 The earliest hominin in our sample, *A. anamensis*, displays a Type 1 transverse crest.  
413 Haile-Selassie et al. (2004:1505) suggested that this is also the case in an *Ardipithecus*  
414 *kadabba* P<sub>3</sub> from 5.6–5.8 Ma: “The transverse crest descends from the tip of the protoconid to  
415 the metaconid, which is hardly expressed as a distinct entity”. This would seem to indicate  
416 that that a Type 1 transverse crest is plesiomorphic for the hominin clade, and evolved  
417 independently in *Pongo*. However it is also possible that this state is plesiomorphic for  
418 hominids, but was lost in *Pan* and *Gorilla*. An analysis of the P<sub>3</sub> EDJ morphology in Miocene  
419 apes may help to shed further light on the evolutionary history of this trait. There is more

420 variation in the transverse crest in modern humans, where transverse crest absence is common  
421 (Type 0; 44%), and among our Neanderthal sample, in which Type 1a is relatively common  
422 (44%). In this form, the transverse crest flattens before reaching the protoconid tip. Here, it is  
423 possible that the relatively low transverse crest seen in Neanderthals is obscured due to the  
424 shape of the crown—in particular the tall, wide lingual face of the protoconid. If this were the  
425 case, then there would be no fundamental developmental difference between Types 1 and 1a;  
426 instead, the difference between the two would be due to a number of factors including the  
427 shape of the crown and the prominence of the transverse crest, which is why Type 1a may be  
428 best viewed as a subtype of Type 1. This would require further investigation, particularly  
429 concerning the developmental basis of the premolar transverse crest. Nonetheless, it appears  
430 that Type 1a, while not ubiquitous in Neanderthals, is an autapomorphy of this species.

431 Previous studies assessed the degree of transverse crest development at the OES (Suwa,  
432 1990; Bailey, 2002; Irish et al., 2018). While we do not score the degree of development of  
433 the crest, presence/absence frequencies can be generated using our scoring system, allowing  
434 direct comparisons. For example, Irish et al. (2018) find all *A. africanus* and *H. naledi*  
435 specimens, as well as a third of *P. robustus* specimens, have no transverse crest. Using  
436 overlapping fossil samples, we instead find that transverse crests are ubiquitous in all three  
437 species (Table 2), suggesting that the appearance of this trait differs fundamentally between  
438 the OES and EDJ. At the OES, the premolar transverse crest is often incised by the  
439 longitudinal fissure, which runs between the two cusps along the mesiodistal axis of the tooth.  
440 A deep longitudinal fissure therefore relates to a weak transverse crest at the OES. However, a  
441 number of crown features such as the development and placement of the metaconid, and the  
442 thickness of the enamel, are likely to play a part in the development of the longitudinal  
443 fissure, and therefore influence the form of the transverse crest observed. The longitudinal  
444 fissure is an enamel feature, however, and as such has no EDJ equivalent. This means that at

445 the EDJ, the transverse crest form is less dependent on the appearance of other crown  
446 features, which is advantageous when scoring multiple crown traits.

447 While hominins display relatively little variation in the form of the transverse crest, there is  
448 more variation in the expression of accessory crests (Fig. 4). These crests form in the occlusal  
449 basin either mesial or distal to the main transverse crest, and may connect to the protoconid  
450 crest, marginal ridge, either of the main cusps, or to other accessory crests. In hominin lower  
451 molars, multiple crests may form between the protoconid and metaconid; a variable feature  
452 called trigonid crest patterning (Wu and Turner, 1993; Skinner et al., 2008; Bailey et al.,  
453 2011; Martínez de Pinillos et al., 2014). Given the location of these crests, it is possible that  
454 these features have similar developmental origins to the transverse crest (and accessory crests)  
455 discussed here for hominin P<sub>3</sub>. Trigonid crests are particularly common in Neanderthal lower  
456 molars (Bailey, 2002, 2006), which is interesting given the high frequency of accessory crests  
457 found here (and previously noted by Bailey, 2006) for the Neanderthal P<sub>3</sub>. Martínez de  
458 Pinillos et al. (2014) found substantial molar trigonid crest variation in the Sima de los  
459 Huesos population at the EDJ, broadly equivalent to that of Neanderthals. Interestingly,  
460 Martínón-Torres et al. (2012) reported that a high proportion of Sima de los Huesos P<sub>3</sub>  
461 specimens show distal accessory crests at the OES, as well as pronounced transverse crests,  
462 when compared with modern humans. This could suggest that the same pattern may exist in  
463 the Sima de los Huesos population as we have found here for Neanderthals, although this  
464 would require comparing the EDJ morphologies of the two samples.

465 In the P<sub>3</sub>, accessory crests are frequently found in large, otherwise empty, areas of the EDJ  
466 occlusal basin, suggesting that the formation of accessory crests could be dependent on the  
467 space available on the crown. The P<sub>3</sub> in Neanderthals usually display a tall protoconid crest,  
468 which creates a steep, almost vertical, lingual-facing surface running from the protoconid  
469 crest to the bottom of the occlusal basin of the tooth. Accessory crests are frequently present

470 on this face in Neanderthals. *Australopithecus africanus* and *P. robustus* far more often show  
471 accessory crests that connect to the main dentine horns, which are often particularly large, or  
472 to the transverse crest itself, which is also well developed. Moreover, Kraus and Furr (1953)  
473 suggested that accessory crests are also found in the P<sub>3</sub> of modern humans. Given the level of  
474 variation seen in premolar accessory crests, as well as molar trigonid crests, it seems likely  
475 that these traits are not individually determined, but are instead the result of upstream  
476 developmental processes. There are a number of ways in which this could operate. Firstly, the  
477 formation of these crests could be genetically determined, but could be only able to form  
478 where there is sufficient space for them within the occlusal basin. In this case, accessory  
479 crests could develop through some of the same developmental processes as other crests and  
480 ridges on the tooth crown (such as the protoconid crest, transverse crest, and marginal ridges  
481 in premolars), but would presumably form later in development than the main crests, which  
482 would explain their variability as they would be dependent on a number of earlier forming  
483 features. This process would be analogous to the patterning cascade model of cusp  
484 development in which cusps form where there is space for them on the crown, and are  
485 prevented from forming too closely to each other by the presence of inhibitor proteins (Polly,  
486 1998; Jernvall, 2000; Kassai et al., 2005), with later forming cusps generally smaller and  
487 more variable than earlier forming cusps (Kondo and Townsend, 2006; Skinner and Gunz,  
488 2010). Crests are different from cusps in that they are often found in association with other  
489 crests. In fact, the accessory cusps identified here were invariably found to be associated with  
490 other crests or cusps on the tooth crown. However, these features appear to be common in the  
491 relatively large distal fovea of *A. africanus* and *P. robustus*, as well as along the tall  
492 protoconid crest of *H. neanderthalensis*, suggesting that the available space on the crown is  
493 important.

494       Alternatively, these crests could arise as the result of biomechanical forces during the  
495 development of the tooth crown. The EDJ preserves the form of the basement membrane of  
496 the inner enamel epithelium, the morphology of which is determined by folding driven by  
497 differential cell division in structures called enamel knots (Jernvall et al., 1994). Since the  
498 accessory crests are most common on relatively tall crown structures (dentine horns of *P.*  
499 *robustus*, and the protoconid crest of Neanderthals), it is possible that during the formation of  
500 these structures, the process of differential cell division creates small buckles and folds in the  
501 inner enamel epithelium as enamel knots do not proliferate themselves, but direct the cell  
502 proliferation of adjacent regions of the developing tooth. These small buckles and folds  
503 created could then go on to become the accessory ridges we see (see discussion of similar  
504 features in molars in Skinner et al, 2010). It is important to note that we did not find any  
505 accessory crests running parallel to the protoconid crest or the dentine horn tip; all run broadly  
506 towards the crest/ridge. In this case accessory crests may be developmentally distinct from the  
507 main crests and ridges of the tooth crown, which are far less variable within species.

508

#### 509   4.2   *Marginal ridge continuity*

510       In line with other studies that have considered the mesial marginal ridge at the OES  
511 (Leonard and Hegmon, 1987; Suwa et al., 1996; Ward et al., 2001; Kimbel et al., 2006;  
512 Delezene and Kimbel, 2011), the morphology of the EDJ supports a transformation series  
513 where a weak mesial marginal ridge, as in extant African apes and *A. anamensis*, is the  
514 primitive state for hominins and a strong mesial marginal ridge, as in *A. africanus* and  
515 *Paranthropus*, is derived. In *A. afarensis* Kimbel et al. (2004) highlighted ‘phylogenetic  
516 polymorphisms’, which refers to the observation that some character states are variable in *A.*  
517 *afarensis* but are fixed in the plesiomorphic condition in older hominins, like *A. anamensis*  
518 and *Ardipithecus ramidus*, and fixed in the apomorphic condition in younger hominins. The

519 form of the mesial marginal ridge observed at the EDJ in this study, and at the OES in other  
520 studies (Delezene and Kimbel, 2011), is congruent with such phylogenetic polymorphism.

521 Differences between our results and those from previous studies of the OES in early  
522 hominins are relatively minor. For example, while Suwa (1990) found that all *A. africanus*  
523 specimens show development of the lingual segment of the mesial marginal ridge, we found  
524 that two *A. africanus* specimens (STW 213 and Taung) have discontinuous P<sub>3</sub> mesial  
525 marginal ridges. However, this may be due to differences in scoring procedures, or in the  
526 fossil samples used, in the two studies (the Taung P<sub>3</sub>, for example, is unerupted and therefore  
527 would not have been included in previous studies), rather than fundamental differences  
528 between the EDJ and OES. The benefit of studying the EDJ is clearer in specimens that  
529 display marginal ridges that are mostly well developed but interrupted; they are flattened at  
530 some point along their length. This is most common in modern human and Neanderthal  
531 specimens where it is likely associated with the secondary reduction of the metaconid, and  
532 may reflect changing masticatory demands of the P<sub>3</sub> in these taxa.

533 Sakai (1967) recorded the presence of a ‘trigonid notch’ when looking at the EDJ of  
534 modern human P<sub>3</sub>, a feature which is equivalent to the mesial marginal ridge interruption  
535 noted here. They found the feature in roughly a quarter of specimens in their sample, but did  
536 not discuss any presence of a similar feature on the distal marginal ridge (although this feature  
537 is less common in our sample). This trait is much clearer at the EDJ than at the OES as the  
538 interruptions can be small, and are often located immediately next to the metaconid where the  
539 enamel of the cusp may obscure visibility of the interruption at the OES. In fact, Sakai (1967)  
540 only found the trigonid notch at the EDJ, and stated that the feature was entirely absent at the  
541 OES in all specimens. In some cases, the marginal ridge interruptions are often large enough  
542 that they are difficult to distinguish from the marginal ridge being entirely absent. Ultimately,  
543 we need a better understanding of the developmental processes that contribute to marginal



544 ridge formation in order to test whether these categories represent distinct traits, or the same  
545 trait with differing levels of expression. It does appear, however, that the interruption and  
546 absence of marginal ridges seen in modern humans and Neanderthals appears to represent a  
547 secondary loss and is not homologous with the minimal expression of the mesial marginal  
548 ridge in early *Australopithecus* and extant apes. Further, the frequent presence of interruptions  
549 in the distal, as well as mesial, marginal ridges suggests that this may be distinct from the  
550 character state we find in earlier taxa.

551 *Australopithecus africanus* specimen STW 401 is the only example within this sample of  
552 an interrupted distal marginal ridge in an *Australopithecus* P<sub>3</sub>, and is particularly interesting  
553 because the form of this trait appears to be different to that seen in later hominin specimens,  
554 and different to that often seen in the mesial marginal ridge in *Australopithecus*. In most  
555 cases, interruptions to the marginal ridge appear either side of the metaconid; the marginal  
556 ridge lowers and flattens before reaching the dentine horn. In this case, however, two portions  
557 of the distal marginal ridge overlap one another and do not meet. This appears to be a  
558 defective form of the marginal ridge in which extensions from the metaconid and protoconid  
559 crest have failed to meet and become continuous, possibly providing insights into the  
560 underlying developmental processes responsible for this structure around the occlusal basin.

561

#### 562 4.3 *Buccal grooves*

563 The grading system used here for buccal grooves (absent, minor, marked) is comparable to  
564 those used by previous authors at the OES (Suwa, 1990) and the EDJ (Sakai, 1967), and can  
565 also be compared to studies using presence/absence scores (Wood and Uytterschaut, 1987).  
566 Our results are largely consistent with those of the OES; while there are some differences,  
567 these may be due to the samples utilized in each case, which overlap but are not identical.

568 However, it is also possible that enamel deposition affects the OES expression of buccal  
569 grooves in some cases.

570 Suwa (1990) suggested that strong distal buccal grooves at the OES are a derived condition  
571 for hominins; this is supported by our finding that extant apes show absent or minor EDJ  
572 distal buccal grooves, while marked grooves are found in a number of *Australopithecus*  
573 specimens, including one specimen of *A. anamensis*, the earliest hominin in our sample.  
574 Further, ASK-VP-3/403, a P<sub>3</sub> of *Ar. kadabba* from 5.6–5.8 Ma, is also described as showing a  
575 distinct distal buccal groove at the OES (Haile-Selassie et al., 2004, 2009). On the mesial  
576 side, Suwa (1990) found that *A. afarensis* was variable, but suggested an increase in well-  
577 developed OES buccal grooves in the earliest hominins compared to apes. Our results are  
578 consistent with this suggestion, finding marked mesial buccal grooves in 5/6 *A. anamensis*  
579 and *A. afarensis* specimens, but not in any of our extant ape specimens. The variability in *A.*  
580 *afarensis* cannot be assessed here however due to the small available sample. In order to  
581 further test these suggestions, the EDJ morphology of fossil apes should also be investigated.

582 *Paranthropus* is derived among hominins in showing a high level of mesial buccal groove  
583 absence as noted previously (Wood and Uytterschaut, 1987; Suwa, 1990). However, we find  
584 more variability in the distal buccal groove expression than Suwa (1990). In modern humans,  
585 absence of mesial and distal buccal grooves is the most common form, while in Neanderthals,  
586 no specimens show marked buccal grooves, and 79% (11/14) show absent distal buccal  
587 grooves. However, Sakai (1967) found a higher proportion of modern human specimens with  
588 buccal grooves at the EDJ, particularly on the mesial side. This could be due to a difference in  
589 scoring procedure; however, it is also possible that buccal grooves are more common in the  
590 Japanese sample used by Sakai (1967) than in the sample used here. This would suggest that  
591 there is more variation in buccal groove expression in modern humans than is captured here,  
592 and should be explored further. The majority of *H. naledi* specimens show minor buccal

593 grooves on the mesial and distal sides, and only one specimen (U.W. 101-144) shows an  
594 absent distal buccal groove as suggested to be typical of early *Homo* at the OES (Wood and  
595 Uytterschaut, 1987; Suwa, 1990). The *H. naledi* expression of minor EDJ mesial buccal  
596 grooves could be interpreted as derived relative to the *Australopithecus*/early *Homo* state of  
597 strongly expressed mesial buccal grooves, particularly considering the weaker expression  
598 observed in modern humans and Neanderthals.

599 Although there are a number of species-specific patterns in buccal groove expression as  
600 outlined above, there is also a large amount of variation. Within our sample, it appears that  
601 buccal grooves are more common in specimens showing straight protoconid crests (in  
602 occlusal view). In such a configuration, there is a more angled intersection between the  
603 protoconid crests and the mesial/distal marginal ridge, often marked by a small accessory  
604 cusp. The buccal ridge is visible as a vertical crest on the buccal surface, as well as a slight  
605 concavity next to the ridge (towards the center of the crown), which could be considered an  
606 extension of the marginal ridge on the buccal surface. In this case, the expression of buccal  
607 grooves would depend on a number of aspects of crown morphology such as the overall shape  
608 of the crown, and the configuration of major occlusal crests, which may explain the variability  
609 in this trait. This should be explored further as it would affect our understanding of the  
610 independence of buccal grooves from other crown traits.

611 It is also important to consider potential serial homologies (de Beer, 1971; Roth, 1994)  
612 with similar traits on other tooth positions. Based on their analysis of the EDJ of mandibular  
613 molars, Skinner et al. (2009) suggested that crest features along the entire buccal face should  
614 be considered part of the expression of the protostylid; it is possible that these crests on the  
615 molar buccal face are developmentally linked to P<sub>3</sub> buccal grooves (note that the name buccal  
616 ‘groove’ may be misleading; in most cases at the EDJ the buccal groove consists of a  
617 ridge/crest with an associated concavity). The pattern noted here in which *A. africanus*

618 specimens show stronger mesial buccal grooves than *P. robustus* is somewhat mirrored in  
619 Skinner et al.'s (2009) study of molar protostylid patterning. They found that *A. africanus*  
620 molars expressed a protostylid crest that extends mesially, whereas the crests in *P. robustus*  
621 specimens were mostly restricted to the area between the protoconid and hypoconid. Further,  
622 we find that STW 213 (Fig. 3, bottom image) exhibits strong buccal grooves in addition to  
623 protostylid-like crests running diagonally across the buccal face. In this case, the buccal  
624 groove and possible protostylid appear to be distinct features; however, it is interesting to note  
625 that STW 213 has the most well defined buccal grooves of any specimen within the sample.  
626 Ultimately, further investigation is required to assess the developmental basis of both of these  
627 traits.

628

#### 629 4.4 *Protoconid form*

630 The transversely expanded dentine horn seen in ULAC 58 (Fig. 5D) may be related to the  
631 'internally placed cusps' identified at the OES of Neanderthal molars (Tattersall and  
632 Schwartz, 1999; Bailey, 2004). Further, Martin et al. (2017) discussed the presence of  
633 'centrally placed dentine horn tips' at the EDJ of Neanderthal and modern human molars, and  
634 the protoconid dentine horn form in the P<sub>3</sub> of ULAC 58 appears to fit within their typology. In  
635 fact, the molars of the ULAC 58 mandible were also included in the sample for Martin et al.  
636 (2017), where they found that the M<sub>1</sub> and M<sub>3</sub> displayed a centrally placed entoconid dentine  
637 horn. Martin et al. (2017) suggested that centrally located dentine horns were particularly  
638 common in Neanderthals. Although we did not find any Neanderthal specimens for which the  
639 protoconid was similar in form to that found in ULAC 58, we did find that the apex of the  
640 protoconid crest and the tip of the protoconid are frequently angled lingually, resulting in a  
641 more centrally located protoconid. Gómez-Robles et al. (2008) suggested that this feature is  
642 distinctive of the P<sub>3</sub> in Neanderthals and *H. heidelbergensis*. Unfortunately, the Mauer P<sub>3</sub>

643 included here (and its antimere) is too worn to assess if this specimen displays the same  
644 morphology.

645 Another trait discussed here, the longitudinally expanded dentine horn (Fig. 5C), also  
646 relates to a feature discussed by Martin et al. (2017): twinned dentine horns. We did not find  
647 this trait in any modern human or Neanderthal specimens, but it was found in several *H.*  
648 *naledi* specimens, as well as a single *Pan* specimen. In some cases, the *H. naledi* P<sub>3</sub>  
649 protoconid appears simply to be expanded, rather than twinned, although it is possible that the  
650 separate apices of the twinned dentine horns are too small to be visible in the scans. Martin et  
651 al. (2017) also found specimens that showed ‘unusually wide’ dentine horns, and suggested  
652 this may be a diminutive form of the twinned dentine horn trait. These traits are particularly  
653 interesting since they are difficult to reconcile with the currently well-accepted patterning  
654 cascade model of cuspal development (Polly, 1998; Jernvall, 2000; Kondo and Townsend,  
655 2006; Skinner and Gunz, 2010), in which cusps develop iteratively across the crown, and that  
656 a zone of inhibition during crown development prevents the formation of cusps in close  
657 proximity to one another. Neither of the protoconid traits described here were seen on P<sub>3</sub>  
658 metaconids, although this may be due a combination of the rarity of the features, and the  
659 reduced number of specimens displaying a well-developed metaconid. Although more  
660 investigation is warranted before strong conclusions can be made, the twinned protoconid  
661 dentine horns of *H. naledi* represent a potential autapomorphy of this species.

662

## 663 **5. Taxonomic implications**

664 These traits can have a bearing on the taxonomic affiliation of important specimens. For  
665 example, two specimens included here, KNM-WT 8556 and W8-978, have not been  
666 definitively assigned to species rank, but have been suggested to belong to *A. afarensis*  
667 (Leonard and Hegmon, 1987; Suwa, 1990; Brown et al., 2001). Our results are consistent with

668 this suggestion; both specimens show fully continuous marginal ridges, a morphology that  
669 first appears in *A. afarensis*, and the buccal groove forms are typical of *Australopithecus* in  
670 general. However, these specimens are also roughly coeval with *Kenyanthropus platyops* and  
671 *Australopithecus deyiremeda*, neither of which are included here. In fact, KNM-WT 8556 is  
672 found in the same Lomekwi locality as *K. platyops*, and the P<sub>4</sub> and M<sub>3</sub> are suggested to show  
673 a derived morphology, relative to *A. afarensis* (Leakey et al., 2001). Ultimately however,  
674 additional fossil material of *K. platyops* is required in order to make direct comparisons to  
675 mandibular specimens.

676 KNM-ER 5431 has been previously assigned to *A. afarensis* (Leonard and Hegmon, 1987),  
677 while other authors have suggested the presence of *Homo*-like traits (Suwa, 1990; Wood,  
678 1991). The P<sub>3</sub> included here, KNM-ER 5431E (Fig 1i), is found to display continuous  
679 marginal ridges, which are common in a number of hominin taxa, while the buccal groove  
680 form was not able to be assessed here due to poor tissue contrast in the cervical region of the  
681 crown (although clear buccal grooves are evident at the OES). The transverse crest form of  
682 this specimen (Type 2) is not seen in any other hominin P<sub>3</sub> in our sample; the crest is  
683 deflected before reaching the metaconid. It is unclear whether this is an apomorphic character  
684 state for a species not included in this sample, or whether this type is simply an uncommon  
685 developmental variant. Since the transverse crest in this specimen is weakly expressed, this  
686 may be difficult to assess from the OES, underlining the importance of trait assessment at the  
687 EDJ as well as the OES.

688 STW 151 has been suggested to display a number of derived features relative to  
689 Sterkfontein *A. africanus* by Moggi-Cecchi et al. (1998). However, the P<sub>3</sub> was not suggested  
690 to show any derived *Homo* discrete traits at the OES, which is supported in our EDJ results. In  
691 general, we did not find clear differences between *A. africanus* and our small sample of early  
692 *Homo* specimens, except for the absence of a distal buccal groove in KNM-ER 806E, which

693 was not seen in any *Australopithecus* specimen. SKX 21204 is assigned to *Homo*, but has no  
694 specific attribution (Grine, 1989). Unfortunately, the small early *Homo* sample here prevents a  
695 detailed assessment of the P<sub>3</sub> of this specimen.

696 The Cave of Hearths mandible has not been assigned to a species, but the original  
697 description and later analyses drew comparisons with the morphology seen in Neanderthals,  
698 and recently Berger et al. (2017) suggested the need for comparisons with *H. naledi*. Although  
699 the Cave of Hearths mandible is poorly dated, it is suggested to antedate the *H. naledi* sample  
700 included here (McNabb et al., 2009; Dirks et al., 2017). We find that the discontinuous mesial  
701 marginal ridge and absence of buccal grooves distinguishes the Cave of Hearths P<sub>3</sub> from *H.*  
702 *naledi*. However, the absent buccal grooves and marginal ridge discontinuity of the P<sub>3</sub> are  
703 common in modern humans and Neanderthals, and are also found in the Mauer P<sub>3</sub>. This,  
704 combined with the overall EDJ shape of the P<sub>3</sub> (Davies et al., 2019), suggests that the Cave of  
705 Hearths specimen is more closely associated with modern humans, Neanderthals and  
706 specimens attributed to *H. heidelbergensis*, while *H. naledi*, despite its Middle Pleistocene  
707 age, displays a morphology which is more primitive. This is also supported by differences  
708 between *H. naledi* and the Cave of Hearths mandible in occlusal topography of the M<sub>2</sub>  
709 (Berthaume et al., 2018) and M<sub>1</sub> root morphology (Kupczik et al., 2019).

710

## 711 **6. Conclusions**

712 We have identified several discrete P<sub>3</sub> traits evident at the EDJ that are variable in  
713 hominoids. In some cases, the form of these traits is largely similar to that of the OES, such as  
714 in buccal grooves, but in other cases, such as the transverse crest, additional morphological  
715 information is evident at the EDJ that may be obscured at the OES by adjacent crown features  
716 and thick enamel. A number of trends are evident in the expression of these traits. For  
717 example, the transverse crest is variable in extant apes, but the majority of hominins show a

718 single crest connecting the protoconid to the metaconid. Absence of this crest is seen in some  
719 specimens of *P. boisei* and *H. sapiens*, while some Neanderthals show a derived form in  
720 which the crest flattens before reaching the protoconid cusp tip. The mesial marginal ridge  
721 undergoes a transformation series early in hominin evolution, from the poorly developed  
722 ancestral state seen in *A. anamensis*, through the polymorphic condition in *A. afarensis*  
723 (Kimbel et al., 2004; Delezene and Kimbel, 2011), to the well-developed marginal ridges seen  
724 in *A. africanus* and *P. robustus*. A number of modern humans and Neanderthal specimens  
725 show a secondary reduction in marginal ridge development, although this is confined to the  
726 mesial marginal ridge in Neanderthals. Buccal grooves are variable throughout hominoids,  
727 with some taxon-specific patterns, but which may be dependent on other aspects of crown  
728 morphology.

729 Our understanding of the development of discrete dental traits is very limited. This is  
730 important when using these traits in phylogenetic studies since the traits used should ideally  
731 be genetically independent, and care must be taken to avoid suggesting that traits that are  
732 superficially similar, but developmentally distinct, can be considered as identical (i.e.,  
733 versions of the same trait) for cladistics analysis. Further, patterning cascade models to  
734 explain variation in dental form have mostly focused on the morphology of the molars.  
735 Similar homologous mechanisms presumably underlie the differences in premolar form;  
736 however, certain traits (i.e., twinned dentine horns) observed in this study are hard to explain  
737 with such a model and highlight the need for investigation of premolar form within an  
738 evolutionary-developmental framework. As shown here, studying the EDJ provides valuable  
739 insights into the development of these traits, and this will be further improved through  
740 inclusion of broader primate samples, and the study of other tooth positions at the EDJ.

741



742 **References**

- 743 Anemone, R.L., Skinner, M.M., Dirks, W., 2012. Are there two distinct types of hypocone in  
744 Eocene primates? The 'pseudohypocone' of notharctines revisited. *Palaeontologia*  
745 *Electronica* 15, 26A.
- 746 Bailey, S.E., 2002. Neandertal dental morphology: implications for modern human origins.  
747 Ph.D. Dissertation, Arizona State University.
- 748 Bailey, S.E., 2004. A morphometric analysis of maxillary molar crowns of Middle-Late  
749 Pleistocene hominins. *Journal of Human Evolution* 47, 183-198.
- 750 Bailey, S.E., 2006. Beyond shovel shaped incisors: Neandertal dental morphology in a  
751 comparative context. *Periodicum Biologorum* 108, 253–267.
- 752 Bailey, S.E., Skinner, M.M., Hublin, J., 2011. What lies beneath? An evaluation of lower  
753 molar trigonid crest patterns based on both dentine and enamel expression. *American*  
754 *Journal of Physical Anthropology* 145, 505-518.
- 755 Berger, L.R., Hawks, J., Dirks, P.H., Elliott, M., Roberts, E.M., 2017. *Homo naledi* and  
756 Pleistocene hominin evolution in subequatorial Africa. *eLife* 6, e24234.
- 757 Berthaume, M.A., Delezene, L.K., Kupczik, K. 2018. Dental topography and the diet of  
758 *Homo naledi*. *Journal of Human Evolution* 118, 14-26.
- 759 Beynon, A.D., Wood, B.A., 1986. Variations in enamel thickness and structure in East  
760 African hominids. *American Journal of Physical Anthropology* 70, 177-193.
- 761 Braga, J., Thackeray, J.F., Subsol, G., Kahn, J.L., Maret, D., Treil, J. and Beck, A., 2010. The  
762 enamel–dentine junction in the postcanine dentition of *Australopithecus africanus*:  
763 intra-individual metameric and antimeric variation. *Journal of Anatomy* 216, 62-79.

- 764 Brown, B., Brown, F.H., Walker, A., 2001. New hominids from the Lake Turkana Basin,  
765 Kenya. *Journal of Human Evolution* 41, 29-44.
- 766 Brunet, M., Guy, F., Pilbeam, D., Mackaye, H.T., Likius, A., Ahounta, D., Beauvilain, A.,  
767 Blondel, C., Bocherens, H., Boisserie, J. De Bonis, L., Coppens, Y., Dejax, J., Denys, C.,  
768 Douring, P., Eisenmann, V., Fanone, G., Fronty, P., Geraads, D., Lehmann, T., Lihoreau,  
769 F., Louchart, A., Mahamat, A., Merceron, G., Mouchelin, G., Otero, O., Campomanes,  
770 P.P., De Leon, M.P., Rage, J., Sapanet, M., Schuster, M., Sudre, J., Tassy, P., Valentin, X.,  
771 Vignaud, P., Viriot, L., Zazzo, A., Zollikofer, C., 2002. A new hominid from the Upper  
772 Miocene of Chad, Central Africa. *Nature* 418, 145-151.
- 773 Burnett, S.E., Irish, J.D., Fong, M.R., 2013. Wear's the problem? Examining the effect of  
774 dental wear on studies of crown morphology. In: Scott, G.R., Irish, J.D. (Eds.),  
775 *Anthropological Perspectives on Tooth Morphology: Genetics, Evolution, Variation*.  
776 Cambridge University Press, Cambridge, pp. 535-554.
- 777 Corruccini, R.S., 1987. The dentinoenamel junction in primates. *International Journal of*  
778 *Primatology* 8, 99-114
- 779 de Beer, S.G., 1971. *Homology, an Unsolved Problem*. Oxford University Press, London.
- 780 Deleuzene, L.K., 2015. Modularity of the anthropoid dentition: Implications for the evolution  
781 of the hominin canine honing complex. *Journal of Human Evolution* 86, 1-12.
- 782 Deleuzene, L.K., Kimbel, W.H., 2011. Evolution of the mandibular third premolar crown in  
783 early *Australopithecus*. *Journal of Human Evolution* 60, 711-730.
- 784 Dempsey, P.A. and Townsend, G.C., 2001. Genetic and environmental contributions to  
785 variation in human tooth size. *Heredity* 86, 685-693.

786 Dirks, P.H., Roberts, E.M., Hilbert-Wolf, H., Kramers, J.D., Hawks, J., Dosseto, A., Duval,  
787 M., Elliott, M., Evans, M., Grün, R., Hellstrom, J., 2017. The age of *Homo naledi* and  
788 associated sediments in the Rising Star Cave, South Africa. *eLife* 6, e24231.

789 Gómez-Robles, A., Martínón-Torres, M., Bermúdez de Castro, J.M., Prado, L., Sarmiento, S.,  
790 Arsuaga, J.L., 2008. Geometric morphometric analysis of the crown morphology of the  
791 lower first premolar of hominins, with special attention to Pleistocene *Homo*. *Journal of*  
792 *Human Evolution* 55, 627-638.

793 Greenfield, L.O., 1990. Canine “honing” in *Australopithecus afarensis*. *American Journal of*  
794 *Physical Anthropology* 82, 135-143.

795 Greenfield, L.O., Washburn, A., 1992. Polymorphic aspects of male anthropoid honing  
796 premolars. *American Journal of Physical Anthropology* 87, 173-186.

797 Grine, F.E., 1989. New hominid fossils from the Swartkrans Formation (1979–1986  
798 excavations): craniodental specimens. *American Journal of Physical Anthropology* 79,  
799 409-449.

800 Grine, F.E., Sponheimer, M., Ungar, P.S., Lee-Thorp, J., Teaford, M.F., 2012. Dental  
801 microwear and stable isotopes inform the paleoecology of extinct hominins. *American*  
802 *Journal of Physical Anthropology* 148, 285-317.

803 Guy, F., Lazzari, V., Gilissen, E., Thiery, G., 2015. To what extent is primate second molar  
804 enamel occlusal morphology shaped by the enamel-dentine junction? *PLoS One* 10,  
805 e0138802.

806 Haile-Selassie, Y., Suwa, G., White, T.D., 2009. Hominidae. In: Haile-Selassie Y.,  
807 WoldeGabriel G., (Eds.), *Ardipithecus kadabba*. Evidence from the Late Miocene of  
808 Africa. University of California Press, Berkeley and Los Angeles, pp.159-236.

809 Haile-Selassie, Y., Suwa, G., White, T.D. 2004. Late Miocene teeth from Middle Awash,  
810 Ethiopia, and early hominid dental evolution. *Science* 303, 1503-1505.

811 Hlusko, L.J., Mahaney, M.C. 2003. Genetic contributions to expression of the baboon  
812 cingular remnant. *Archives of Oral Biology* 48, 663-672.

813 Irish, J.D., Guatelli-Steinberg, D., 2003. Ancient teeth and modern human origins: an  
814 expanded comparison of African Plio-Pleistocene and recent world dental samples. *Journal*  
815 *of Human Evolution* 45, 113-144.

816 Irish, J.D., Bailey, S.E., Guatelli-Steinberg, D., Deleuzene, L.K., Berger, L.R., 2018. Ancient  
817 teeth, phenetic affinities, and African hominins: Another look at where *Homo naledi* fits in.  
818 *Journal of Human Evolution* 122, 108-123

819 Jernvall, J., 2000. Linking development with generation of novelty in mammalian teeth.  
820 *Proceedings of the National Academy of Sciences USA* 97, 2641-2645.

821 Jernvall, J., Kettunen, P., Karavanova, I., Martin, L.B., Thesleff, I., 1994. Evidence for the  
822 role of the enamel knot as a control center in mammalian tooth cusp formation: non-  
823 dividing cells express growth stimulating Fgf-4 gene. *The International Journal of*  
824 *Developmental Biology* 38, 463-469.

825 Kassai, Y., Munne, P., Hotta, Y., Penttilä, E., Kavanagh, K., Ohbayashi, N., Takada, S.,  
826 Thesleff, I., Jernvall, J., Itoh, N., 2005. Regulation of mammalian tooth cusp patterning by  
827 ectodin. *Science* 309, 2067-2070.

828 Kettunen, P., Thesleff, I. 1998. Expression and function of FGFs-4, -8, and -9 suggest  
829 functional redundancy and repetitive use as epithelial signals during tooth morphogenesis.  
830 *Developmental Dynamics* 211, 256-268.

831 Kimbel, W.H., Rak, Y., Johanson, D.C., 2004. *The Skull of Australopithecus afarensis*.  
832 Oxford University Press, New York.

833 Kimbel, W.H., Lockwood, C.A., Ward, C.V., Leakey, M.G., Rak, Y., Johanson, D.C., 2006.  
834 *Was Australopithecus anamensis* ancestral to *A. afarensis*? A case of anagenesis in the  
835 hominin fossil record. *Journal of Human Evolution* 51, 134-152.

836 Kondo, S., Townsend, G.C., 2006. Associations between Carabelli trait and cusp areas in  
837 human permanent maxillary first molars. *American Journal of Physical Anthropology* 129,  
838 196-203.

839 Korenhof, C.A.W., 1960. *Morphogenetical Aspects of the Human Upper Molar*.  
840 Uitgeversmaatschappij Neerlandia, Utrecht.

841 Kraus, B.S., Furr, M.L., 1953. Lower first premolars. Part I. A definition and classification of  
842 discrete morphologic traits. *Journal of Dental Research* 32, 554-564.

843 Krenn, V.A., Fornai, C., Wurm, L., Bookstein, F.L., Haeusler, M., Weber, G.W., 2019.  
844 Variation of 3D outer and inner crown morphology in modern human mandibular  
845 premolars. *American Journal of Physical Anthropology* 168, 646-663.

846 Kupczik, K., Delezene, L.K., Skinner, M.M. 2019. Mandibular molar root and pulp cavity  
847 morphology in *Homo naledi* and other Plio-Pleistocene hominins. *Journal of Human*  
848 *Evolution* 130, 83-95.

849 Leakey, M.G., Spoor, F., Brown, F.H., Gathogo, P.N., Kiarie, C., Leakey, L.N., McDougall,  
850 I., 2001. New hominin genus from eastern Africa shows diverse middle Pliocene lineages.  
851 Nature 410, 433-440.

852 Lee-Thorp, J.A., Sponheimer, M., van der Merwe, N.J., 2003. What do stable isotopes tell us  
853 about hominid dietary and ecological niches in the Pliocene? International Journal of  
854 Osteoarchaeology 13, 104-113.

855 Leonard, W.R., Hegmon, M., 1987. Evolution of P<sub>3</sub> morphology in *Australopithecus*  
856 *afarensis*. American Journal of Physical Anthropology 73, 41-63

857 Liao, W., Xing, S., Li, D., Martínón-Torres, M., Wu, X., Soligo, C., de Castro, J.M.B., Wang,  
858 W., Liu, W., 2019. Mosaic dental morphology in a terminal Pleistocene hominin from  
859 Dushan Cave in southern China. Scientific Reports 9, 2347.

860 Liu, W., Schepartz, L.A., Xing, S., Miller-Antonio, S., Wu, X., Trinkaus, E., Martínón-  
861 Torres, M., 2013. Late middle pleistocene hominin teeth from Panxian Dadong, South  
862 China. Journal of Human Evolution 64, 337-355.

863 Macho, G.A., Thackeray, J.F., 1992. Computed tomography and enamel thickness of  
864 maxillary molars of Plio-Pleistocene hominids from Sterkfontein, Swartkrans, and  
865 Kromdraai (South Africa): An exploratory study. American Journal of Physical  
866 Anthropology 89, 133-143.

867 Martin, R.M., Hublin, J.-J., Gunz, P., Skinner, M.M., 2017. The morphology of the enamel-  
868 dentine junction in Neanderthal molars: Gross morphology, non-metric traits, and temporal  
869 trends. Journal of Human Evolution 103, 20-44.

870 Martínez de Pinillos, M., Martínón-Torres, M., Skinner, M.M., Arsuaga, J.L., Gracia-Téllez,  
871 A., Martínez, I., Martín-Francés, L., Bermúdez de Castro, J.M., 2014. Trigonid crests  
872 expression in Atapuerca-Sima de los Huesos lower molars: Internal and external  
873 morphological expression and evolutionary inferences. *Comptes Rendus Palevol* 13, 205-  
874 221.

875 Martínón-Torres, M., Bermúdez de Castro, J.M., Gómez-Robles, A., Prado-Simón, L.,  
876 Arsuaga, J.L., 2012. Morphological description and comparison of the dental remains from  
877 Atapuerca-Sima de los Huesos site (Spain). *Journal of Human Evolution* 62, 7-58.

878 Martínón-Torres, M., de Pinillos, M.M., Skinner, M.M., Martín-Francés, L., Gracia-Téllez,  
879 A., Martínez, I., Arsuaga, J.L., Bermúdez de Castro, J.M., 2014. Talonid crests expression  
880 at the enamel–dentine junction of hominin lower permanent and deciduous molars.  
881 *Comptes Rendus Palevol* 13, 223-234.

882 McNabb, J., Sinclair, A., Wadley, L., Maguire, J., Latham, A., Herries, A., Ogola, C., Curnoe,  
883 D., Underhill, D. 2009. *The Cave of Hearths: Makapan Middle Pleistocene Research*  
884 *Project: Field Research by Anthony Sinclair and Patrick Quinney, 1996-2001.*  
885 Archaeopress, Oxford.

886 Moggi-Cecchi, J., Tobias, P.V., Beynon, A.D., 1998. The mixed dentition and associated  
887 skull fragments of a juvenile fossil hominid from Sterkfontein, South Africa. *American*  
888 *Journal of Physical Anthropology* 106, 425-465.

889 Morita, W., Yano, W., Nagaoka, T., Abe, M., Ohshima, H., Nakatsukasa, M. 2014. Patterns  
890 of morphological variation in enamel–dentin junction and outer enamel surface of human  
891 molars. *Journal of Anatomy* 224, 669-680.

- 892 Nager, G., 1960. Der vergleich zwischen dem räumlichen verhalten des dentin-kronenreliefs  
893 und dem schmelzrelief der zahnkrone. *Cells Tissues Organs* 42, 226-250.
- 894 Ortiz, A., Skinner, M.M., Bailey, S.E., Hublin, J.-J., 2012. Carabelli's trait revisited: An  
895 examination of mesiolingual features at the enamel–dentine junction and enamel surface of  
896 *Pan* and *Homo sapiens* upper molars. *Journal of Human Evolution* 63, 586-596.
- 897 Polly, P.D., 1998. Variability, selection, and constraints: development and evolution in  
898 viverravid (Carnivora, Mammalia) molar morphology. *Paleobiology* 24, 409-429.
- 899 Richards, M.P., Pettitt, P.B., Stiner, M.C., Trinkaus, E., 2001. Stable isotope evidence for  
900 increasing dietary breadth in the European mid-Upper Paleolithic. *Proceedings of the*  
901 *National Academy of Sciences USA* 98, 6528-6532.
- 902 Robinson, J.T., 1956. *The Dentition of the Australopithecinae*. Transvaal Museum, Pretoria.
- 903 Roth, V.L., 1994. Within and Between Organisms: Replicators, Lineages, and Homologues.  
904 In: Hall, B.K., (Ed.), *Homology: The Hierarchical Basis of Comparative Biology*.  
905 Academic Press, San Diego, pp 301–337
- 906 Sakai, T., 1967. Morphologic study of the dentinoenamel junction of the mandibular first  
907 premolar. *Journal Dental Research* 46, 927-932
- 908 Schulze, M.A., Pearce, J.A., 1994. A morphology-based filter structure for edge-enhancing  
909 smoothing. In: *Proceedings of the 1994 IEEE International Conference on Image*  
910 *Processing*, pp. 530–534
- 911 Skinner, M.M., 2008. Enamel-dentine junction morphology of extant hominoid and fossil  
912 hominin mandibular molars. Ph.D. Dissertation, George Washington University.



913 Skinner, M.M., Gunz, P., 2010. The presence of accessory cusps in chimpanzee lower molars  
914 is consistent with a patterning cascade model of development. *Journal of Anatomy* 217,  
915 245-253.

916 Skinner, M.M., Wood, B.A., Boesch, C., Olejniczak, A.J., Rosas, A., Smith, T.M., Hublin,  
917 J.J., 2008. Dental trait expression at the enamel-dentine junction of lower molars in extant  
918 and fossil hominoids. *Journal of Human Evolution* 54, 173-186.

919 Skinner, M.M., Wood, B.A., Hublin, J.-J., 2009. Protostylid expression at the enamel-dentine  
920 junction and enamel surface of mandibular molars of *Paranthropus robustus* and  
921 *Australopithecus africanus*. *Journal of Human Evolution* 56, 76-85.

922 Skinner, M.M., Evans, A., Smith, T., Jernvall, J., Tafforeau, P., Kupczik, K., Olejniczak, A.J.,  
923 Rosas, A., Radovčić, J., Thackeray, J.F., Toussaint, M., Hublin, J.-J., 2010. Brief  
924 communication: Contributions of enamel-dentine junction shape and enamel deposition to  
925 primate molar crown complexity. *American Journal of Physical Anthropology* 142, 157-  
926 163.

927 Smith, T.M., Olejniczak, A.J., Zermeno, J.P., Tafforeau, P., Skinner, M.M., Hoffmann, A.,  
928 Radovčić, J., Toussaint, M., Kruszynski, R., and Menter, C., 2012. Variation in enamel  
929 thickness within the genus *Homo*. *Journal of Human Evolution* 62, 395-411.

930 Sponheimer, M., Lee-Thorp, J.A., 1999. Isotopic evidence for the diet of an early hominid,  
931 *Australopithecus africanus*. *Science* 283, 368-370.

932 Suwa, G., 1988. Evolution of the “robust” australopithecines in the Omo succession: evidence  
933 from mandibular premolar morphology. In: Grine, F.E. (Ed.), *Evolutionary History of the*  
934 *“Robust” Australopithecines*. Aldine de Gruyter, New York, pp. 199-222.

- 935 Suwa, G., 1990. A comparative analysis of hominid dental remains from the Shungura and  
936 Usno Formations, Omo valley, Ethiopia. Ph.D. Dissertation, University of California,  
937 Berkeley.
- 938 Suwa, G., White, T.D., Howell, F.C., 1996. Mandibular postcanine dentition from the  
939 Shungura Formation, Ethiopia: crown morphology, taxonomic allocations, and Plio-  
940 Pleistocene hominid evolution. *American Journal of Physical Anthropology* 101, 247-282.
- 941 Suwa, G., Kono, R.T., Simpson, S.W., Asfaw, B., Lovejoy, C.O., White, T.D., 2009.  
942 Paleobiological implications of the *Ardipithecus ramidus* dentition. *Science* 326, 69-99.
- 943 Tattersall, I, Schwartz, J.H., 1999. Hominids and hybrids: The place of Neanderthals in  
944 human evolution. *Proceedings of the National Academy of Sciences USA* 96, 7117-7119.
- 945 Townsend, G.C., Martin, N.G. 1992. Fitting genetic models to Carabelli trait data in South  
946 Australian twins. *Journal of Dental Research* 71, 403-409.
- 947 Visualization Sciences Group, 2010. AVIZO, Version 6.3. Visualization Sciences Group,  
948 Bordeaux.
- 949 Walker, A., 1984. Mechanisms of honing in the male baboon canine. *American Journal of*  
950 *Physical Anthropology* 65, 47-60.
- 951 Walker, A., Hoeck, H.N., Perez, L., 1978. Microwear of mammalian teeth as an indicator of  
952 diet. *Science* 201, 908-910.
- 953 Ward, C.V., Leakey, M.G., Walker, A., 2001. Morphology of *Australopithecus anamensis*  
954 from Kanapoi and Allia Bay, Kenya. *Journal of Human Evolution* 41, 255-368.

- 955 Wollny, G., Kellman, P., Ledesma-Carbayo, M.J., Skinner, M.M., Hublin, J.-J., Hierl, T.,  
956 2013. MIA-A free and open source software for gray scale medical image analysis. Source  
957 Code for Biology and Medicine 8, 20.
- 958 Wood, B.A., 1991. Koobi Fora Research Project: Volume 4. Hominid Cranial Remains.  
959 Clarendon Press, Oxford.
- 960 Wood, B.A., Abbott, S.A., 1983. Analysis of the dental morphology of Plio-Pleistocene  
961 hominids. I. Mandibular molars: crown area measurements and morphological traits.  
962 Journal of Anatomy 136, 197-219.
- 963 Wood, B.A., Uytterschaut, H., 1987. Analysis of the dental morphology of Plio-Pleistocene  
964 hominids. III. Mandibular premolar crowns. Journal of Anatomy 154, 121-156.
- 965 Wood, B.A., Abbott, S.A., Graham, S.H., 1983. Analysis of the dental morphology of Plio-  
966 Pleistocene hominids. II. Mandibular molars – study of cusp areas, fissure pattern and  
967 cross sectional shape of the crown. Journal of Anatomy 137, 287-314.
- 968 Wu, L., Turner II, C.G., 1993. Variation in the frequency and form of the lower permanent  
969 molar middle trigonid crest. American Journal of Physical Anthropology 91, 245-248.
- 970 Zanolli, C., 2015. Molar crown inner structural organization in Javanese *Homo erectus*.  
971 American Journal of Physical Anthropology 156, 148-157.
- 972 Zanolli, C., Mazurier, A., 2013. Endostructural characterization of the *H. heidelbergensis*  
973 dental remains from the early Middle Pleistocene site of Tighenif, Algeria. Comptes  
974 Rendus Palevol 12, 293-304.
- 975 Zanolli, C., Pan, L., Dumoncel, J., Kullmer, O., Kundrať, M., Liu, W., Macchiarelli, R.,  
976 Mancini, L., Schrenk, F., Tuniz, C., 2018. Inner tooth morphology of *Homo erectus* from

977 Zhoukoudian. New evidence from an old collection housed at Uppsala University, Sweden.  
978 Journal of Human Evolution 116, 1-13.

979

## 980 **Figure captions**

981

982 **Figure 1.** Transverse crest variation. Illustration of the 5 main transverse crest forms seen in  
983 hominoid P<sub>3</sub>, as well as examples of each type. Schematic diagrams represent a left sided  
984 tooth in occlusal view. Filled black circle indicates the protoconid, dashed circle indicates the  
985 metaconid (or equivalent point on the crown, when a metaconid is not present). Example  
986 EDJs are chosen to illustrate the range of variation encompassed within each type (some are  
987 reversed such that all specimens appear left-sided): a) ULAC 797 (*H. sapiens*, reversed); b)  
988 ULAC 171 (*H. sapiens*); c) SK 100 (*P. robustus*, reversed); d) ZMB 38607 (*Pongo*, reversed);  
989 e) KRP 51 (*H. neanderthalensis*, reversed); f) KRP D33 (*H. neanderthalensis*); g) KRP D114  
990 (*H. neanderthalensis*); h) ZMB 7826 (*Hylobates*); i) KNM-ER 5431E (Hominidae gen. et sp.  
991 indet.); j) ZMB 31435 (*Gorilla*, reversed); k) MPITC 11776 (*Pan*); l) ZMB 7814 (*Hylobates*).  
992 Abbreviations: B = buccal; L = lingual; M = mesial; D = distal.

993

994 **Figure 2.** Marginal ridge continuity. Specimens are presented in lingual view at the OES  
995 (left) and EDJ (right), illustrating continuous and three types of discontinuous marginal ridges  
996 (indicated by white arrows). Specimens used, top to bottom: DNH 107 (*P. robustus*), KRP 54  
997 (*H. neanderthalensis*), ULAC 801 (*H. sapiens*), and ULAC 806 (*H. sapiens*). Abbreviations:  
998 M = mesial; D = distal.

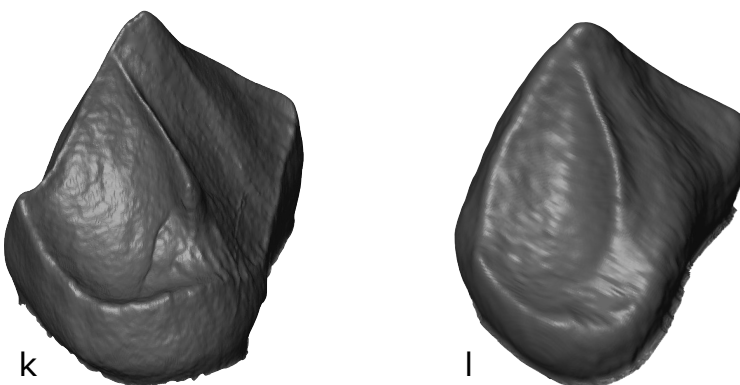
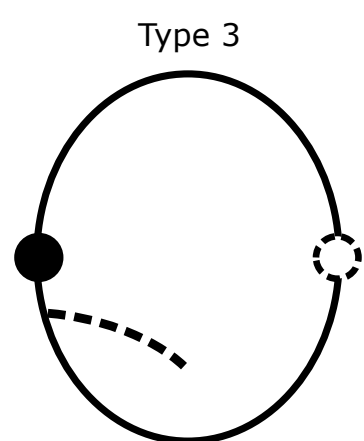
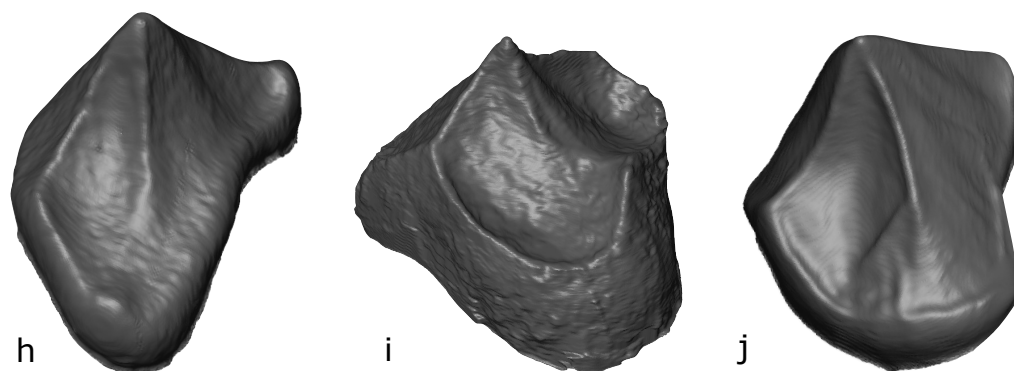
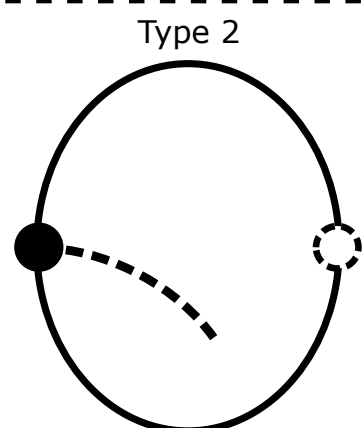
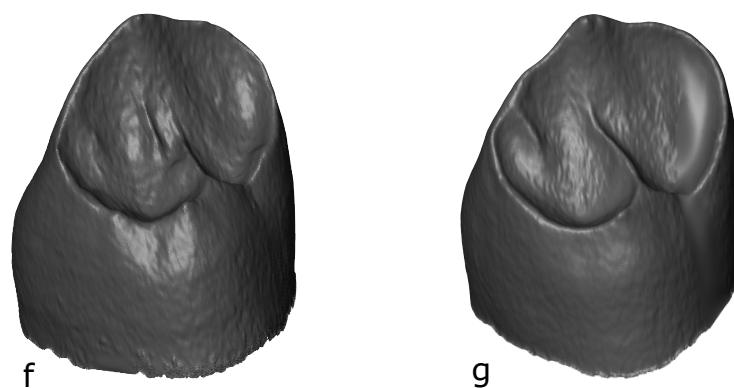
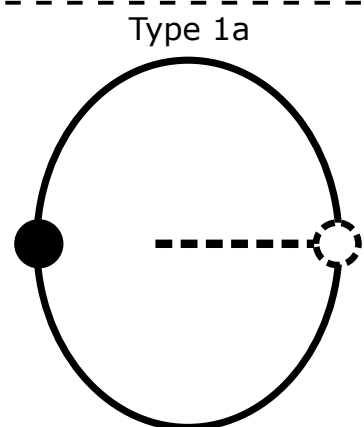
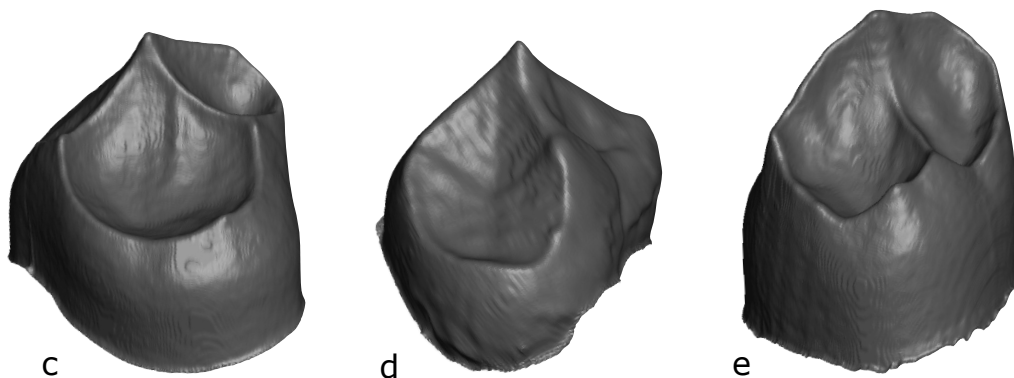
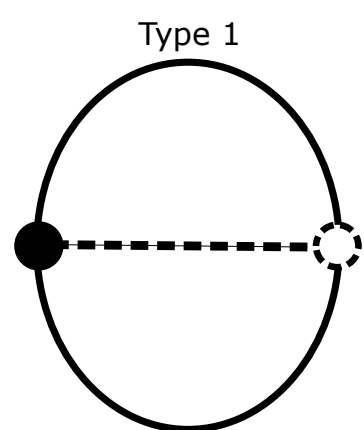
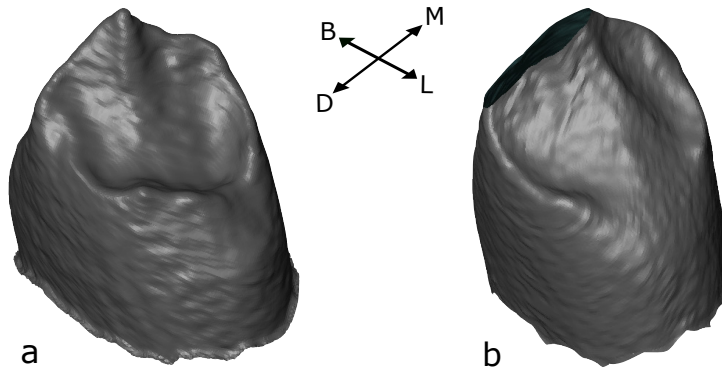
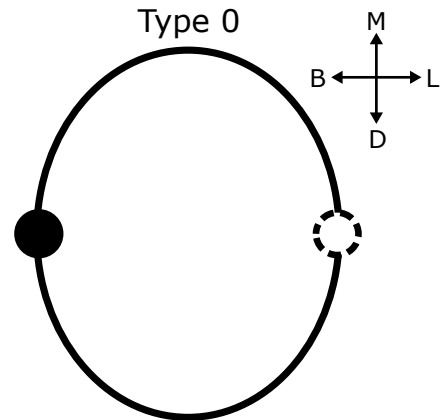
999 **Figure 3.** Buccal groove variation. Three specimens in buccal view at the OES (left) and EDJ  
1000 (right), illustrating the range in buccal groove expression, and the scoring system used. Buccal

1001 grooves are indicated by white arrows. Examples used are those which display the same  
1002 buccal groove score mesially and distally. Top: KRP 54 (*H. neanderthalensis*). Middle:  
1003 UW101 889 (*H. naledi*). Bottom: STW 213 (*A. africanus*, reversed). Abbreviations: M =  
1004 mesial; D = distal.

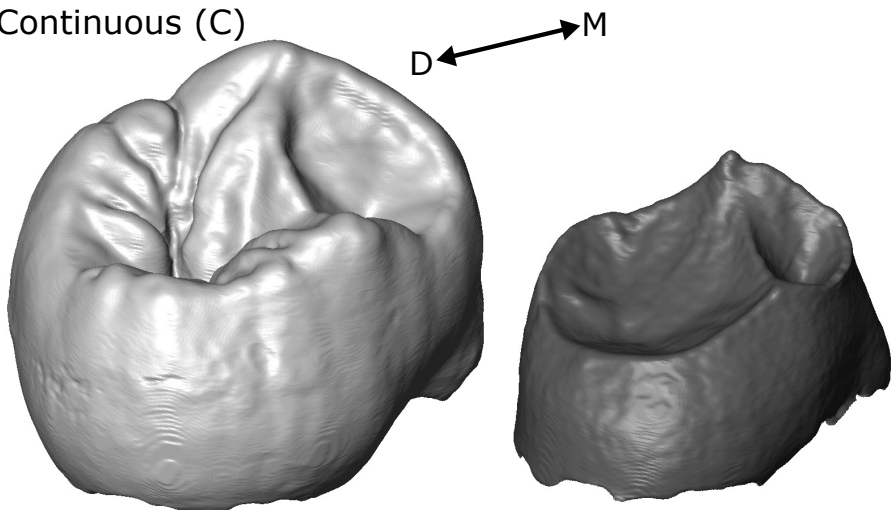
1005 **Figure 4.** Accessory crest examples. A selection of P<sub>3</sub> specimens displaying accessory crests,  
1006 marked with white arrows. Top row in oblique view, bottom row in lingual view. The images  
1007 of SK 100, DNH 46 and Combe-Grenal I have been flipped such that all specimens appear  
1008 left sided. Abbreviations: B = buccal; L = lingual; M = mesial; D = distal.

1009 **Figure 5.** Protoconid variation. Four specimens showing variation in protoconid form are  
1010 displayed. A) ‘Standard’ simple conic dentine horn in SK 100 (*P. robustus*). B) Flat ridge in  
1011 ULAC 790 (*H. sapiens*, image flipped). C) ‘Double’ dentine in UW101 377 (*H. naledi*). D)  
1012 Transversely expanded dentine horn in ULAC 58 (*H. sapiens*). A–C presented as right sided  
1013 in lingual view, D left sided in distal view. Abbreviations: B = buccal; L = lingual; M =  
1014 mesial; D = distal.

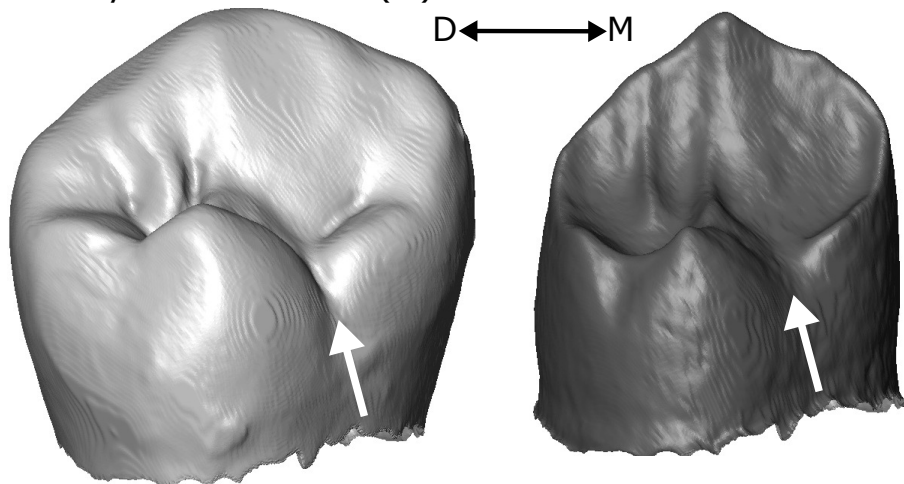
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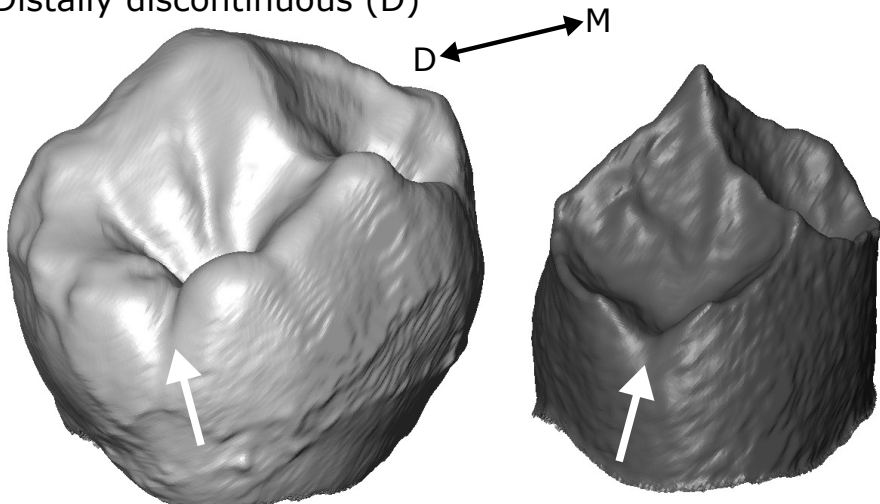
Continuous (C)



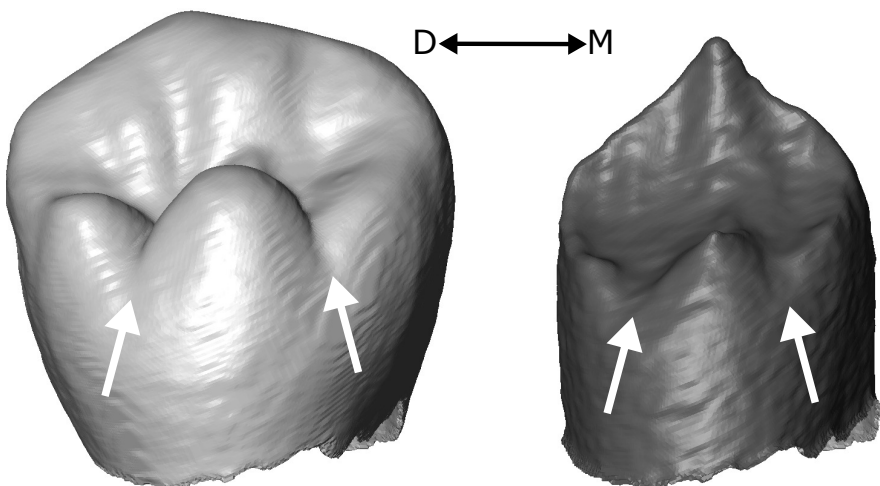
Mesially discontinuous (M)



Distally discontinuous (D)

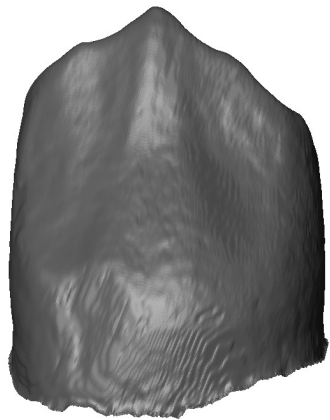
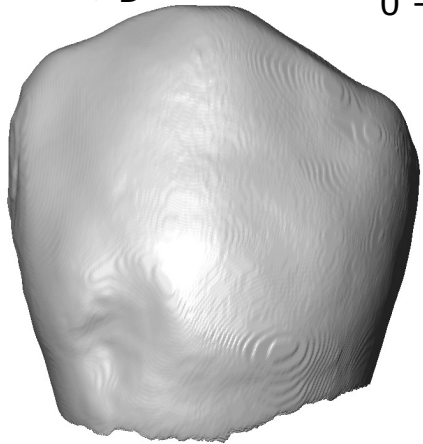


Mesially and distally discontinuous (MD)

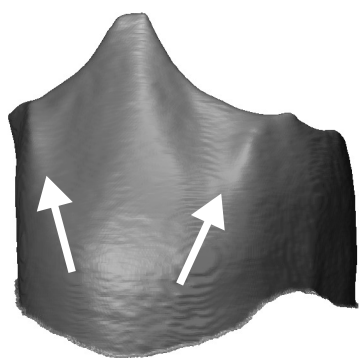
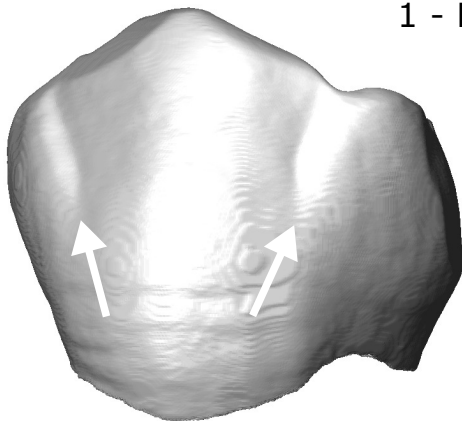


M ← → D

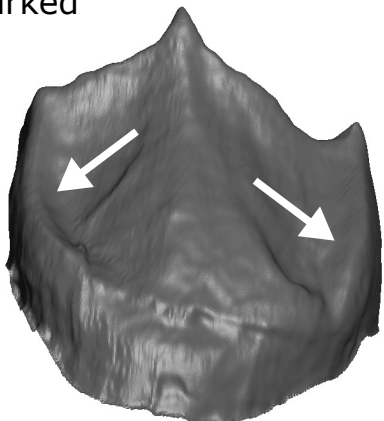
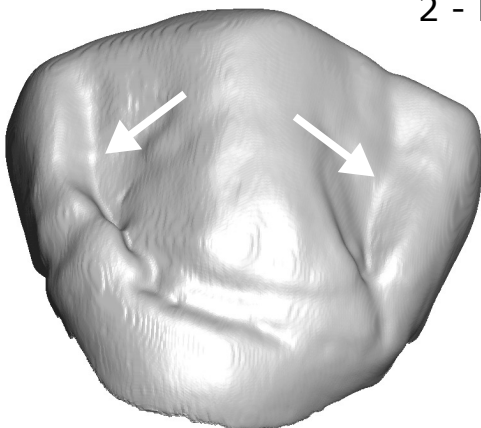
0 - Absent



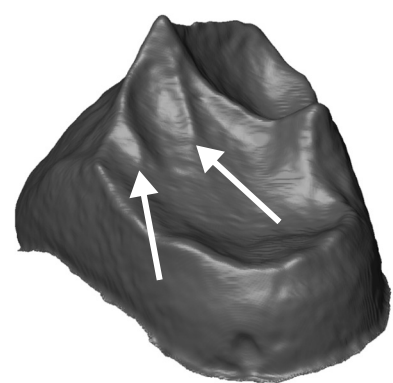
1 - Minor



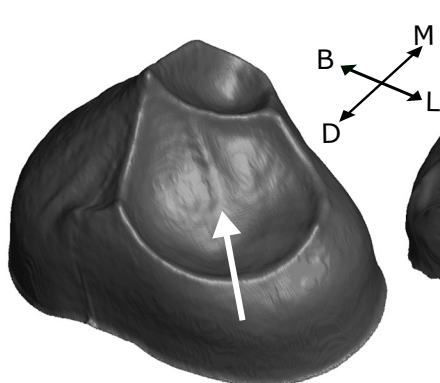
2 - Marked



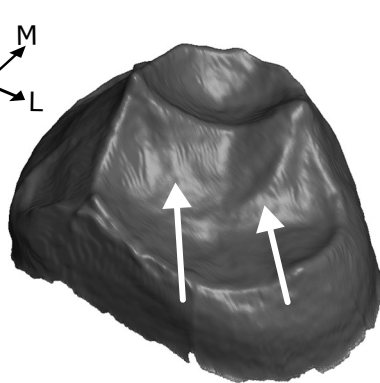




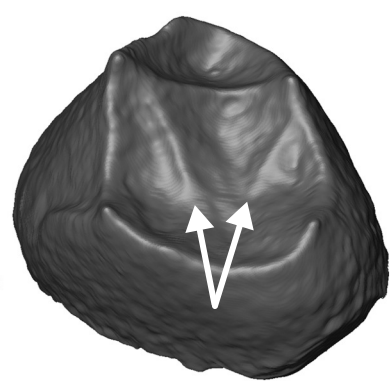
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*A. africanus*



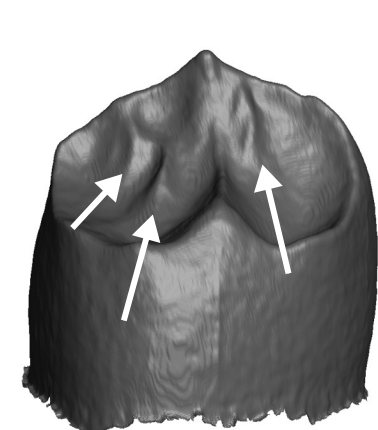
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*P. robustus*



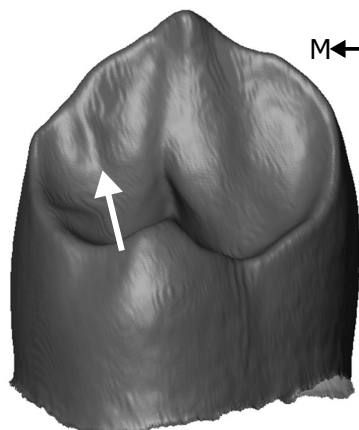
DNH 46  
*P. robustus*



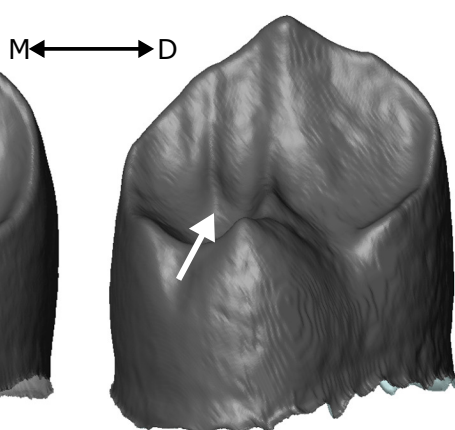
SK 62  
*P. robustus*



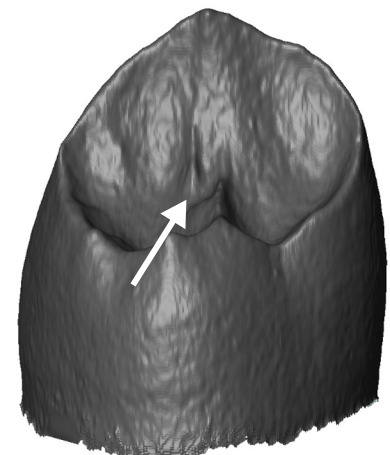
Combe-Grenal I  
*H. neanderthalensis*



KRP 52  
*H. neanderthalensis*

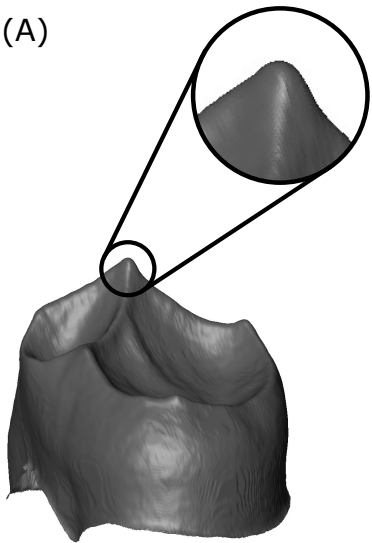


KRP 54  
*H. neanderthalensis*

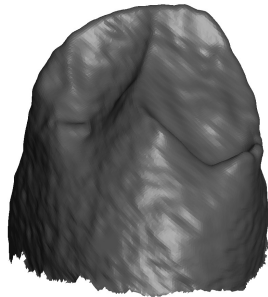


KRP D33  
*H. neanderthalensis*

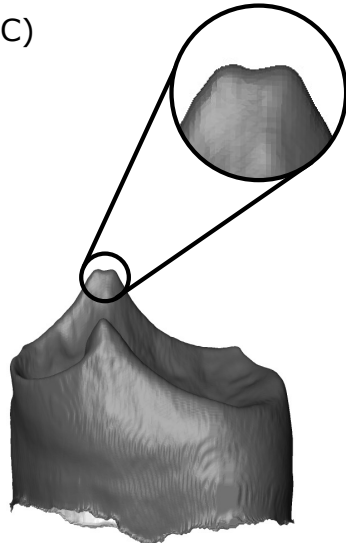
(A)

M  $\longleftrightarrow$  D

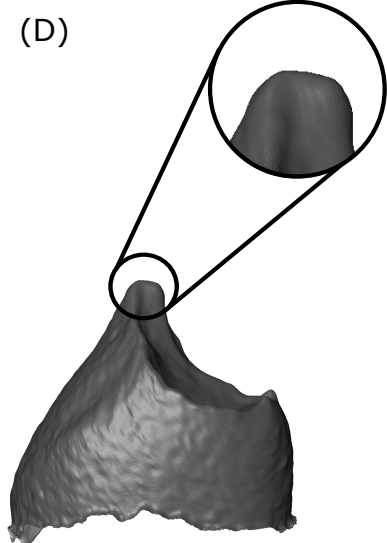
(B)

M  $\longleftrightarrow$  D

(C)

M  $\longleftrightarrow$  D

(D)

B  $\longleftrightarrow$  L

**Table 1**

P<sub>3</sub> study sample summary. The extant and fossil taxa included in the sample are listed, along with their locality, and the sample size for each of the discrete traits scored. Full specimen list can be found in SOM Table S1.<sup>a</sup>

Taxon	Locality	Transverse crest	Marginal ridge	Buccal grooves
<i>Hylobates</i>	South East Asia ( <i>Hy. muelleri</i> and <i>Hy. agilis</i> )	4	—	4
<i>Pongo</i>	Borneo; Sumatra ( <i>Po. pygmaeus</i> and <i>Po. abelii</i> )	6	—	6
<i>Gorilla</i>	Cameroon; Congo ( <i>G. gorilla</i> )	5	—	5
<i>Pan</i>	Côte d'Ivoire ( <i>Pa. troglodytes verus</i> )	5	—	5
<i>A. anamensis</i>	Kanapoi, Kenya	2	3	3
<i>A. afarensis</i>	Hadar and Omo, Ethiopia	4	3	3
<i>A. africanus</i>	Sterkfontein and Taung, South Africa	7	12	13
<i>P. robustus</i>	Drimolen and Swartkrans, South Africa	7	9	11
<i>P. boisei</i>	Koobi Fora and West Turkana, Kenya; Omo, Ethiopia	4	3	3
<i>Homo</i> sp.	Koobi Fora, Kenya; Swartkrans, South Africa	1	1	2
<i>H. naledi</i>	Rising Star cave system, South Africa	4	5	7
<i>H. heidelbergensis</i>	Mauer, Germany	0	1	1
<i>H. neanderthalensis</i>	Combe Grenal, France; Krapina, Croatia; Scladina, Belgium	9	13	14
<i>H. sapiens</i>	Qafzeh, Israel; Anatomical collection, various localities	9	14	14

<sup>a</sup> Specimens not included in these taxon groups are Cave of Hearths, KNM-ER 5431E, KNM-WT 8556, Mauer, STW 151 and W8-978.



**Table 2**

Transverse crest (TC) variation by taxon. The percentage of specimens displaying each transverse crest type is shown for each taxon (sample sizes in parentheses). See main text and Figure 2 for full details of the typology. For results by specimen, see SOM Table S1.<sup>a</sup>

Taxon ( <i>n</i> )	TC type (%)				
	0	1	1a	2	3
<i>Hylobates</i> (4)	0	0	0	50	50
<i>Pongo</i> (6)	0	100	0	0	0
<i>Gorilla</i> (5)	0	0	0	100	0
<i>Pan</i> (5)	0	0	0	20	80
<i>A. anamensis</i> (2)	0	100	0	0	0
<i>A. afarensis</i> (4)	0	100	0	0	0
<i>A. africanus</i> (7)	0	100	0	0	0
<i>P. robustus</i> (7)	0	100	0	0	0
<i>P. boisei</i> (4)	50	50	0	0	0
<i>Homo</i> sp. (1)	0	100	0	0	0
<i>H. naledi</i> (4)	0	100	0	0	0
<i>H. neanderthalensis</i> (9)	0	56	44	0	0
<i>H. sapiens</i> (9)	44	56	0	0	0
Extant ape total (20)	0	30	0	40	30
Hominin total (51)	12	78	8	2	0

<sup>a</sup> Specimens which could not be assigned to taxon groups (transverse crest type in parentheses): Cave of Hearths (1), KNM-ER 5431E (2), KNM-WT 8556 (1), STW 151 (1).

**Table 3**

Marginal ridge (MR) variation. The percentage of specimens displaying each marginal ridge type is shown for each taxon (sample sizes in parentheses). See main text and Figure 3 for full details of the typology. For results by specimen, see SOM Table S1.

Taxon ( <i>n</i> )	MR type (%)			
	C	M	D	MD
<i>A. anamensis</i> (3)	0	100	0	0
<i>A. afarensis</i> (3)	33	67	0	0
<i>A. africanus</i> (12)	75	17	8	0
<i>P. robustus</i> (9)	100	0	0	0
<i>P. boisei</i> (3)	100	0	0	0
<i>Homo</i> sp. (1)	100	0	0	0
<i>H. naledi</i> (5)	100	0	0	0
<i>H. heidelbergensis</i> (1)	0	0	0	100
<i>H. neanderthalensis</i> (13)	69	31	0	0
<i>H. sapiens</i> (14)	50	14	7	29
Hominin total (69)	70	20	3	7

Abbreviations: C = continuous; M = mesially discontinuous; D = distally discontinuous; MD = mesially and distally discontinuous

<sup>a</sup> Specimens which could not be assigned to taxon groups (marginal ridge type in parentheses): Cave of Hearths (M), KNM-ER 5431E (C), KNM-WT 8556 (C), STW 151 (C), W8-978 (C).

**Table 4**

Buccal groove variation. The percentage of specimens displaying each mesial and distal buccal groove type is shown for each taxon (sample sizes in parentheses). See main text and Figure 4 for full details of the typology. For results by specimen, see SOM Table S1.<sup>a</sup>

Taxon ( <i>n</i> )	MBG type (%) <sup>b</sup>			DBG type (%) <sup>b</sup>		
	0	1	2	0	1	2
<i>Hylobates</i> (4)	75	25	0	100	0	0
<i>Pongo</i> (6)	100	0	0	100	0	0
<i>Gorilla</i> (5)	60	40	0	0	100	0
<i>Pan</i> (5)	40	60	0	0	100	0
<i>A. anamensis</i> (3)	0	0	100	0	67	33
<i>A. afarensis</i> (3)	0	33	67	0	33	67
<i>A. africanus</i> (13)	0	15	85	0	69	31
<i>P. robustus</i> (11)	91	9	0	18	64	18
<i>P. boisei</i> (3)	67	0	33	33	33	33
<i>Homo</i> sp. (2)	0	0	100	50	50	0
<i>H. naledi</i> (7)	14	86	0	14	86	0
<i>H. heidelbergensis</i> (1)	100	0	0	100	0	0
<i>H. neanderthalensis</i> (14)	43	57	0	79	21	0
<i>H. sapiens</i> (14)	93	7	0	86	14	0
Extant ape total (20)	70	30	0	50	50	0
Hominin total (75)	45	25	29	40	45	15

Abbreviations: MBG = mesial buccal groove; DBG = distal buccal groove.

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<sup>a</sup> Specimens which could not be assigned to taxon groups (buccal groove types in parentheses in the form MBG:DBG): Cave of Hearths (0:0), KNM-WT 8556 (2:1), STW 151 (2:2), W8-978 (2:1).

<sup>b</sup> MBG and DBG states: 0 = absent; 1 = minor; 2 = marked.



## Supplementary Online Material (SOM)

Endostructural morphology in hominoid mandibular third premolars: Discrete traits at the enamel-dentine junction

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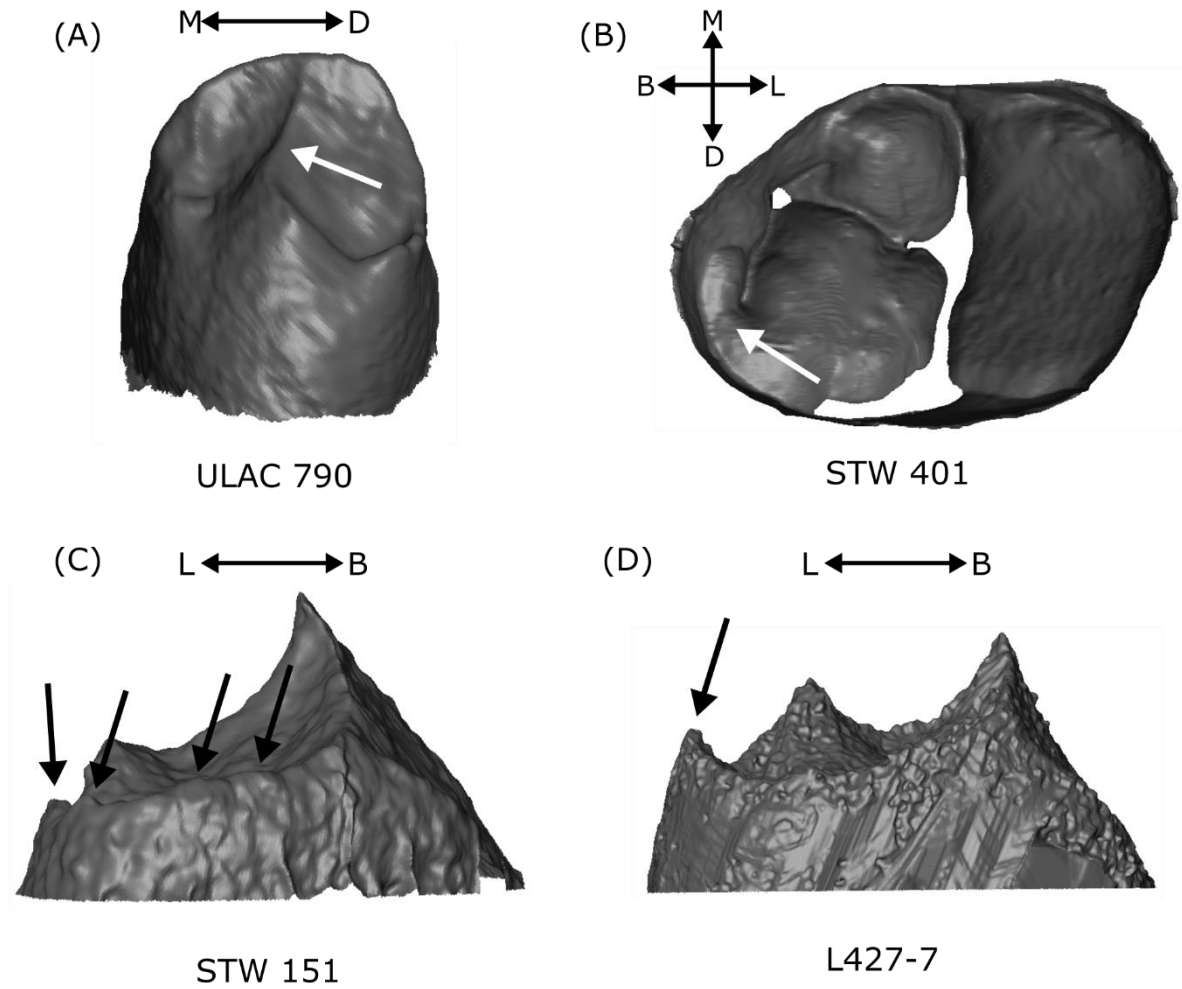
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**SOM Figure S1.** EDJ surface for selected specimens discussed in the main text. A) ULAC 790 (reversed) in lingual view, arrow indicates transverse crest. B) STW 401 in occlusal view, arrow indicates discontinuous distal marginal ridge. C) STW 151 in distal view, arrows indicate possible accessory cusps. D) L427-7 in distal view, arrow indicates accessory cusp.

**SOM Table S1.** Detailed study sample, including which analyses each specimen is included in.

Specimen	Side	Site/Origin	Taxonomy	Source	Position basis <sup>a</sup>	Position source	TC	MR	MBG	DBG	Recon ?
ZMB 7814	L	Borneo	<i>Hylobates muelleri</i>	ZMB records	1	ZMB records	3	—	0	0	—
ZMB 7826	L	Borneo	<i>Hylobates muelleri</i>	ZMB records	1	ZMB records	2	—	1	0	—
ZMB 7828	L	Borneo	<i>Hylobates muelleri</i>	ZMB records	1	ZMB records	3	—	0	0	—
ZMB 85368	L	Sumatra, Indonesia	<i>Hylobates agilis</i>	ZMB records	1	ZMB records	2	—	0	0	Prd
ZMB 6948	R	Borneo	<i>Pongo pygmaeus</i>	ZMB records	1	ZMB records	1	—	0	0	Prd
ZMB 6957	L	Borneo	<i>Pongo pygmaeus</i>	ZMB records	1	ZMB records	1	—	0	0	—
ZMB 12209	R	Sumatra, Indonesia	<i>Pongo abelii</i>	ZMB records	1	ZMB records	1	—	0	0	Prd
ZMB 38607	R	Sumatra, Indonesia	<i>Pongo abelii</i>	ZMB records	1	ZMB records	1	—	0	0	—
ZMB 83509	R	Sumatra, Indonesia	<i>Pongo abelii</i>	ZMB records	1	ZMB records	1	—	0	0	—
ZMB 83511	L	Sumatra, Indonesia	<i>Pongo abelii</i>	ZMB records	1	ZMB records	1	—	0	0	Prd
ZMB 17963	L	Cameroon	<i>Gorilla gorilla</i>	ZMB records	1	ZMB records	2	—	1	1	Prd
ZMB 30940	R	Cameroon	<i>Gorilla gorilla</i>	ZMB records	1	ZMB records	2	—	0	1	—
ZMB 30941	L	Congo	<i>Gorilla gorilla</i>	ZMB records	1	ZMB records	2	—	0	1	Prd
ZMB 31435	R	Cameroon	<i>Gorilla gorilla</i>	ZMB records	1	ZMB records	2	—	1	1	—
ZMB 83561	R	Cameroon	<i>Gorilla gorilla</i>	ZMB records	1	ZMB records	2	—	0	1	Prd
MPITC 11776	L	Taï, Côte d'Ivoire	<i>Pan troglodytes verus</i>	MPI records	1	ZMB records	3	—	1	1	—
MPITC 11800	R	Taï, Côte d'Ivoire	<i>Pan troglodytes verus</i>	MPI records	1	ZMB records	3	—	1	1	—
MPITC 11903	R	Taï, Côte d'Ivoire	<i>Pan troglodytes verus</i>	MPI records	1	ZMB records	3	—	1	1	Prd
MPITC 13430	R	Taï, Côte d'Ivoire	<i>Pan troglodytes verus</i>	MPI records	1	ZMB records	2	—	0	1	—
MPITC 13437	R	Taï, Côte d'Ivoire	<i>Pan troglodytes verus</i>	MPI records	1	ZMB records	3	—	0	1	—
KNM-KP 29281	R	Kanapoi, Kenya	<i>Australopithecus anamensis</i>	Leakey et al. 1995	1	Ward et al., 2001	—	M	2	1	—
KNM-KP 29286	R	Kanapoi, Kenya	<i>Australopithecus anamensis</i>	Leakey et al. 1995	1	Ward et al., 2001	1	M	2	2	—
KNM-KP 53160	L	Kanapoi, Kenya	<i>Australopithecus anamensis</i>	Ward et al. 2017	1	Ward et al. 2017	1	M	2	1	—
AL266-1	R	Hadar, Ethiopia	<i>Australopithecus afarensis</i>	Johanson et al., 1982	1	Johanson et al., 1982	1	M	1	1	Prd
AL333-10	L	Hadar, Ethiopia	<i>Australopithecus afarensis</i>	Johanson et al., 1982	3	Johanson et al., 1982	1	C	2	2	Prd
AL333w-1c	R	Hadar, Ethiopia	<i>Australopithecus afarensis</i>	Johanson et al., 1982	2	Johanson et al., 1982	1	M	2	2	—
AL655-1	L	Hadar, Ethiopia	<i>Australopithecus afarensis</i>	Kimbel and Delezeze, 2009	3	Kimbel and Delezeze, 2009	1	—	—	—	—
W8-978	R	Omo, Ethiopia	Indet.	Suwa, 1990	3	Suwa, 1990	—	C	2	1	—
KNM-WT 8556	L	West Turkana, Kenya	Indet.	Brown et al., 2001	1	Brown et al., 2001	1	C	2	1	—
STW 7	L	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Moggi-Cecchi et al., 2006	3	Moggi-Cecchi et al., 2006	1	C	1	1	Prd
STW 104	L	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Moggi-Cecchi et al., 2006	1	Moggi-Cecchi et al., 2006	1	C	2	1	—

STW 142	R	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Moggi-Cecchi et al., 2006	1	Moggi-Cecchi et al., 2006	—	C	2	1	Med
STW 193	R	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Moggi-Cecchi et al., 2006	2	Moggi-Cecchi et al., 2006	—	—	2	1	—
STW 213	R	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Moggi-Cecchi et al., 2006	2	Moggi-Cecchi et al., 2006	1	M	2	2	Prd
STW 401	R	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Moggi-Cecchi et al., 2006	3	Moggi-Cecchi et al., 2006	—	D	2	2	Med
STW 404	R	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Moggi-Cecchi et al., 2006	1	Moggi-Cecchi et al., 2006	—	C	2	1	Prd
STW 420B	L	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Moggi-Cecchi et al., 2006	2	Moggi-Cecchi et al., 2006	1	C	2	2	—
STW 498c	L	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Moggi-Cecchi et al., 2006	1	Moggi-Cecchi et al., 2006	—	C	2	1	—
STS 24	L	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Brain, 1981	1	Brain, 1981	1	C	1	1	—
STS 51	R	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Brain, 1981	2	Brain, 1981	1	C	2	1	—
STS 52b	R	Sterkfontein, S. Africa	<i>Australopithecus africanus</i>	Dart, 1954	1	Dart, 1954	—	C	2	2	Prd
Taung1	R	Taung, S. Africa	<i>Australopithecus africanus</i>	Dart, 1925	1	Dart, 1925	1	M	2	1	—
DNH8	L	Drimolen, S. Africa	<i>Paranthropus robustus</i>	Moggi-Cecchi et al., 2010	1	Moggi-Cecchi et al., 2010	1	C	0	1	—
DNH46	R	Drimolen, S. Africa	<i>Paranthropus robustus</i>	Moggi-Cecchi et al., 2010	1	Moggi-Cecchi et al., 2010	1	—	—	—	—
DNH51	R	Drimolen, South Africa	<i>Paranthropus robustus</i>	Moggi-Cecchi et al., 2010	1	Moggi-Cecchi et al., 2010	—	C	0	1	—
DNH107	L	Drimolen, S. Africa	<i>Paranthropus robustus</i>	Museum records	2	Museum records	1	C	0	2	—
SK23	L	Swartkrans, S. Africa	<i>Paranthropus robustus</i>	Robinson, 1956	1	Robinson, 1956	—	C	0	0	—
SK30	L	Swartkrans, S. Africa	<i>Paranthropus robustus</i>	Robinson, 1956	3	Robinson, 1956	—	—	0	1	—
SK61	R	Swartkrans, S. Africa	<i>Paranthropus robustus</i>	Robinson, 1956	1	Robinson, 1956	1	C	0	1	—
SK62	L	Swartkrans, S. Africa	<i>Paranthropus robustus</i>	Robinson, 1956	1	Robinson, 1956	1	C	1	2	—
SK63	L	Swartkrans, S. Africa	<i>Paranthropus robustus</i>	Robinson, 1956	1	Robinson, 1956	—	—	0	1	—
SK100	R	Swartkrans, S. Africa	<i>Paranthropus robustus</i>	Robinson, 1956	3	Oakley, 1977	1	C	0	1	—
SK857	R	Swartkrans, S. Africa	<i>Paranthropus robustus</i>	Robinson, 1956	3	Oakley, 1977	1	C	0	0	—
SKW5	R	Swartkrans, S. Africa	<i>Paranthropus robustus</i>	Grine and Daegling, 1993	1	Grine and Daegling, 1993	—	C	0	1	Prd
KNM-ER 1820	L	Koobi Fora, Kenya	<i>Paranthropus boisei</i>	Wood, 1991	1	Wood, 1991	1	C	—	—	—
KNM-ER 6082	L	Koobi Fora, Kenya	<i>Paranthropus boisei</i>	Wood, 1991	3	Wood, 1991	0	C	—	—	—
KNM-ER 15951H	L	Koobi Fora, Kenya	<i>Paranthropus boisei</i>	Wood and Leakey, 2011	2	Wood and Leakey, 2011	—	—	0	0	—
KNM-WT 16005	L	West Turkana, Kenya	<i>Paranthropus boisei</i>	Leakey and Walker, 1988	1	Leakey and Walker, 1988	1	C	0	1	Prd
L427-7	R	Omo, Ethiopia	<i>Paranthropus boisei</i>	Suwa et al., 1996	1	Suwa et al., 1996	0	—	2	2	—
KNM-ER 806E	L	Koobi Fora, Kenya	<i>Homo sp. (Homo ergaster)</i>	Wood, 1991	2	Wood, 1991	—	—	2	0	—
KNM-ER 5431E	L	Koobi Fora, Kenya	Indet.	Wood, 1991	2	Wood, 1991	2	C	—	—	—
SKX 21204	R	Swartkrans, S. Africa	<i>Homo sp.</i>	Grine, 1989	1	Grine, 1989	1	C	2	1	—
STW 151	R	Sterkfontein, S. Africa	Indet.	Moggi-Cecchi et al., 1998	1	Moggi-Cecchi et al., 1998	1	C	2	2	—
U.W. 101-0010	R	Rising Star, S. Africa	<i>Homo naledi</i>	Berger et al. 2015	1	Berger et al. 2015	—	C	1	1	—
U.W. 101-144	L	Rising Star, S. Africa	<i>Homo naledi</i>	Berger et al. 2015	3	Berger et al. 2015	1	C	1	0	—

U.W. 101-377 <sup>b</sup>	R	Rising Star, S. Africa	<i>Homo naledi</i>	Berger et al. 2015	1	Berger et al. 2015	—	—	—	—	—
U.W. 101-506 <sup>b</sup>	R	Rising Star, S. Africa	<i>Homo naledi</i>	Berger et al. 2015	3	Berger et al. 2015	—	—	—	—	—
U.W. 101-850	R	Rising Star, S. Africa	<i>Homo naledi</i>	Berger et al. 2015	3	Berger et al. 2015	—	—	1	1	—
U.W. 101-889	L	Rising Star, S. Africa	<i>Homo naledi</i>	Berger et al. 2015	3	Berger et al. 2015	1	C	1	1	—
U.W. 101-1261	R	Rising Star, S. Africa	<i>Homo naledi</i>	Berger et al. 2015	1	Berger et al. 2015	1	C	1	1	Prd
U.W. 101-1565	L	Rising Star, S. Africa	<i>Homo naledi</i>	Berger et al. 2015	1	Berger et al. 2015	1	C	0	1	—
U.W. 102-0023	R	Rising Star, S. Africa	<i>Homo naledi</i>	Hawks et al. 2017	3	Hawks et al. 2017	—	—	1	1	—
Cave of hearths	R	Cave of hearths, S. Africa	Indet.	Tobias, 1971	1	Tobias, 1971	1	M	0	0	—
Mauer 1	R	Mauer, Germany	<i>Homo heidelbergensis</i>	Schoetensack, 1908	1	Schoetensack, 1908	—	MD	0	0	—
Combe-Grenal I	R	Combe Grenal, France	<i>Homo neanderthalensis</i>	Garralda and Vandermeersch, 2000	1	Garralda and Vandermeersch, 2000	1	C	1	0	—
KRP 51	R	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	1	Radovčić, 1988	1	C	1	1	—
KRP 52	L	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	1	Radovčić, 1988	1a	C	1	0	—
KRP 54	L	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	1	Radovčić, 1988	1a	M	0	0	—
KRP 55	L	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	1	Radovčić, 1988	1	C	0	0	—
KRP 58	R	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	1	Radovčić, 1988	—	C	1	0	—
KRP D27	L	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	2	Radovčić, 1988	—	C	1	0	—
KRP D28	R	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	2	Radovčić, 1988	—	M	0	0	—
KRP D29	R	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	2	Radovčić, 1988	—	M	0	0	—
KRP D33	L	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	2	Radovčić, 1988	1a	C	0	0	—
KRP D34	R	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	3	Radovčić, 1988	—	—	1	1	Prd
KRP D111	L	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	3	Radovčić, 1988	1	M	1	1	—
KRP D114	L	Krapina, Croatia	<i>Homo neanderthalensis</i>	Radovčić, 1988	2	Radovčić, 1988	1a	C	1	0	—
SCLA 4A 6	R	Scladina, Belgium	<i>Homo neanderthalensis</i>	Toussaint et al., 1998	2	Toussaint et al., 1998	1	C	0	0	—
Qafzeh 10	R	Qafzeh, Israel	Fossil <i>Homo sapiens</i>	Vandermeersch, 1981	1	Vandermeersch, 1981	1	C	0	0	—
Qafzeh 11	R	Qafzeh, Israel	Fossil <i>Homo sapiens</i>	Vandermeersch, 1981	1	Vandermeersch, 1981	1	M	0	0	—
ULAC 1	R	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	—	C	0	0	—
ULAC 58	L	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	1	M	0	0	—
ULAC 58 <sup>b</sup>	R	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	—	—	—	—	—
ULAC 66	L	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	0	MD	0	0	Prd
ULAC 74	L	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	—	C	0	1	—
ULAC 171	L	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	0	MD	0	0	—
ULAC 522	L	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	—	C	0	0	—
ULAC 536	R	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	1	C	0	0	Prd
ULAC 607	R	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	—	C	0	0	—

ULAC 790	L	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	*	MD	0	0	—
ULAC 797	R	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	0	C	0	0	—
ULAC 801	L	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	1	D	1	1	—
ULAC 806	L	Anatomical collection	<i>Homo sapiens</i>	ULAC records	1	ULAC records	0	MD	0	0	—

Abbreviations: TC = transverse crest form; see main text for typology (\* = EDJ preserved but transverse crest form could not be determined—see main text for details); MR = marginal ridge (C = continuous; M = mesially discontinuous; D = distally discontinuous; MD = mesially and distally discontinuous); MBG = mesial buccal groove (0 = absent; 1 = minor; 2 = marked; for description of these character states, see main text); DBG = distal buccal groove (character states as for MBG); Recon? = specimens with reconstructed dentine horns (Prd = protoconid reconstructed; Med = metaconid reconstructed).

<sup>a</sup> Position basis: 1 = In jaw; 2 = associated dentition; 3 = based on morphology.

<sup>b</sup> Antimere specimens that are not included in analyses, but are discussed in the main text.

**SOM Table S2**

Additional information on the modern human sample, as listed in the records of the Anatomical Collection of the University of Leipzig

Specimen number	Region	Age	Sex
ULAC_1	Germany/Rheinland	Adult	Male
ULAC_58	Norway	Adult	Male
ULAC_66	Norway/Sweden	Adult	Female
ULAC_74	Italy (Etruscan, Tarquinii)	Adult	Male
ULAC_171	Italy (Etruscan, Tarquinii)	Adult	Male
ULAC_522	Egypt (Thebes)	Adult	Male
ULAC_536	Egypt (Thebes)	Adult	Male
ULAC_607	Egypt (Thebes)	Adult	Male
ULAC_790	Africa (Americans/New Orleans)	Adult	Male
ULAC_797	Africa (Americans/New Orleans)	Adult	Male
ULAC_801	Africa (Americans/New Orleans)	Adult	Female
ULAC_806	Africa (Americans/New Orleans)	Adult	Male

**SOM Table S3**

Results of the inter- and intra-observer error tests for the four P<sub>3</sub> discrete traits scored.

Trait	Inter-observer agreement (%)	Intra-observer agreement (%)
Transverse crest	100	98
Marginal ridge	92	99
Mesial buccal groove	96	89
Distal buccal groove	88	93



## SOM References

- Berger, L.R., Hawks, J., de Ruiter, D.J., Churchill, S.E., Schmid, P., Deleuzene, L.K., Kivell, T.L., Garvin, H.M., Williams, S.A., DeSilva, J.M., Skinner, M.M., Musiba, C.M., Cameron, N., Holliday, T.W., Harcourt-Smith, W., Ackermann, R.R., Bastir, M., Bogin, B., Bolter, D., Brophy, J., Cofran, Z.D., Congdon, K.A., Deane, A.S., Dembo, M., Drapeau, M., Elliott, M.C., Feuerriegel, E.M., Garcia-Martinez, D., Green, D.J., Gurtov, A., Irish, J.D., Kruger, A., Laird, M.F., Marchi, D., Meyer, M.R., Nalla, S., Negash, E.W., Orr, C.M., Radovic, D., Schroeder, L., Scott, J.E., Throckmorton, Z., Tocheri, M.W., VanSickle, C., Walker, C.S., Wei, P., Zipfel, B., 2015. *Homo naledi*, a new species of the genus *Homo* from the Dinaledi Chamber, South Africa. *eLife* 4, e09560.
- Brain, C.K., 1981. *The Hunters or the Hunted?: An Introduction to African Cave Taphonomy*. University of Chicago Press, Chicago.
- Brown, B., Brown, F.H., Walker, A., 2001. New hominids from the Lake Turkana basin, Kenya. *Journal of Human Evolution* 41, 29-44.
- Dart, R.A., 1925. *Australopithecus africanus* the man-ape of South Africa. *Nature* 115, 195-199.
- Dart, R.A., 1954. The second, or adult, female mandible of *Australopithecus prometheus*. *American Journal of Physical Anthropology* 12, 313-344.
- Garralda, M.-D., Vandermeersch, B., 2000. Les Néandertaliens de la grotte de Combe-Grenal (Domme, Dordogne, France)/The Neanderthals from Combe-Grenal cave (Domme, Dordogne, France). *Paléo* 12, 213-259.
- Grine, F.E., 1989. New hominid fossils from the Swartkrans Formation (1979-1986 excavations): craniodental specimens. *American Journal of Physical Anthropology* 79, 409-449.
- Grine, F.E., Daegling, D.J., 1993. New mandible of *Paranthropus robustus* from Member 1, Swartkrans Formation, South Africa. *Journal of Human Evolution* 24, 319-333.
- Hawks, J., Elliott, M., Schmid, P., Churchill, S.E., Ruiter, D.J.d., Roberts, E.M., Hilbert-Wolf, H., Garvin, H.M., Williams, S.A., Deleuzene, L.K., Feuerriegel, E.M., Randolph-Quinney, P., Kivell, T.L., Laird, M.F., Tawane, G., DeSilva, J.M., Bailey, S.E., Brophy, J.K., Meyer, M.R., Skinner, M.M., Tocheri, M.W., VanSickle, C., Walker, C.S., Campbell, T.L., Kuhn, B., Kruger, A., Tucker, S., Gurtov, A., Hlophe, N., Hunter, R., Morris, H., Peixotto, B., Ramalepa, M., Rooyen, D.v., Tsikoane, M., Boshoff, P., Dirks, P.H.G.M., Berger, L.R., 2017. New fossil remains of *Homo naledi* from the Lesedi Chamber, South Africa. *eLife* 6, e24232.
- Johanson, D.C., White, T.D., Coppens, Y., 1982. Dental remains from the Hadar Formation, Ethiopia: 1974-1977 collections. *American Journal of Physical Anthropology* 57, 545-603.
- Kimbel, W.H., Deleuzene, L.K., 2009. "Lucy" redux: A review of research on *Australopithecus afarensis*. *American Journal of Physical Anthropology* 140, 2-48.
- Kimbel, W.H., Johanson, D.C., Rak, Y., 1994. The first skull and other new discoveries of *Australopithecus afarensis* at Hadar, Ethiopia. *Nature* 368, 449.
- Leakey, M.G., Feibel, C.S., McDougall, I., Walker, A., 1995. New four-million-year-old hominid species from Kanapoi and Allia Bay, Kenya. *Nature* 376, 565.
- Leakey, R., Walker, A., 1988. New *Australopithecus boisei* specimens from east and west Lake Turkana, Kenya. *American Journal of Physical Anthropology* 76, 1-24.
- Moggi-Cecchi, J., Grine, F.E., Tobias, P.V., 2006. Early hominid dental remains from Members 4 and 5 of the Sterkfontein Formation (1966-1996 excavations): catalogue, individual associations, morphological descriptions and initial metrical analysis. *Journal of Human Evolution* 50, 239-328.
- Moggi-Cecchi, J., Menter, C., Boccone, S., Keyser, A., 2010. Early hominin dental remains from the Plio-Pleistocene site of Drimolen, South Africa. *Journal of Human Evolution* 58, 374-405.
- Moggi-Cecchi, J., Tobias, P.V., Beynon, A., 1998. The mixed dentition and associated skull fragments of a juvenile fossil hominid from Sterkfontein, South Africa. *American Journal of Physical Anthropology* 106, 425-465.

- Oakley, K.P., Campbell, B.G., Molleson, T.I., Museum, B., 1977. Catalogue of Fossil Hominids. Part I: Africa. Trustees of the British Museum (Natural History), London.
- Radovčić, J., 1988. The Krapina Hominids: An Illustrated Catalog of Skeletal Collection. Mladost, Zagreb.
- Robinson, J.T., 1956. The Dentition of Australopithecinae. Transvaal Museum, Pretoria.
- Schoetensack, O., 1908. Der Unterkiefer des *Homo heidelbergensis* aus den Sanden von Mauer bei Heidelberg. Ein Beitrag zur Paläontologie des Menschen. Zeitschrift für Induktive Abstammungs- und Vererbungslehre 1, 408-410.
- Suwa, G., 1990. A Comparative analysis of hominid dental remains from the Sungura and Usno Formations. Omo Valley, Ethiopia. Ph.D. Dissertation, University of California, Berkeley.
- Suwa, G., White, T.D., Howell, F.C., 1996. Mandibular postcanine dentition from the Shungura Formation, Ethiopia: Crown morphology, taxonomic allocations, and Plio-Pleistocene hominid evolution. American Journal of Physical Anthropology 101, 247-282.
- Tobias, P.V., 1971. Human skeletal remains from the Cave of Hearths, Makapansgat, northern Transvaal. American Journal of Physical Anthropology 34, 335-367.
- Toussaint, M., Otte, M., Bonjean, D., Bocherens, H., Falguères, C., Yokoyama, Y., 1998. Les restes humains néandertaliens immatures de la couche 4A de la grotte Scladina (Andenne, Belgique). Comptes Rendus de l'Académie des Sciences Paris 326, 737-742.
- Vandermeersch, B., 1981. Les Hommes Fossiles de Qafzeh (Israël). Editions du Centre National de la Recherche Scientifique, Paris.
- Walker, A., Leakey, R.E. (Eds.), 1993. The Nariokotome *Homo erectus* Skeleton. Harvard University Press, Cambridge.
- Ward, C., Manthi, F., Plavcan, J., 2017. New fossils of *Australopithecus anamensis* from Kanapoi, West Turkana, Kenya (2012–2015). Journal of Human Evolution. <https://doi.org/10.1016/j.jhevol.2017.07.008>
- Ward, C.V., Leakey, M.G., Walker, A., 2001. Morphology of *Australopithecus anamensis* from Kanapoi and Allia Bay, Kenya. Journal of Human Evolution 41, 255-368.
- Wood, B., 1991. Koobi Fora Research Project Volume 4: Hominid Cranial Remains from Koobi Fora. Clarendon, Oxford.
- Wood, B., Leakey, M., 2011. The Omo-Turkana Basin fossil hominins and their contribution to our understanding of human evolution in Africa. Evolutionary Anthropology 20, 264-292.