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1 **The Biorhythm of Human Skeletal Growth**

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29 The biorhythm of human skeletal growth

30

31 **KEYWORDS**

32 Retzius [line](#) periodicity. Enamel thickness. Daily enamel secretion rates. Osteocyte lacunar density.
33 Stature.

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35

Abstract

Evidence of a [periodic](#) biorhythm is retained in tooth enamel in the form of Retzius lines. The periodicity of Retzius lines (RP) correlates with body mass and the scheduling of life history events when compared between [some mammalian](#) species. The correlation has led to the development of the inter-specific Havers-Halberg Oscillation (HHO) hypothesis, which holds great potential for studying [aspects of a fossil species biology from](#) teeth. Yet, our understanding of if, or how the HHO relates to human skeletal growth is limited. The goal here is to explore associations between the biorhythm and two hard tissues that form at different times during human ontogeny, [within the context of the HHO](#). First, we investigate the relationship of RP to permanent molar enamel thickness and the underlying daily rate that ameloblasts secrete enamel during childhood. Following this, we develop preliminary research conducted on small samples of adult human bone by testing associations between RP, adult femoral length (as a proxy for attained adult stature), and cortical osteocyte lacunae density (as a proxy for the rate of osteocyte proliferation). Results reveal RP is positively correlated with enamel thickness, negatively correlated with femoral length, but weakly associated with the rate of enamel secretion and osteocyte proliferation. These new data imply that a slower biorhythm predicts thicker enamel for children but shorter stature for adults. Our results develop the intra-specific HHO hypothesis suggesting that there is a common underlying systemic biorhythm that has a role in the final products of human enamel and bone growth.

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64 **Introduction**

65 Biorhythms are cyclic changes in an organism's growth, development, or functioning that are
66 driven by an internal biological 'clock' and synchronized through environmental cues
67 (Hastings, 1998). They have been linked to variations in human body temperature,
68 metabolism, testosterone production, ovulation, and rate of tooth eruption (Reinberg et al.,
69 1965; Little and Rimmel, 1971; Sothorn, 1974; Lee and Profitt, 1995; Garde et al., 2000).
70 Human tooth enamel retains evidence of periodic fluctuations that occur as enamel forming
71 cells (secretory ameloblasts) deposit mineralising protein matrix (Retzius, 1837; Asper,
72 1916). One of these fluctuations manifests as cross striations, which are incremental enamel
73 markings that correspond with a circadian rhythm (Schour and Poncher, 1937; Boyde, 1979,
74 1989; Risnes, 1986; Bromage 1991; Antoine et al., 2009; Lacruz et al., 2012; Zheng et al.,
75 2013). Another longer-period infradian rhythm leads to enamel Retzius lines (e.g., Dean,
76 1987; Risnes, 1990; Beynon, 1992). Retzius lines mark 'layers' of forming enamel that are
77 usually separated by six to 12 days of growth in human permanent teeth (**Fig. 1**), depending
78 upon the individual (Schwartz et al., 2001; Reid and Dean, 2006; Reid and Ferrell, 2006;
79 Mahoney, 2008). The Havers Halberg Oscillation (HHO) hypothesis proposes that Retzius
80 line periodicity (RP), the number of days between adjacent Retzius lines, is a manifestation of
81 a central infradian biorhythm that regulates the rate of bone cell proliferation and adult body
82 mass via metabolism, with links to life history traits, when compared between some
83 mammalian species (Bromage et al., 2009, 2012). Much less is known about the potential
84 role of this oscillation for human skeletal growth. Here, we extend our previous intra-specific
85 research into the HHO in which we established associations between human deciduous
86 enamel growth and RP (Mahoney et al., 2016; 2017). We construct and test predictions about
87 the relationship of RP to human permanent enamel thickness and the underlying daily rate
88 that enamel forms during the childhood years. We develop preliminary research conducted on
89 small samples of adult human bone (Bromage et al., 2016a), by assessing the relationship of

90 adult femoral length and the underlying density of bone maintenance cells (osteocytes) to RP.
91 Our goal is to explore the periodicity of the biorhythm against two hard tissues that form at
92 different but overlapping times during human ontogeny, within the context of the HHO.

93

94 **Enamel biorhythms of mammals and the Havers-Halberg Oscillation hypothesis**

95 Research into relationships between the periodicity of Retzius lines and somatic growth
96 commenced in the 1990's with those that suspected RP might relate to mammalian body size
97 (Dean, 1995; Dean and Scandrett 1996). Soon after, studies established a significant inter-
98 specific positive correlation between RP and average body mass in a selection of extant and
99 fossil mammals (Smith et al., 2003; Smith, 2008; Bromage et al., 2009). Not all primate
100 species followed this pattern (Hogg et al., 2015), and a lower rather than a higher RP related
101 to a larger estimated body mass for three fossil species (Schwartz et al., 2002, 2005; Le
102 Cabec et al., 2017). Further work reported inter-specific associations between the periodicity
103 of the biorhythm and the scheduling of life history traits for some primate species (Bromage
104 et al., 2012). The inter-specific HHO hypothesis developed out of these studies, and earlier
105 research on mammals (Mullender et al., 1996; Bromage et al., 2009, 2012).

106 The hypothesised biological 'clock' that regulates Retzius lines is unknown, but given
107 that RP subdivides into multiples' of daily intervals, the suprachiasmatic nucleus of the
108 hypothalamus (SCN) has been identified as one likely contender (Bromage et al., 2012). The
109 SCN is a source of circadian rhythmic activity in mammals (Richter, 1965; Ralph et al., 1990;
110 Sujino et al., 2003) and has a role in regulating metabolism via the pituitary gland (Weaver,
111 1998; Kalsbeek et al., 2011; Coomans et al., 2013). The HHO hypothesis drew upon this
112 biological pathway proposing that Retzius lines were a manifestation of a longer-period
113 oscillation stemming from the hypothalamus that employed SCN 'machinery' in its pathway
114 to stimulate pituitary secretions that linked to metabolism, body mass, and primate life
115 history traits (Bromage et al., 2012). Experimental research on domestic pig links aspects of

116 metabolism to RP (Bromage et al., 2016b). Further support for an inter-specific HHO is
117 provided by [some](#) mammalian species with slower metabolisms, and a larger body size
118 combined with a higher mean RP, relative to those with smaller body size (Bromage et al.
119 2009).

120

121 **Enamel biorhythms of humans and the Havers-Halberg Oscillation hypothesis**

122 The idea that [human](#) enamel growth might be controlled by an underlying biological ‘clock’
123 is not a new one, as the presence of daily cross-striations along enamel prisms implies that
124 secretory ameloblasts may be under circadian control via clock genes (maintainers of
125 circadian rhythms) [during amelogenesis](#) (Schour and Poncher, 1937; Bromage, 1991; Antoine
126 et al., 2009; Lacruz et al., 2012; Zheng et al., 2013). However, much less is known about the
127 potential role of the longer-period HHO for human enamel growth. Recently we reported
128 links between RP, the width of enamel ‘layers’ between adjacent Retzius lines, and [two](#)
129 [dimensional \(2D\)](#) average and relative enamel thickness of human deciduous maxillary
130 second molar crowns (dm^2) (Mahoney et al., 2016, 2017). We also identified an association
131 between RP and dm^2 paracone cusp formation time (Mahoney et al., 2016). The relationship
132 of RP to [daily enamel secretion rates](#) (DSRs) was however less clear. When RP and DSRs
133 were calculated for dm^2 in one homologous dental location and compared between
134 individuals there was a weaker association between these variables (Mahoney et al., 2017).
135 Prior to our research, enamel growth had not been considered within the context of the HHO
136 hypothesis. Based upon our data, we proposed that if RP is evidence of the HHO, an
137 underlying biorhythm that affects physiological systems (Bromage et al., 2009, 2012), then
138 its influence extends to enamel thickness and formation time of deciduous molar enamel, but
139 [was](#) less clearly associated with deciduous DSRs (Mahoney et al., 2017). Up until now, no
140 study has determined if there is a relationship between RP and human permanent molar
141 enamel thickness, or DSRs calculated for this tooth type.

142 Research on four adult humans hints at a negative correlation between adult stature and
143 RP (Bromage et al., 2016a). This [shift](#) away from the [positive correlation](#) reported in inter-
144 specific research [on mammalian species](#) (see above) is to be expected within the context of
145 the HHO. Inter-specifically, mammals with larger bodies tend to have an extended growth
146 period with slower rates of metabolism and associated cell proliferation, which is reflected by
147 a higher mean RP ([and thus slower oscillation of the biorhythm](#)) relative to smaller bodied
148 species (Mullender et al., 1996; Bromage et al., 2009). Within humans, the growth period is
149 constrained between birth and adulthood, so greater stature is achieved by ‘speeding up’ the
150 biorhythm (reducing the periodicity), and thus increasing skeletal metabolism or the rate of
151 cell proliferation (Bromage et al., 2016a). Thus, our current understanding of the HHO is
152 that inter-specific scaling trends between RP and body size may be associated with alterations
153 in the *duration* of development, whereas within humans, RP may relate to adult stature
154 through variation in growth *rates* (Bromage et al., 2009).

155 Preliminary support for [the HHO hypothesis within humans](#) is provided by a study of
156 bone osteocyte lacunar density (Ot.Dn) (Bromage et al., 2016a). Osteocytes are former
157 osteoblasts that become trapped as they finish producing bone matrix (e.g., Palumbo et al.,
158 1990). These cells have a complex functionality, that includes sensing mechanical ([shear or](#)
159 [strain](#)) forces applied to bone, which activates remodeling via the linked action of osteoblasts
160 and osteoclasts (e.g., Frost, 1987; Robling and Turner, 2002; Mullender et al., 2007;
161 Bonewald, 2007; Tatsumi et al., 2007; Noble, 2008); detecting and initiating micro-damage
162 repair (e.g., Verborgt et al., 2000; Herman et al., 2010); and in mineral homeostasis (e.g.,
163 Cullinane, 2002; [Teti and Zallone, 2009](#); Nakashima et al., 2011). The Ot.Dn of healthy bone
164 can also vary when compared to pathological bone (e.g., Mullender et al; 2005; van Hove et
165 al, 2009). In addition to these potential influences, Ot.Dn can correspond with body size,
166 whereby Ot.Dn of the mid-shaft femur from 12 adult humans scaled positively with final
167 attained adult stature (Bromage et al., 2016a). This scaling relationship suggests that the rate

168 of osteocyte proliferation is greater in taller adult individuals, which is consistent with the
169 hypothesised effect of the HHO on human body size.

170

171 **Research questions and predictions**

172 The background research provides a foundation from which to formulate four research
173 questions, and from these, predictions, that will be tested by calculating RP [from thin sections](#)
174 [of teeth](#) and comparing these values to measures of human skeletal growth. The research
175 questions are as follows:

176

177 ***Do daily enamel secretion rates correlate with RP in permanent molars?***

178 We have previously shown RP does not exert a consistent influence on the daily rate that
179 ameloblasts secrete structural matrix proteins as they increase the length of hydroxyapatite
180 crystallites in deciduous enamel (Mahoney et al., 2017). Instead, it seems more likely from
181 links we have reported, and by others, that the intra-specific HHO is related to the end state
182 of enamel growth (i.e., final enamel thickness) through formation time. These links
183 commence as ameloblasts secrete matrix for an additional number of days between adjacent
184 Retzius lines, leading to [thicker 'enamel layers' with higher RP's](#) (Mahoney et al., 2017).
185 [Layers become thicker because ameloblasts do not greatly alter their DSRs in outer lateral](#)
186 [enamel regions as RP increases, when compared between human molars from different](#)
187 [individuals. Thicker layers accumulate leading to greater average enamel thickness \(AET\) of](#)
188 [dm² crowns with higher RP's, relative to molars with lower RP's](#) (Mahoney et al. 2016).
189 Thicker crown enamel takes a longer period of time to form, for deciduous molars (Mahoney
190 2011), and permanent teeth (Dean et al., 2001). Formation time is correlated with RP, for
191 deciduous molar [paracone cusps](#) (Mahoney et al., 2016) and for permanent mandibular canine
192 lateral enamel (Reid and Ferrell, 2006). Thus, unlike the HHO intra-specific prediction for
193 body mass, [where RP links](#) to final attained adult stature through variation in growth rates
194 (Bromage et al., 2009), we suggest that RP is not strongly related to final enamel thickness

195 via daily enamel secretion rates (and is thus more likely to be related to enamel formation
196 time). Thus, we predict a weak association between RP and DSRs of permanent molars. To
197 examine the relationship between the circadian and the infradian rhythm in permanent
198 enamel, we separated out $n=15$ M1's from our sample, and calculated and compared RPs and
199 DSRs in one homologous location in outer lateral enamel of each crown.

200

201 ***Does permanent molar enamel thickness correlate with RP?***

202 [Two dimensional measurements of AET](#) from human dm^2 correlated positively with RP
203 (Mahoney et al., 2016). Assuming that RP is evidence of a biorhythm that affects multiple
204 physiological systems, including enamel growth, then we predict that its influence will extend
205 to permanent molar enamel thickness. To test this prediction we calculate [2D AET](#) and
206 enamel area (EA) for human permanent first (M1) and second molars (M2) [from thin](#)
207 [sections](#), and compare these values to RP's of the same teeth. Based upon findings for
208 deciduous molars, RP of permanent molars should scale positively with our measures of
209 enamel thickness.

210

211 ***Is adult femoral length correlated with RP?***

212 The intra-specific HHO predicts that greater adult height is achieved through a biorhythm
213 that is accelerated (Bromage et al. 2016a), with a shorter periodicity. To assess RP against
214 stature, we selected a sample of younger adult males with shorter femora, and compared these
215 to younger adult males with longer femora. We calculated RP for each male and compared
216 this value to his stature (reconstructed from femoral length). The femur has been used within
217 regression equations for the past fifty years to reconstruct stature (e.g., Trotter, 1970; [and see](#)
218 [methods](#)). We also compare RP to femoral length.

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225 *Is adult femoral length correlated with cortical bone osteocyte lacunae density?*

226 The intra-specific HHO predicts that taller humans (with longer femora) grow more rapidly
227 with a faster rate of osteocyte proliferation, relative to shorter individuals. Data for 12
228 individuals indicate these faster rates are then maintained as adults (Bromage et al., 2016a).
229 As Ot.Dn can sometimes vary with age (e.g., Mullender et al., 1996) we subdivided our entire
230 adult male sample into age groups and explored associations between Ot.Dn and stature,
231 within each group. We also assess Ot.Dn against adult femoral length, and against RP.

232

233 **Materials and Methods**

234 Our samples are human skeletons from one cemetery in Canterbury, England, that dates to
235 the early 16th century AD (Hicks and Hicks, 2001). Historical texts state that burials were
236 from a single lower socio-economic group that lived and worked in Canterbury and
237 represented non-catastrophic mortality (Somner 1703; Duncombe 1785; Brent 1879). We
238 have previously shown that the periodicity of the biorhythm can change in response to non-
239 specific pathology (Mahoney, et al., 2017). We limited this type of variation in our data by
240 only selecting skeletons and teeth without skeletal or radiographic signs of pathology,
241 drawing upon an extensive collection of accompanying radiographs that were produced at
242 Kent and Canterbury Hospital (Radiology Department) for any skeleton with suspected
243 trauma or pathology. Age-at-death is reconstructed for all skeletons; sex is reconstructed for
244 adults (see Methods). These collections are curated in the Skeletal Biology Research Centre,
245 University of Kent, UK. All sectioning adhered to the British Association of Biological
246 Anthropology and Osteoarchaeology code of practice (2014). No permits were required for
247 this study as these are archaeological samples from before the 19th Century AD.

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253 **Samples and the chronology of skeletal growth**

254 We selected three samples. Throughout, RP is calculated for lateral enamel of permanent M1
255 and M2. Lateral enamel of these tooth types forms between [approximately](#) 1.5 to 5.7 years of
256 age (Reid and Dean, 2006). Sample sizes varied depending upon the variables examined and
257 are given in the corresponding tables. One tooth ([either M1 or M2](#)) represents one individual.
258 Raw data is [available](#) in Supporting Information.

259

260 **a)** The first sample was juveniles ($n=40$). We assessed RP against [daily](#) enamel secretion
261 rates, and against enamel thickness of the same molars. We chose juveniles ([<8yrs of age](#)
262 [for M1's; <13yrs for M2's](#)) because enamel is often worn in adults, and this would have
263 affected our measurements of 2D AET and EA. [Daily enamel secretion rates of M1 and](#)
264 [M2 are a measure of the rate that ameloblasts previously deposited matrix during the](#)
265 [secretory phase of enamel growth in the childhood years. Average enamel thickness and](#)
266 [EA of M1 and M2 are a measure of the end state of the secretory stage of enamel growth](#)
267 [that is attained in childhood.](#)

268

269 **b)** The second sample was young adult males, aged between 18 to 34yrs ($n=27$). We
270 assessed RP of their M1 or M2 (representing their childhood years), against their femoral
271 cortical bone osteocyte lacunar density, and final attained adult stature. Osteocyte
272 lacunar density in adult cortical bone likely represents a combination of lamellae
273 deposited during later ontogeny and in adulthood. Final attained adult stature is the [end](#)
274 [state](#) of linear growth of long bones via endochondral ossification over the course of
275 postnatal development [from birth to adulthood](#). In this study, we measure adult femoral
276 length as a proxy for attained adult stature. The slight occlusal wear of some molars did
277 not affect our calculation of RP in lateral enamel, which is located cervical to wear on
278 the occlusal surface. We did not include older adults because of their greater enamel
279 wear.

280 c) The third sample was adult males, subdivided into two age groups (younger males 18-
281 34yrs, $n=28$; older males 35-50yrs, $n=94$). We assessed femoral cortical Ot.Dn against
282 their estimated stature, and femoral length.

283

284 **Sample preparation for histology**

285 We used standard histological techniques (Bancroft and Gamble, 2008; Mahoney, 2008;
286 Miskiewicz, 2016). Each tooth was embedded in polyester resin to reduce the risk of
287 splintering while sectioning. Using a diamond-wafering blade (Buehler® IsoMet 4000
288 precision saw), buccal-lingual sections captured the paracone and protocone of maxillary
289 molars and the protoconid and metaconid of mandibular permanent molars. Each section was
290 mounted on a microscope slide, lapped using a graded series of grinding pads (Buehler®
291 Eco-Met 300) to reveal incremental lines, polished with a $0.3\mu\text{m}$ aluminum oxide powder
292 (Buehler® Micro-Polish II), placed in an ultrasonic bath to remove surface debris, dehydrated
293 through a series of alcohol baths, cleared (Histoclear®), and mounted with a coverslip using a
294 xylene-based mounting medium (DPX®).

295 Dry, un-decalcified bone measuring 1cm in depth was removed from the posterior
296 femoral mid-shaft cortex using a drill (Dremel Rotary®) with a circular metal blade (**Fig. 2**).
297 Only the posterior portion of the femoral diaphysis was used in order to keep the overall
298 integrity of the femur preserved for future research purposes. The bone was embedded in
299 epoxy resin, reduced in thickness (Buehler® IsoMet 4000 precision saw), ground, polished,
300 and cover-slipped following the same procedures used to embed and prepare the teeth. Thin
301 sections measured approximately $100\mu\text{m}$ in depth.

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309 Retzius line periodicity

310 Using a high-resolution microscope (Olympus® BX51), each section was examined at
311 magnification (10-60x). Images were captured with a microscope digital camera (Olympus®
312 DP25) and analyzed in CELL® Live Biology imaging software. We counted the number of
313 cross-striations along a prism between several adjacent Retzius lines in outer lateral enamel
314 of M1 and M2 to determine the number of days between two adjacent Retzius lines. For
315 twelve thin sections, cross-striations were not clearly visible and continuous [along prisms](#)
316 between adjacent Retzius lines. For these twelve sections, we divided the distance between
317 several adjacent Retzius lines by local mean daily secretion rates (e.g., [Schwartz et al., 2001](#);
318 [Mahoney et al., 2007](#); [Lacruz et al., 2008](#)). We did not include these sections in the analysis
319 of RP and secretion rates. Retzius periodicity was recorded by SC and PM. After they had
320 finished recording all slides they conducted an inter-observer error test. The test revealed one
321 difference in RP calculations between the two observers. This slide was removed from the
322 study.

323

324 Enamel thickness

325 The 2D AET in mm was calculated by dividing the area of the enamel cap (EA) by the length
326 of the dentin-enamel junction (DEJ), which provides the average straight-line distance
327 between the DEJ and outer enamel surface (Martin, 1983, 1985). EA is given in mm².

328

329 Enamel daily secretion rates

330 Secretion rates in μm per day were calculated for outer lateral enamel in the same region that
331 we recorded RP (ie., avoiding inner and mid enamel regions as DSRs can vary from one
332 region to the next within a crown: [Lacruz and Bromage 2006](#)). Rates were measured along
333 the long axis of an enamel prism. A distance corresponding to five days of enamel secretion
334 was measured, and then divided by five to yield a mean daily rate. The procedure was
335 repeated a minimum of six times in each region, which allowed a grand mean value and

336 standard deviation (SD) to be calculated. The grand mean value was compared to RP
337 calculated in the same enamel region.

338

339 **Osteocyte lacunae density**

340 We use Ot.Dn as a proxy for the rate of (past) femoral cortical bone cell proliferation.
341 Osteocyte lacunae density data were collected as part of a PhD project (Miszkievicz, 2014).
342 We selected femoral Ot.Dn, rather than osteon population density, so that we could directly
343 test prior research (see above). Osteocyte lacunae density is significantly correlated with
344 osteon population density in this skeletal sample (Miszkievicz, 2016). Exploring associations
345 between Ot.Dn and age, or at the initiation of remodeling (e.g., Metz et al., 2003), were not
346 aims of this study.

347 Using a high-resolution microscope (Olympus® BX51, and Olympus DP25 microscope
348 camera) osteocyte lacunae were counted within secondary osteonal bone and interstitial bone.
349 Ot.Dn were counted from a maximum of six main regions of interest (ROI; mag =10X, 2.44
350 mm²) positioned adjacent to the periosteum, and sub-divided into smaller ROIs (mag =40X,
351 0.13 mm²) (**Fig. 2**). See Miszkievicz (2016) for a detailed methodology of ROI's. Using
352 CELL® Live Biology Imaging software, all visible osteocyte lacunae (including cavities
353 which appeared “empty” or transparent) were counted using a “touch count” tool (identical in
354 premise to the “point count technique” recommended by Parfitt, 1983). Densities were
355 calculated by dividing the total number of osteocyte lacunae by the area of bone examined (in
356 mm²). We acknowledge that automated methods of osteocyte lacunae detection are available,
357 and ideally a whole long bone cross-section should be examined (e.g. Hunter and Agnew,
358 2016). However, those techniques are better suited to fresh or “recent” bone with excellent
359 microstructural preservation. Given the archaeological background (localised diagenetic
360 alteration of micro-anatomy) of our samples, there needed to be flexibility in our ROI
361 selection procedures. This is because the ROI would sometimes have to be moved

362 fractionally to avoid an area of diagenesis or one that was affected by taphonomy. Clear
363 differences in osteocyte lacunar densities were observed across the sample (see Fig. 2).

364

365 **Stature estimation and femoral length**

366 Femoral length data were previously included in robusticity index calculations as part of
367 another project (Miszkiwicz and Mahoney, 2016), but correlations between Ot.Dn and
368 stature/femur length are examined here for the first time. The maximum length of each femur
369 was measured by placing it flat on an osteometric board, in its anatomical position, with the
370 posterior femoral aspect facing down. Femoral length was measured from the most superior
371 surface of the femoral head to the most distal surface of the medial condyle (Buikstra and
372 Ubelaker, 1994). Standard, and most commonly used formulae for reconstructing stature in
373 skeletal remains were used (Trotter, 1970; White et al., 2011). These were specific to sex and
374 appropriate for individuals of European descent. Male stature was estimated using the
375 regression equation: $2.38 \times \text{femur maximum length in cm} + 61.41 (+/- 3.27)$ (Trotter, 1970;
376 White et al., 2011).

377

378 **Sex determination and age-at-death**

379 Sex determination was carried out using multiple standard methods to increase the accuracy
380 of the determination. We relied upon standard morphological characteristics of the pelvis and
381 cranium. The pelvic methods were based upon 25 morphological characteristics of the human
382 pelvis taken from Schwartz (1995), Ferembach et al., (1980), Krogman and Iscan (1986) and
383 Phenice (1969). Cranial features included the mastoid process, supraorbital margin, mental
384 eminence, and nuchal crest (Buikstra and Ubelaker, 1994). When determinations from cranial
385 and pelvic features conflicted, priority was given to the pelvic criteria (White et al., 2011). In
386 the analyses, 'probable males' were classified as male.

387 Age was estimated from age-specific morphology of the pubic symphysis, and the
388 auricular surface of the pelvis (e.g., Meindl et al., 1985; Lovejoy et al., 1985). Two age
389 categories were constructed: younger adult males, 18-34 years; older adult males 35-50 years.

390

391

392 **Analyses**

393 Data were analyzed in IBM SPSS® 22 (2014). Each variable was log-transformed. A one
394 sample Kolmogorov-Smirnov test indicated that the distribution of the data for each variable
395 was normal. Data from right and left femora (one femur was selected from each individual,
396 and either the right or left depending upon preservation) were pooled. We analyze the data
397 using linear regression statistics. In Tables 1-2 we present the r^2 value (coefficient of
398 determination) which measures the proportion of explained variation, and we also show the r
399 value (correlation coefficient) which measures the strength and direction of the relationship
400 between variables. The residual, presented as a percentage in the Tables, is the error not
401 explained by the regression equation.

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414 **Results**

415 **Retzius line periodicity, enamel thickness and secretion rates**

416 Regression statistics are in Table 1. Corresponding data for the sample of juveniles is
417 available in Supporting Information Table S1. When data for all tooth types are combined,
418 the enamel areas and AET of permanent molar crowns were significantly and positively
419 related with RP, increasing from minimum values that were associated with an RP of 6 days
420 to maximum values that were associated with RP's of 10 and 11 days respectively (**Fig 3a.**
421 **Fig 4a**). When subdivided into either M1's or M2's and re-analyzed, RP was significantly
422 related to EA and AET (Table 1). When further subdivided into upper or lower molars, RP
423 was significantly related to EA (**Fig 4b-d**). AET was also significantly related to RP for each
424 upper and lower molar type, except lower M2 where this relationship approached
425 significance ($r^2=0.287$; $p=0.072$).

426 When 15 permanent first molars were separated from the sample, and RPs and DSRs
427 were measured and compared between the molars in one homologous location in outer lateral
428 enamel of each crown, there was no consistent or significant association with the periodicity
429 of Retzius lines.

430

431 **Retzius line periodicity, femoral length and osteocyte lacunae density**

432 Regression statistics are in Table 2. The corresponding data sets for younger and older male
433 adults are available in Supporting Information Tables S2 and Table S3. Estimated stature
434 (and femoral length) was significantly and negatively related with RP (**Fig. 3b**). The density
435 of osteocyte lacunae did not relate significantly with RP (Table 2). Osteocyte lacunae density
436 was not significantly related to femoral length or stature for younger males (Table 2). There
437 was a weak relationship between these latter variables that approached significance in older
438 males though the residual was high ($r^2=0.030$; $p=0.089$).

439

440 **Discussion**

441 This study builds upon our previous work that examined relationships of RP to human
442 deciduous molar enamel growth, and extends preliminary research into associations between
443 RP and human adult femoral cortical bone growth (Bromage et al., 2016a; Mahoney et al.,
444 2016, 2017). We examined the relationship of permanent molar daily enamel secretion rates
445 to RP, and of osteocyte proliferation to RP. We find limited evidence for either of these
446 relationships, but did find stronger evidence of linkages between RP, permanent molar
447 enamel thickness, and stature.

448

449 **Retzius line periodicity, enamel thickness and secretion rates**

450 Our data support the prediction that the **periodicity of the** biorhythm is associated with
451 enamel thickness when considered within a smaller intra-specific scale, within humans.
452 However, as with deciduous molars (Mahoney, et al., 2016), RP was more weakly associated
453 with DSRs, when compared between permanent molars from different individuals. **Therefore,**
454 **even though RP is calculated by a count of cross striations, variation in the biorhythm is not**
455 **always associated with the amount of matrix deposited by ameloblasts in 24 hour periods**
456 **(Fig. 5).** Instead, it seems likely that RP can link to the final enamel thickness of a human
457 crown through formation time. RP is related to the time taken to form part of a deciduous
458 and permanent tooth crown (Reid and Ferrell, 2005 Mahoney et al., 2016), and formation
459 time is related to human enamel thickness (Dean et al., 2001; Mahoney 2011). **Thus, inter-**
460 **individual variation in the periodicity of the biorhythm may have a clearer association with**
461 **final enamel thickness through the duration, rather than the daily rate of enamel growth. More**
462 **work is needed to understand if and how these developmental mechanisms change within a**
463 **species (Fig 5).**

464 The proposal that aspects of enamel growth are controlled by a long-period biological
465 'clock' with an infradian rhythm, whether it is the HHO via the SCN of the brain, or a

466 different ‘peripheral’ independent ‘clock’ (Hastings, 1998), or even more than one ‘clock’
467 (Newman and Poole, 1974, 1993), is a hypothesis. Our data for human permanent teeth, and
468 deciduous teeth (Mahoney et al., 2016, 2017), provide support for this hypothesis. The
469 infradian rhythm (reflected by RP) appears to have an association with **final** enamel thickness
470 of a crown, but is inconsistently related to the daily *amount* of enamel secreted by
471 ameloblasts as these cells respond to a circadian rhythm (reflected by cross striations). The
472 infradian rhythm likely has a systemic origin, as RP can alter within a single crown in
473 response to non-specific pathology (Mahoney et al., 2017). The longer-period rhythm is
474 intrinsic to enamel growth, not only relating to **final** enamel thickness, but also the
475 microstructural components of enamel (prisms) which **can be** reduced in size, or have an
476 altered morphology when associated with Retzius lines (Risnes, 1990,1998; Li and Risnes,
477 2004). Perhaps therefore, the infradian rhythm periodically modifies ameloblast metabolism,
478 interfering with enamel secretion of ameloblasts, leading to the altered prism structure **that**
479 **can be associated with Retzius lines**.

480 There is substantial residual in the relationship of RP to enamel thickness (Table 1), as
481 even the strongest correlations explain just over half of the variation in our data. So, there are
482 other factors operating as well. Enamel thickness is a product of several mechanisms, other
483 than those considered here, such as the number of active ameloblasts and their life spans
484 (Grine and Martin, 1988; Macho, 1995). We have only considered the rate that enamel grows
485 in thickness, but whether the rate that enamel crowns extend in height (**enamel extension**
486 **rates**), as epithelium cells differentiate into pre-ameloblasts down along the dentin-enamel
487 junction (DEJ), is linked to RP, has yet to be determined. Guatelli-Steinberg and colleagues
488 (2012) have already shown links between DEJ lengths and lateral enamel formation time. As
489 RP is correlated with enamel formation times (Reid and Ferrell, 2006; Mahoney et al., 2016),
490 it would seem possible that extension rates can relate to RP.

491

492 **Retzius line periodicity, femoral length, and osteocyte lacunae density**

493 Our data support the intra-specific HHO prediction that taller adults (with longer femora)
494 have a lower RP (Bromage et al., 2016a). Thus, the biorhythm oscillates with a faster
495 periodicity in taller humans, compared to those with shorter femora. However, we found less
496 support for the prediction that taller adults maintain significantly faster rates of femoral
497 osteocyte proliferation, relative to shorter adults. Osteocyte density did not relate to stature
498 or femoral length amongst our sample of young adult males, though it appeared to be
499 trending towards significance with a high residual amongst older males (Table 2, and
500 footnotes). Neither did RP relate to Ot.Dn in a small sample. Thus, the biorhythm is
501 significantly linked to adult stature, but neither the biorhythm nor stature are linked to
502 osteocyte proliferation of the femur.

503 Osteocytes have a complex functionality (see Introduction) that, in addition to potential
504 influences of body size, probably influences their distribution in cortical bone leading to
505 significant variation in their numbers across the femoral shaft (e.g., Carter et al. 2013, 2014).
506 For example, an anatomical region can adapt to mechanical loading, adding and removing
507 new bone tissue in response to loading or disuse (Wolff, 1892; Robling et al., 2001; Burr et
508 al., 2002). Our osteocyte lacunae data are from one anatomical region, the posterior femoral
509 mid-shaft cortex, and just the sub-periosteal pocket, which is where new bone is usually
510 deposited in response to excessive load (Robling et al., 2006). There is substantial inter-
511 individual variation in Ot.Dn values from this region (younger adults range between 394.87
512 and 1307.69; middle aged adults between 305.77 and 1255.13). Some of this variation in
513 Ot.Dn probably reflects differences in femoral mechanical loading between individuals, as
514 some adults in our sample would have been employed in the physically demanding
515 occupations that were typical of lower socio-economic lifestyles in medieval Canterbury
516 (Miszkievicz and Mahoney, 2016).

517

518 Variation in adult stature is not strongly related to differences among individuals in the
519 rate of femoral osteocyte proliferation, but it is related to RP (Table 2). This finding makes
520 sense if RP is linked to the duration in which stature is attained. [Pre-pubertal](#) growth velocity
521 differences can underlie adult stature differences within some populations (e.g., Gasser 1990;
522 Gasser et al., 2001), but not all populations. Instead, the timing of the pubertal growth spurt
523 can contribute to the *age* adult height is attained, for females compared to males (e.g.,
524 Tanner, 1990; Roche, 1992; Gasser et al., 2000), and within the sexes (Hägg and Taranger,
525 1991; Baer et al., 2006). Late maturing Swedish boys continued to grow between 18 to 25
526 years of age, attaining significantly greater growth in height during this period and a greater
527 final stature, compared to early maturing boys whose height increased only slightly after age
528 18 (Hägg and Taranger, 1991). The Nurses' Health Study (II) in the USA, which is based
529 upon large sample sizes, indicates that females with delayed puberty are older when they
530 attain their final and greater adult height, compared to females with a shorter adult stature
531 (Baer et al., 2006). Further research might explore potential linkages between [the frequency](#)
532 [that the biorhythm oscillates and the age that adult stature is attained, as the duration of the](#)
533 [growth period may be an important link to RP for aspects of both enamel and bone growth.](#)
534 [Variation in growth velocities \(and Ot.Dn\) compared to RP amongst children should also be](#)
535 [examined.](#)

536

537 **The biorhythm of human skeletal growth**

538 The direction of the correlation between RP and enamel thickness is positive, but negative
539 when RP is related to stature. [Our data implies that a child from Canterbury with a slow](#)
540 [biorhythm between birth and five years of age attained thicker deciduous \(Mahoney et al.,](#)
541 [2016\)](#) and permanent molar enamel, compared to another child with a faster biorhythm that
542 developed thinner enamel (**Fig. 6**). A child from the same population [with a fast biorhythm](#)
543 attained a greater adult stature. These findings imply that the biorhythm may coordinate

544 aspects of human skeletal growth, perhaps by increasing the duration of crown enamel
545 growth early on in ontogeny leading to thicker enamel, at the expense of subsequent femoral
546 growth in length and attained adult height. Alternatively, the change in the direction of the
547 correlation may reflect a biorhythm that does not remain constant within an individual. We
548 have previously shown that RP can change within an individual at the end of the first post-
549 natal year (Mahoney et al., 2017). The change in RP, from deciduous to permanent molars,
550 suggests that the biorhythm produces a sequence of RPs for an individual, rather than a static
551 value. In the present study, we focused on permanent M1s and M2s, whose enamel forms
552 between birth and five to six years of age (Reid and Dean, 2006). It seems likely that RP
553 remains constant during this age-range within an individual, as comparisons between small
554 samples of permanent anterior teeth that form at about the same time as permanent molars
555 (FitzGerald, 1998), as well as comparisons between molar types within four individuals (Reid
556 et al., 1998), reveal no variation in RP. Whether the periodicity of the biorhythm changes in
557 humans beyond 11 years of age, after third molar crown enamel has formed, is unknown.
558 Therefore, the relationship we describe, between RP during the early childhood years and
559 adult stature, might not describe this relationship in later ontogeny, if RP changes closer to
560 adulthood, or, if bone modifies its response to the biorhythm with age.

561

562 **Conclusion**

563 We examined the relationship of enamel secretion rates to evidence of a biorhythm retained
564 in human teeth as Retzius line periodicity, and of cortical bone osteocyte proliferation to
565 Retzius periodicity. We found only limited evidence for either of these relationships, but we
566 did find stronger evidence of linkages between RP and permanent molar enamel thickness
567 (end state of enamel growth), and RP and final adult stature (end state of linear growth in
568 long bones). Our findings develop the intra-specific HHO hypothesis suggesting that the
569 biorhythm has a role in human skeletal growth and the development of more than one hard
570 tissue.

571 **Conflict of Interest**

572 The authors have no conflict of interest to declare.

573

574 **REFERENCES**

575 **Antoine D, Hillson S, Dean MC** (2009) The developmental clock of dental enamel: a test for the
576 periodicity of prism cross striations and an evaluation of the likely sources of error in histological
577 studies of this kind. *J Anat* **214**, 45–55.

578

579 **von Asper H** (1916) *Über die Braune Retzius'sche Parallelstreifung im Schmelz der menschlichen*
580 *Zähne*. PhD thesis: Universität Zurich.

581

582 **Baer HJ, Rich-Edwards JW, Colditz GA, Hunter DJ, Willett WC, Michels KB** (2006) Adult
583 height, age at attained height, and incidence of breast cancer in premenopausal women. *Int J Cancer*
584 **119**, 2231–2235.

585

586 **Bancroft JD, Gamble M** (2008) *Theory and practice of histological techniques*. Elsevier Health
587 Sciences.

588

589 **Beynon AD** (1992) Circaseptan rhythms in enamel development in modern humans and Plio-
590 Pleistocene hominids. In *Structure, Function and Evolution of Teeth* (eds Smith P, Tchernov E), pp.
591 295–309. London: Freund Publishing House Ltd.

592

593 **Boas F** (1935) The tempo of growth of fraternities. *Proc Nat Acad Sci* **21**, 413–418.

594

595 **Bonewald LF** (2007) Osteocytes as dynamic multifunctional cells. *Ann NY Acad Sci* **1116**, 281–290.

596

597 **Boyde A** (1979) Carbonate concentration, crystal centres, core dissolution, caries, cross striation,
598 circadian rhythms and compositional contrast in the SEM. *J Dent Res* **58**, 981–983.

599

600 **Boyde A** (1989) Enamel. In: *Teeth. Handbook of microscopic anatomy* (eds Berkovitz BKB, Boyde
601 A, Frank RM, et al.), pp. 309–473. Berlin: Springer-Verlag.

602

603 **Brent J** (1879) *Canterbury in the olden time*. London: Simpkin, Marshall and Co.

604

605 **Bromage TG** (1991) Enamel incremental periodicity in the pigtailed macaque: a polychrome
606 fluorescent labelling study of dental hard tissues. *Am J Phys Anthropol* **86**, 205–214.

607

608 **Bromage TG, Hogg RT, Lacruz RS. et al.** (2012) Primate enamel evinces long period biological
609 timing and regulation of life history. *J Theor Biol* **305**, 131–144.

610

611 **Bromage TG, Idaghdour Y, Lacruz RS. et al.** (2016b) The swine plasma metabolome chronicles
612 'many days' biological timing and functions linked to growth. *PLoS One* e0145919.

613

614 **Bromage TG, Juwayeyic YM, Katrisa JA. et al.** (2016a) The scaling of human osteocyte lacuna
615 density with body size and metabolism. *C R Palevol* **15**, 32–39.

616

617 **Bromage TG, Lacruz RS, Hogg R. et al.** (2009) Lamellar bone is an incremental tissue reconciling
618 enamel rhythms, body size, and organismal life history. *Calcif Tiss Int* **84**, 388–404.

619

620 **Buikstra JE, Ubelaker DH** (1994) *Standards for data collection from human skeletal remains*.
621 Fayetteville: Arkansas Archaeology Survey.

622

623 **Burr DB, Robling AG, Turner CH** (2002) Effects of biomechanical stress on bones in animals.
624 *Bone* **30**(5), 781–6.

- 625 **Le Cabec A, Dean MC, Begun DR** (2017) Dental development and age at death of the holotype of
626 *Anapithecus hernyaki* (RUD 9) using synchrotron virtual histology. *J Hum Evol* **108**, 161-175.
627
- 628 **Carter Y, Suchorab JL, Thomas CD, Clement JG, Cooper DM** (2014) Normal variation in
629 cortical osteocyte lacunar parameters in healthy young males. *J Anat* **225**, 328-36.
630
- 631 **Carter Y, Thomas CD, Clement JG, Peele AG, Hannah K, Cooper DM** (2013) Variation in
632 osteocyte lacunar morphology and density in the human femur--a synchrotron radiation_
633 study. *Bone* **52**(1), 126-32.
634
- 635 **Cullinane DM** (2002) The role of osteocytes in bone regulation: mineral homeostasis versus
636 mechanoreception. *J Musc Neuronal Interact* **2**, 242-4.
637
- 638 **Coomans CP, van den Berg SA, Houben T, van Klinken JB, van den Berg R, Pronk AC,
639 Havekes LM, Romijn JA, van Dijk KW, Biermasz NR, Meijer JH** (2013) Detrimental effects of
640 constant light exposure and high-fat diet on circadian energy metabolism and insulin sensitivity.
641 *FASEB J* **27**, 1721-1732.
- 642 **Dean MC** (1987). Growth layers and incremental markings in hard tissues; a review of the literature
643 and some preliminary observations about enamel structure in *Paranthropus boisei*. *J Hum Evol* **16**,
644 157-172.
645
- 646 **Dean MC** (1995) The nature and periodicity of incremental lines in primate dentine and their
647 relationship to periradicular bands in OH 16 (*Homo habilis*). In *Aspects of Dental Biology:
648 Paleontology, Anthropology and Evolution* (ed Moggi-Cecchi J), pp. 239-265. Florence: International
649 Institute for the Study of Man.
650
- 651 **Dean MC, Leakey MG, Reid DJ. et al.** (2001) Growth processes in teeth distinguish modern
652 humans from *Homo erectus* and earlier hominins. *Nature* **414**, 628-631.
653
- 654 **Dean MC, Scandrett AE** (1996) The relation between long-period incremental markings in dentine
655 and daily cross-striations in enamel in human teeth. *Arch Oral Biol* **41**, 233-241.
656
- 657 **Duncombe J** (1785) The history and antiquities of the three archiepiscopal hospitals and other
658 charitable foundations at and near Canterbury. *Bibliotheca Topographica Britannica* No XXX,
659 London.
660
- 661 **Ferembach D, Schwindevky I, Stoukal M** (1980) Recommendations for age and sex diagnoses of
662 skeletons. *Journal of Human Evolution* **9**: 517-549.
663
- 664 **FitzGerald CM** (1998) Do enamel microstructures have regular time dependency? Conclusions from
665 the literature and a large scale study. *J Hum Evol* **35**, 371-386.
666
- 667 **Frost HM** (1987) Bone "mass" and the "mechanostat": a proposal. *Anat Rec* **219**, 1-9.
668
- 669 **Gasser T** (1990) A method for determining the dynamics and intensity of average growth. *An Hum
670 Biol* **17**, 459-457.
671
- 672 **Gasser T, Sheehy A, Molinari L, Largo RH** (2000) Sex dimorphism in growth. *An Hum Biol* **27**,
673 187-197.
674
- 675 **Gasser T, Sheehy A, Molinari L, Largo RH** (2001) Growth processes leading to a large or small
676 adult height. *An Hum Biol* **28**, 319-327.
677

- 678 **Garde AH, Hansen AM, Skovgaard LT, Christensen JM** (2000). Seasonal and biological variation
679 of blood concentrations of total cholesterol, dehydroepiandrosterone sulfate, hemoglobin A(1c), IgA,
680 prolactin, and free testosterone in healthy women. *Clin Chem* **46**(4), 551–559.
681
- 682 **Grine FE, Martin LB** (1988) Enamel thickness and development in Australopithecus and
683 Paranthropus. In *The evolutionary history of the robust Australopithecines* (ed Grine FE), pp. 3–
684 42. New York: Aldyne de Gruiter.
685
- 686 **Guatelli-Steinberg D, Floyd BA, Dean MC, Reid DJ** (2012) Enamel extension rate patterns in
687 modern human teeth: two approaches designed to establish an integrated comparative context for
688 fossil primates. *J Hum Evol* **63**, 475–486.
689
- 690 **Hägg U, Taranger J** (1991) Height and height velocity in early, average and late maturers followed
691 to the age of 25: a prospective longitudinal study of Swedish urban children from birth to adulthood.
692 *Ann Hum Bio* **18**(1), 47–56.
693
- 694 **Hasting M** (1998) The brain, circadian rhythms, and clock genes. *Brit Med J*, **317**, 1704–1707.
695
- 696 **Herman BC, Cardoso L, Majeska RJ, Jepsen KJ, Schaffler MB** (2010) Activation of Bone
697 Remodeling after Fatigue: Differential Response to Linear Microcracks and Diffuse Damage. *Bone*
698 **47**(4), 766–772.
699
- 700 **Hicks M, Hicks A** (2001) *St. Gregory's Priory, Northgate, Canterbury Excavations 1988–1991*.
701 Canterbury Archaeological Trust Ltd: Volume II.
702
- 703 **Hogg RT, Godfrey LR, Schwartz GT, Dirks W, Bromage TG** (2015) Lemur biorhythms and life
704 history evolution. *PLoS ONE* **10**(8), e0134210. doi:10.1371/journal.pone.0134210
705
- 706 **Hunter RL, Agnew AM** (2016) Intraskkeletal variation in human cortical osteocyte lacunar density:
707 Implications for bone quality assessment. *Bone Reports* **5**, 252–261.
708
- 709 **Kalsbeek A, Scheer FA, Perreau-Lenz S. et al.** (2011) Circadian disruption and SCN control of
710 energy metabolism. *FEBS Letters* **585**, 1412–1426.
711
- 712 **Krogman WM, Iscan MY** (1986) *The human skeleton in forensic medicine* (2nd ed). Springfield:
713 Charles C Thomas. P 156–162
714
- 715 **Lacruz RS, Dean MC, Ramirez-Rozzi F, Bromage TG** (2008) Megadontia, striae periodicity and
716 patterns of enamel secretion in Plio-Pleistocene fossil hominins. *J Anat.* **213**, 148–158
717
- 718 **Lacruz RS, Bromage TG** (2006) Appositional enamel growth in molars of South African fossil
719 hominids. *J Anat* **209**, 13–20.
720
- 721 **Lacruz RS, Hacia JG, Bromage TG. et al.** (2012) The circadian clock modulates enamel
722 development. *J Biol Rhyth* **27**, 237–245.
723
- 724 **Lee CF, Profitt WR** (1995) The daily rhythm of tooth eruption. *Am J Orthod Dentofacial Orthop*
725 **107**(1), 38–47.
726
- 727 **Li C, Risnes S** (2004) SEM observations of Retzius lines and prism cross-striations in human dental
728 enamel after different acid etching regimes. *Arch Oral Biol* **49**, 45–52.
729
- 730 **Little MA, Rummel JA** (1971) Circadian variations in thermal and metabolic responses to heat
731 exposure. *J Appl Physiol* **31**(4), 556–561.
732

- 733 **Lovejoy CO, Meindl RS, Pryzbeck TR, Mensforth TJ** (1985) Chronological metamorphosis of the
734 auricular surface of the ilium: A new method for the determination of age at death. *Am J Phys*
735 *Anthropol* **68**, 15-28.
736
- 737 **Macho G** (1995) The significance of hominid enamel thickness for phylogenetic and life-history
738 reconstruction. In: *Aspects of Dental Biology: Palaeontology, Anthropology and Evolution* (ed.
739 Moggi-Cecchi J), pp. 51–68. Florence: International Institute for the Study of Man.
740
- 741 **Mahoney P** (2008) Intraspecific variation in M1 enamel development 601 in modern humans:
742 implications for human evolution. *J Hum Evo* **55**, 131–147.
743
- 744 **Mahoney P** (2011) Human deciduous mandibular molar incremental enamel development. *Am J Phys*
745 *Anthropol*, 144, 204–214.
746
- 747 **Mahoney P** (2013) Testing functional and morphological interpretations of enamel thickness along
748 the deciduous tooth row in human children. *Am J Phys Anthropol* **151**, 518–525.
749
- 750 **Mahoney P** (2015) Dental fast track: prenatal enamel growth, incisor eruption, and weaning in human
751 infants. *Am J Phys Anthropol* **156**, 407–421.
752
- 753 **Mahoney P, Miszkiewicz JJ, Pitfield R, Schlecht SH, Deter C, Guatelli-Steinberg D** (2016)
754 Biorhythms, deciduous enamel thickness, and primary bone growth in modern human children: a test
755 of the Havers-Halberg Oscillation hypothesis. *J Anat* **228**, 919–928.
756
- 757 **Mahoney P, Miszkiewicz JJ, Pitfield R, Deter C, Guatelli-Steinberg D** (2017) Enamel biorhythms
758 of humans and great apes: the Havers-Halberg Oscillation hypothesis reconsidered. *J Anat* **230**, 272-
759 281.
760
- 761 **Mahoney P, Smith TM, Schwartz GT, Dean MC, Kelley J** (2007) Molar crown formation in the
762 Late Miocene Asian hominoids, *Sivapithecus parvada* and *Sivapithecus indicus*. *J Human Evol* **53**,
763 61-68
764
- 765 **Martin LB** (1983) Relationships of the later Miocene Hominoidea. Ph.D. Dissertation, University
766 College London.
767
- 768 **Martin LB** (1985) Significance of enamel thickness in hominoid evolution. *Nature* **314**, 260-263.
769
- 770 **Meindl RS, Lovejoy CO, Mensforth RP, Walker RA** (1985) A revised method of age
771 determination using the os pubis, with a review and tests of accuracy of other current methods of
772 pubic symphyseal aging. *Am J Phys Anth* **68**, 29–45.
773
- 774 **Metz LN, Martin RB, Turner AS** (2003) Histomorphometric analysis of the effects of osteocyte
775 density on osteonal morphology and remodeling. *Bone* **33**(5), 753-759.
776
- 777 **Miszkiewicz JJ** (2014) Ancient human bone histology and behaviour. PhD Dissertation. University
778 of Kent: Canterbury, UK.
779
- 780 **Miszkiewicz JJ** (2016) Investigating histomorphometric relationships at the human femoral midshaft
781 in a biomechanical context. *J Bone Min Metab* **34**(2), 179-192.
782
- 783 **Miszkiewicz JJ, Mahoney P** (2016) Ancient human bone microstructure in medieval England:
784 comparisons between two socio-economic groups. *Anat Rec* **299**, 42-59.
785
- 786 **Mullender MG, Huiskes R, Versleyen H, Buma P** (1996) Osteocyte Density and
787 Histomorphometric Parameters in Cancellous Bone of the Proximal Femur in Five Mammalian
788 Species. *J Orth Res* **14**, 972-979.

- 789 **Mullender MG, Tan SD, Vico L, Alexandre C, Klein-Nulend J** (2005). Differences in osteocyte
790 density and bone histomorphometry between men and women and between healthy and osteoporotic
791 subjects. *Calcif Tiss Int* **77**(5), 291-296.
- 792 **Nakashima T, Hayashi M, Fukunaga T, Kurata K, Oh-Hora M, Feng JQ, Bonewald LF,**
793 **Kodama T, Wutz A, Wagner EF, Penninger JM, Takayanagi H** (2011) Evidence for osteocyte
794 regulation of bone homeostasis through RANKL expression. *Nat Med* **17**, 1231-4.
- 795 **Newman H, Poole D** (1974) Observations with scanning and transmission electron microscopy on the
796 structure of human surface enamel. *Arch Oral Biol* **19**, 1135-1143.
- 797
798 **Newman H, Poole D** (1993) *Dental enamel growth*. *J Roy Soc Med* **86**, 61.
- 799
800 **Noble BS** (2008) The osteocyte lineage. *Arch Biochem Biophys* **473**, 106–111.
- 801
802 **Palumbo C, Palazzini S, Marotti G** (1990) Morphological study of inter-cellular junctions during
803 osteocyte differentiation. *Bone* **11**, 401–406.
- 804
805 **Parfitt AM** (1983) Steriologic basis of bone histomorphometry; theory of quantitative microscopy
806 and reconstruction of the third dimension. In: *Bone Histomorphometry: Techniques and Interpretation*
807 (Recker RR, ed). pp, 53–87. Boca Raton, FL: CRC Press.
- 808
809 **Phenice TW** (1969) A newly developed visual method for sexing the os pubis. *Am J Phys Anthropol*
810 **30**, 297-302.
- 811
812 **Ralph MR, Foster RG, Davis FC, Menaker M** (1990) Transplanted suprachiasmatic nucleus
813 determines circadian period. *Science* **247**, 975–978.
- 814
815 **Reid DJ, Beynon AD, Ramirez Rozzi FV** (1998) Histological reconstruction of dental development
816 in four individuals from a medieval site in Picardie, France. *J Hum Evol* **35**, 463–477.
- 817
818 **Reid DJ, Dean MC** (2006) Variation in modern human enamel formation times. *J Hum Evol* **50**,
819 329–346.
- 820
821 **Reid DJ, Ferrell R** (2006) The relationship between number of striae of Retzius and their periodicity
822 in imbricational enamel formation. *J Hum Evol* **50**, 195–202.
- 823
824 **Reinberg A, Sidi E, Ghata J** (1965) Circadian reactivity rhythms of human skin to histamine or
825 allergen and the adrenal cycle. *J Allergy* **36**, 273-283.
- 826
827 **Retzius A** (1837) Bemerkungenq ber den inneren Bau der Zahne, mit besonderer Rücksicht auf dem
828 in Zahnknochen Vorkommenden Röhrenbau. *Arch Anat Physiol* 486–566.
- 829 **Richter CP** (1965) *Biological Clocks in Medicine and Psychiatry*. Springfield, IL: CC Thomas.
- 830
831 **Risnes S** (1986) Enamel apposition rate and the prism periodicity in human teeth. *Scand J Dent Res*
832 **94**, 394–404.
- 833
834 **Risnes S** (1990) Structural characteristics of staircase-type retzius lines in human dental enamel
835 analyzed by scanning electron microscopy. *Anatom Rec* **226**, 135–146.
- 836
837 **Risnes S** (1998) Growth tracks in dental enamel. *J Hum Evol* **35**, 331-350.
- 838
839 **Robling AG, Duijvelaar KM, Geevers JV, Ohashi N, Turner CH** (2001) Modulation of
840 appositional and longitudinal bone growth in the rat ulna by applied static and dynamic force. *Bone*,
841 **29**, 105–113.

- 842 **Robling AG, Castillo AB, Turner CH** (2006) Biomechanical and molecular regulation of bone
843 remodeling. *Annu Rev Biomed Eng* **8**, 455-498.
844
- 845 **Robling AG, Turner CH** (2002). Mechanotransduction in bone: genetic effects on
846 mechanosensitivity in mice. *Bone* **31**(5), 562-569.
847
- 848 **Roche AF** (1992) Growth, maturation and body composition. The Fels longitudinal study 1929-1991.
849 Cambridge University Press.
850
- 851 **Schour I, Poncher HG** (1937) Rate of apposition of enamel and dentin, measured by the effect of
852 acute fluorosis. *Am J Dis Child* **54**, 757-776.
853
- 854 **Schwartz GT, Mahoney P, Godfrey LR, Cuzzo FP, Jungers WL, Randria GFN** (2005) Dental
855 development in *Megaladapis edwardsi* (primates, lemuriformes): implications for understanding life
856 history variation in subfossil lemurs. *J Hum Evol* **49**, 702-721.
857
- 858 **Schwartz GT, Reid DJ, Dean C** (2001) Developmental aspects of sexual dimorphism in hominoid
859 canines. *Int J Primatol* **22**, 837-860.
860
- 861 **Schwartz GT, Samonds KE, Godfrey LR, Jungers WL, Simons EL** (2002) Dental microstructure
862 and life history in subfossil Malagasy lemurs. *Proc Natl Acad Sci USA* **99**, 6124-6129.
863
- 864 **Schwartz JH** (1995) *Skeleton Keys*. Oxford University Press.
865
- 866 **Smith TM** (2008) Incremental dental development: methods and applications in hominoid
867 evolutionary studies. *J Hum Evol*, **54**, 205-224.
868
- 869 **Smith TM, Dean MC, Kelley J, Martin LB, Reid DJ, Schwartz GT** (2003) Molar crown formation
870 in Miocene hominoids: a preliminary synthesis. *Am J Phys Anthropol.* **36** (Suppl.), 196.
871
- 872 **Somner W** (1703) *The antiquities of Canterbury*. EP Publishing Limited.
873
- 874 **Sothorn RB** (1974) Chronobiologic serial section on 8876 oral temperatures collected during 4.5
875 years by presumably healthy man (age 20.5 at start of study). In *Chronobiology* (eds Scheving LE,
876 Halberg F, Pauly JE), pp. 245-248. Tokyo: Igaku Shoin Ltd.
877
- 878 **Sujino M, Masumoto K, Yamaguchi S, van der Horst GT, Okamura H, Inouye SI** (2003)
879 Suprachiasmatic nucleus grafts restore circadian behavioral rhythms of genetically arrhythmic mice.
880 *Curr Biol* **13**(8), 664-668.
881
- 882 **Tanner JM** (1990) *Fetus into man: physical growth from conception to maturity*. Cambridge, MA:
883 Harvard University Press.
884
- 885 **Tatsumi S, Ishii K, Amizuka N, Li M, Kobayashi T, Kohno K, Ito M, Takeshita S, Ikeda K**
886 (2007) Targeted ablation of osteocytes induces osteoporosis with defective mechanotransduction. *Cell*
887 *Metab* **5**, 464-75.
888
- 889 **Teti A, Zallone A** (2009) Do osteocytes contribute to bone mineral homeostasis? Osteocytic
890 osteolysis revisited. *Bone* **44**, 11-16.
891
- 892 **Trotter M** (1970) Estimation of stature from intact long bones. In *Personal Identification in Mass*
893 *Disasters* (eds Stewart TD), pp. 71-83. Washington, DC: Smithsonian Institution Press.
894
- 895 **Verborgt O, Gibson GJ, Schaffler MB** (2000) Loss of osteocyte integrity in association with
896 microdamage and bone remodeling after fatigue in vivo. *J Bone Miner Res* **15**, 60-7.
897

898 **van Hove RP, Nolte PA, Vatsa A, Semeins CM, Salmon PL, Smit TH, Klein-Nulend J** (2009)
899 Osteocyte morphology in human tibiae of different bone pathologies with different bone mineral
900 density—is there a role for mechanosensing? *Bone* **45**(2), 321-329.

901

902 **Weaver D** (1998) The Suprachiasmatic Nucleus: A 25-Year Retrospective. *J Biol Rhythms* **13**(2):
903 100-112.

904

905 **White T D, Black MT, Folkens PA** (2011) *Human Osteology*. 3rd ed. Boston: Academic Press.

906

907 **Wolff J** (1982) *Das Gesetz der Transformation der Knochen*. Berlin: Hirschwald.

908

909 **Zheng L, Seon YJ, Mourão MA. et al.** (2013). Circadian rhythms regulate amelogenesis. *Bone* **55**,
910 158–165.

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Table 1 Linear regression analyses of log-Retzius periodicity against log-enamel growth.

ENAMEL	<i>n</i>	Intercept	Slope	<i>r</i>	<i>r</i> ²	<i>p</i>	Residual
<i>Thickness. RP v EA</i>							
All	40	0.755	0.569	0.697	0.486	<0.001*	55%
M1	25	0.826	0.489	0.615	0.378	0.001*	64%
M2	15	0.639	0.703	0.806	0.650	<0.001*	43%
<i>Thickness. RP v AET</i>							
All	40	-0.426	0.432	0.604	0.365	0.002*	63%
M1	25	-0.391	0.384	0.577	0.333	0.004*	68%
M2	15	-0.542	0.580	0.720	0.519	0.002*	44%
<i>Rate. RP v DSR</i>							
M1	15	0.817	-0.098	0.009	0.000	0.714	98%

949 Tooth types: M1, permanent first molar; M2, permanent second molar. *Significant. EA:
 950 Enamel area. AET: Average enamel thickness. DSR: Daily secretion rate. RP: Retzius
 951 periodicity. **RP v EA:** Lower M1 (*n*=13): $r^2 = 0.491$, $p = 0.007^*$. Upper M1 (*n*=12): $r^2 = 0.633$,
 952 $p = 0.001^*$. Lower M2 (*n*=12): $r^2 = 0.603$, $p = 0.002^*$. **RP v AET:** Lower M1 (*n*=13): $r^2 =$
 953 0.338 , $p = 0.037^*$. Upper M1 (*n*=12): $r^2 = 0.482$, $p = 0.012^*$. Lower M2 (*n*=12): $r^2 = 0.287$, $p =$
 954 0.072 . Upper M2 excluded from separate analysis as $n = 3$.

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Table 2 Linear regression analyses of log-Retzius periodicity against log-bone growth.

BONE	<i>n</i>	Intercept	Slope	<i>r</i>	<i>r</i>²	<i>p</i>	Residual
<i>Stature. RP v S^a</i>							
Younger M	27	2.309	-0.082	-0.417	0.213	0.015*	74%
<i>Rate. RP v Ot.Dn</i>							
Younger M	10	3.232	-0.401	-0.370	0.159	0.326	90%
<i>Rate. S v Ot.Dn^b</i>							
Younger M	28	2.171	0.019	0.199	0.039	0.317	94%
Older M	94	2.185	0.016	0.175	0.030	0.089	93%

971 S: Estimated stature. RP: Retzius periodicity. M: Males. Ot.Dn: Osteocyte lacunae density.

972 ^aFemoral length v RP: Young M intercept= 3.232, slope= -0.135, $r=-0.492$, $p=0.020^*$.

973 ^bOt.Dn v femoral length: Young M intercept= 1.567, slope= 0.030, $r=0.199$, $p=0.317$. Older
 974 M intercept= 1.587, slope= 0.025, $r=0.176$, $p=0.088$.

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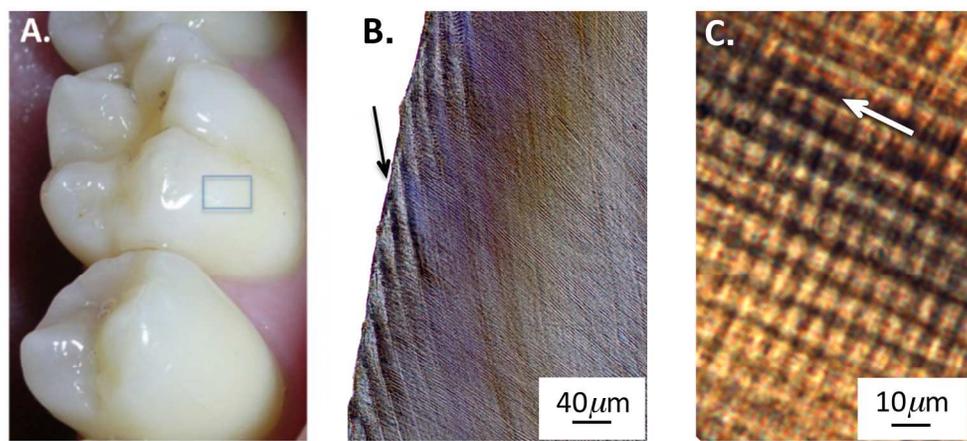


Fig 1. Human first molar with lateral enamel highlighted (A), black arrow points to infradian Retzius line (B), white arrow points in direction of prisms, with circadian cross striations at right angle to the arrow (C).

179x83mm (300 x 300 DPI)

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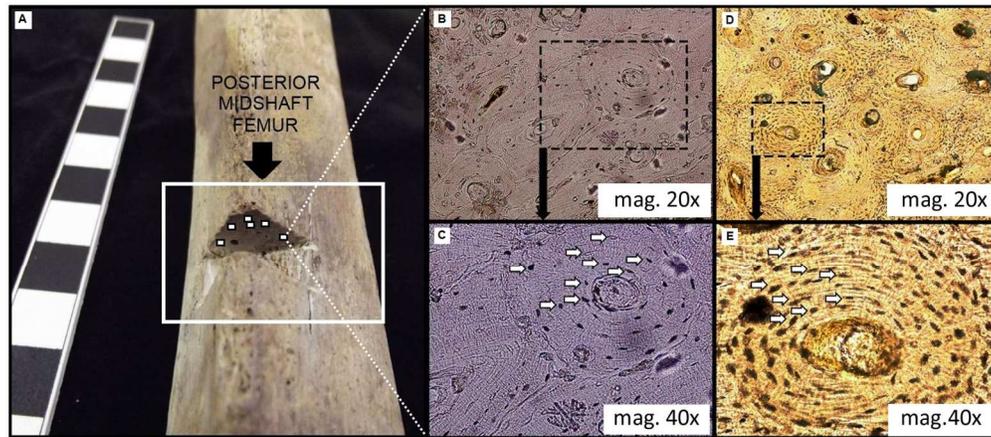


Fig 2. The figure illustrates the regions of interest (ROI) in the posterior mid-shaft femur (A). The sectioned cortical location is highlighted, with approximate (not to scale) positioning of ROIs adjacent to the periosteum (these are placed on the superior region in the sectioned area only for illustrative purposes in this figure, as we examined the removed cortical bone). The series of four histology images on the right show lower (B, and magnified in C) and higher densities (D, and magnified in E) of osteocyte lacunae (white arrows).

149x65mm (300 x 300 DPI)

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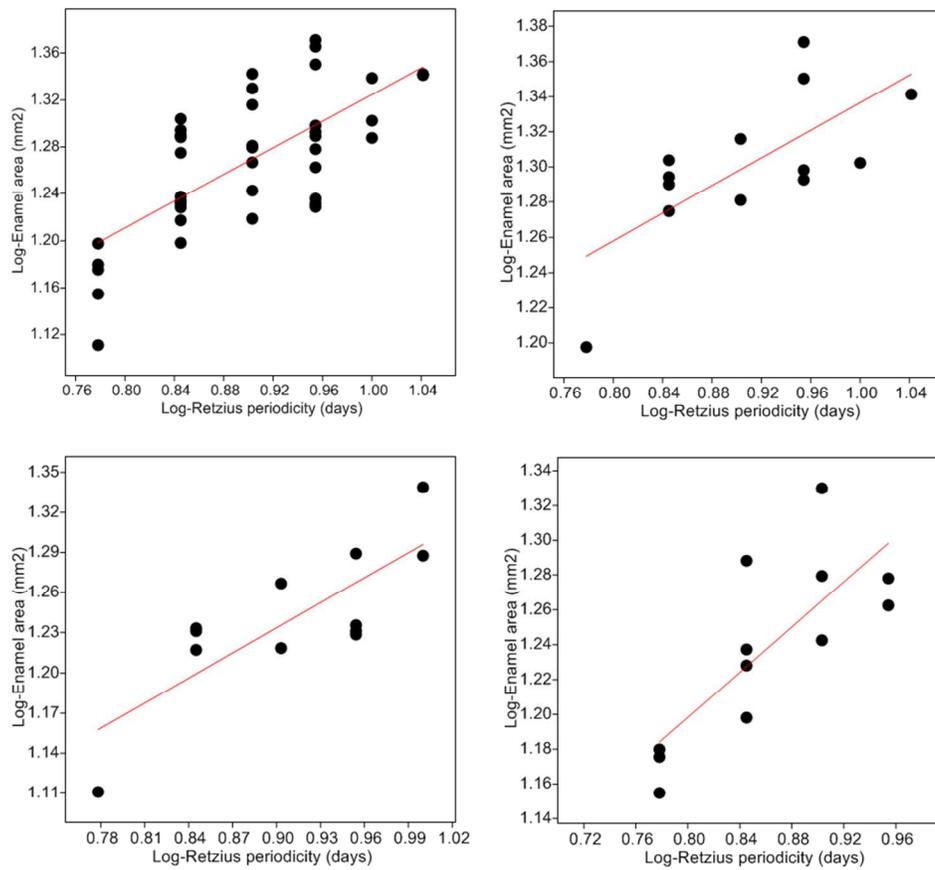


Fig. 4 Plot of log-Retzius periodicity against log-enamel area with regression lines fitted to the data. All molar types combined $n=40$ (A), which are separated into tooth types for lower first molars (B), upper first molars (C), and lower second molars (D). Upper second molars ($n=3$) are excluded from separate analysis because of the small sample size. Regression statistics are in Table 1 and footnotes. Corresponding data sets are available in Supporting Information.

83x75mm (300 x 300 DPI)



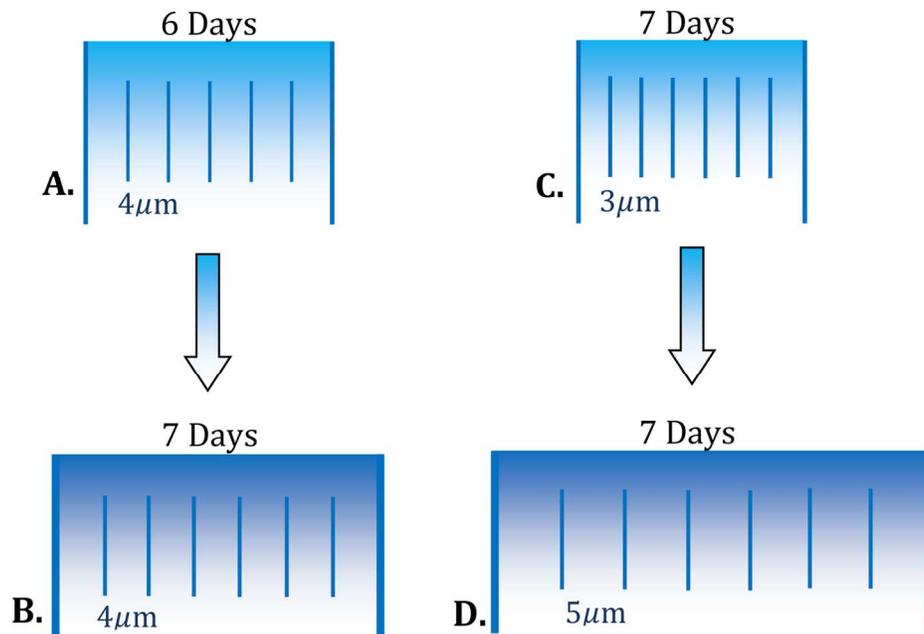


Fig 5. Cell mechanisms underlying different thicknesses of human enamel layers. Long lines illustrate two adjacent Retzius lines (enamel layer) in one outer lateral region of the crown. Short lines represent daily cross striations with either 3, 4 or $5\mu\text{m}$ of enamel between adjacent striations, representing different daily enamel secretion rates (DSR). Retzius periodicity increases from six days in A to seven days in B. Layer B increases in width because ameloblasts have secreted enamel for an extra day, relative to A, but the DSR remains constant (e.g., this study, and Mahoney et al., 2017). Another developmental mechanism is illustrated by C and D. Retzius periodicity remains the same in both illustrations. Layer D increases in width because of the greater DSR, relative to C, which might be expected when deciduous incisors are compared to second molars along the tooth row of the same individual (Mahoney, 2015).

134x93mm (300 x 300 DPI)

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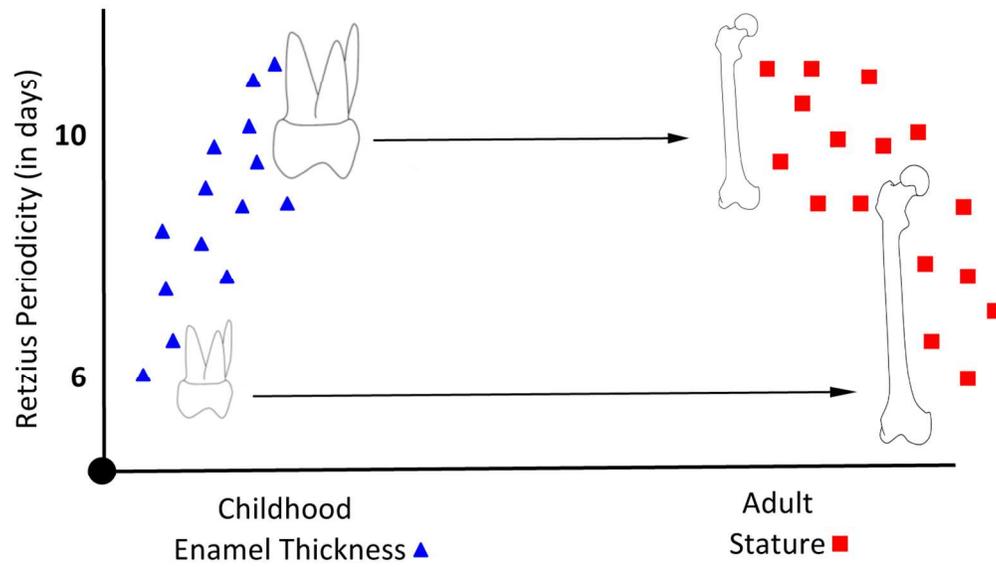


Fig 6. Hypothetical relationship between Retzius line periodicity, and the final enamel thickness of a tooth crown, and final attained adult stature.

118x70mm (300 x 300 DPI)

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1 **SUPPORTING INFORMATION**

2

3 **The Biorhythm of Human Skeletal Growth**

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33 **Table S1** Retzius periodicity (RP), enamel area, and average enamel thickness
 34 measurements for juveniles. Abbreviations: Low M1, lower first molar; Low M2, lower
 35 second molar; Up M1, upper first molar; Up M2, upper second molar. **One tooth represents**
 36 **one individual.**

Project n	Tooth type	Age category	RP (days)	Enamel area (mm ²)	AET (mm)
1	Low-M1	Juvenile	6	15.76	0.82773109
2	Low-M1	Juvenile	11	21.95	1.06864654
3	Low-M1	Juvenile	9	23.5	1.12601821
4	Low-M1	Juvenile	9	19.62	0.91596639
5	Low-M1	Juvenile	7	18.83	0.92576205
6	Low-M1	Juvenile	8	19.1	0.93996063
7	Low-M1	Juvenile	10	20.06	0.92145154
8	Low-M1	Juvenile	7	19.5	0.85152838
9	Low-M1	Juvenile	8	20.7	0.9059081
10	Low-M1	Juvenile	9	22.41	1.06107955
11	Low-M1	Juvenile	7	20.13	0.94109397
12	Low-M1	Juvenile	9	19.87	0.88626227
13	Low-M1	Juvenile	7	19.69	0.97138629
14	Low-M2	Juvenile	8	21.38	1.00328484
15	Low-M2	Juvenile	6	15.13	0.89579633
16	Low-M2	Juvenile	9	18.97	1.09022989
17	Low-M2	Juvenile	7	17.27	0.92254274
18	Low-M2	Juvenile	7	15.78	0.82230328
19	Low-M2	Juvenile	7	19.42	0.73436254
20	Low-M2	Juvenile	8	17.48	0.85351563
21	Low-M2	Juvenile	7	16.91	0.93322296
22	Low-M2	Juvenile	9	18.3	0.94524793
23	Low-M2	Juvenile	8	19.03	0.9228904
24	Low-M2	Juvenile	6	14.98	0.93918495
25	Low-M2	Juvenile	6	14.28	0.72634791
26	Up-M1	Juvenile	6	12.91	0.69428925
27	Up-M1	Juvenile	10	19.39	0.92157795
28	Up-M1	Juvenile	9	16.93	0.85591507
29	Up-M1	Juvenile	7	16.49	0.81917536
30	Up-M1	Juvenile	8	16.54	0.78537512
31	Up-M1	Juvenile	10	21.82	1.12532233
32	Up-M1	Juvenile	9	17.2	0.90669478
33	Up-M1	Juvenile	7	17.03	0.86887755
34	Up-M1	Juvenile	9	19.46	0.8413316
35	Up-M1	Juvenile	8	18.48	0.95454545
36	Up-M1	Juvenile	9	17.03	0.86490604
37	Up-M1	Juvenile	7	17.11	0.86632911
38	Up-M2	Juvenile	8	22	0.98434004
39	Up-M2	Juvenile	9	23.2	1.16407426
40	Up-M2	Juvenile	11	21.99	1.16592386

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39 **Table S2** Osteocyte density, and estimated stature (from femoral length) for younger and
 40 older adult males.

Project n	Sex	Age Category (y: 18-34yrs, m: 35-50yrs)	Side	Ot.Dn	Estimated Stature (cm)
41	M	Younger adult	left	607.69	177.08
42	M	Younger adult	right	394.87	163.99
43	M	Younger adult	right	415.38	170.89
44	M	Younger adult	right	426.92	167.56
45	M	Younger adult	right	532.05	161.61
46	M	Younger adult	right	540.38	178.98
47	M	Younger adult	right	543.59	163.51
48	M	Younger adult	right	590.38	167.32
49	M	Younger adult	right	640.00	169.94
50	M	Younger adult	right	656.41	168.75
51	M	Younger adult	right	660.00	166.13
52	M	Younger adult	right	671.16	168.75
53	M	Younger adult	left	684.62	172.32
54	M	Younger adult	right	715.38	169.70
55	M	Younger adult	left	761.54	161.61
56	M	Younger adult	right	769.23	160.89
57	M	Younger adult	right	776.92	166.13
58	M	Younger adult	right	792.31	168.99
59	M	Younger adult	right	820.51	162.56
60	M	Younger adult	right	844.62	165.89
61	M	Younger adult	right	858.97	170.89
62	M	Younger adult	right	897.44	166.84
63	M	Younger adult	right	1055.39	166.37
64	M	Younger adult	right	1062.82	166.61
65	M	Younger adult	left	1098.72	173.03
66	M	Younger adult	right	1144.23	179.22
67	M	Younger adult	right	1307.69	175.17
68	M	middle-aged	right	346.15	177.79
69	M	middle-aged	right	392.31	165.65
70	M	middle-aged	right	465.65	164.70
71	M	middle-aged	right	474.36	172.08
72	M	middle-aged	right	480.77	172.79
73	M	middle-aged	right	692.31	169.94
74	M	middle-aged	right	1255.13	170.18
75	M	middle-aged	right	305.77	169.46
76	M	middle-aged	right	365.38	179.46
77	M	middle-aged	right	376.92	178.27
78	M	middle-aged	right	383.08	169.94
79	M	middle-aged	right	403.85	176.36
80	M	middle-aged	right	421.16	171.84

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44 **Table S2** Osteocyte density, and estimated stature for younger and older adult males.

Project n	Sex	Age Category (y: 18-34yrs, m:35-50yrs)	Side	Ot.Dn	Estimated Stature (cm)
81	M	middle-aged	right	441.03	177.08
82	M	middle-aged	right	442.31	167.56
83	M	middle-aged	right	461.54	165.42
84	M	middle-aged	right	480.77	169.94
85	M	middle-aged	right	483.33	165.89
86	M	middle-aged	right	494.23	160.42
87	M	middle-aged	right	503.85	168.03
88	M	middle-aged	right	511.54	162.08
89	M	middle-aged	right	512.82	168.03
90	M	middle-aged	right	516.67	173.27
91	M	middle-aged	right	517.31	164.70
92	M	middle-aged	right	520.00	174.94
93	M	middle-aged	right	525.64	170.89
94	M	middle-aged	right	534.62	166.37
95	M	middle-aged	right	538.46	168.27
96	M	middle-aged	right	538.47	165.42
97	M	middle-aged	right	546.16	166.84
98	M	middle-aged	right	548.08	173.27
99	M	middle-aged	right	551.28	166.13
100	M	middle-aged	right	557.69	163.75
101	M	middle-aged	right	563.08	168.51
102	M	middle-aged	right	563.46	170.89
103	M	middle-aged	right	565.38	163.75
104	M	middle-aged	right	569.23	161.13
105	M	middle-aged	right	578.46	166.84
106	M	middle-aged	left	578.21	164.94
107	M	middle-aged	right	579.49	172.32
108	M	middle-aged	right	582.05	166.13
109	M	middle-aged	right	597.44	161.37
110	M	middle-aged	right	601.92	167.56
111	M	middle-aged	right	611.54	162.80
112	M	middle-aged	left	615.38	169.70
113	M	middle-aged	right	624.17	166.13
114	M	middle-aged	right	634.62	176.36
115	M	middle-aged	right	635.90	172.79
116	M	middle-aged	right	638.46	165.65
117	M	middle-aged	left	650.00	173.51
118	M	middle-aged	right	651.28	169.94

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48 **Table S2** Osteocyte density, and estimated stature for younger and older adult males.

Project n	Sex	Age Category <i>(y:18-34yrs, m:35-50yrs)</i>	SIDE	Ot.Dn	Estimated Stature (cm)
119	M	middle-aged	right	659.62	178.98
120	M	middle-aged	left	667.31	167.56
121	M	middle-aged	right	669.23	178.27
122	M	middle-aged	right	669.23	163.51
123	M	middle-aged	right	687.18	172.32
124	M	middle-aged	right	689.75	167.56
125	M	middle-aged	right	692.31	169.94
126	M	middle-aged	right	711.54	168.51
127	M	middle-aged	right	714.10	165.89
128	M	middle-aged	right	715.38	167.56
129	M	middle-aged	right	730.77	168.51
130	M	middle-aged	right	730.77	170.89
131	M	middle-aged	right	746.15	168.75
132	M	middle-aged	right	776.92	167.56
133	M	middle-aged	right	782.05	167.08
134	M	middle-aged	right	792.31	177.08
135	M	middle-aged	right	794.87	167.56
136	M	middle-aged	right	798.46	177.08
137	M	middle-aged	left	805.77	175.65
138	M	middle-aged	left	812.82	181.12
139	M	middle-aged	right	813.46	161.61
140	M	middle-aged	right	817.95	178.27
141	M	middle-aged	right	834.62	165.42
142	M	middle-aged	left	841.02	174.94
143	M	middle-aged	right	841.54	169.94
144	M	middle-aged	right	844.62	169.22
145	M	middle-aged	left	846.15	170.89
146	M	middle-aged	right	851.28	170.65
147	M	middle-aged	right	867.69	175.17
148	M	middle-aged	right	869.23	170.89
149	M	middle-aged	right	917.95	167.32
150	M	middle-aged	right	923.08	170.41
151	M	middle-aged	right	924.36	175.65
152	M	middle-aged	left	925.64	173.51
153	M	middle-aged	left	998.08	175.41
154	M	middle-aged	right	1000.00	174.70
155	M	middle-aged	left	1000.00	172.32
156	M	middle-aged	left	1030.77	175.65

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52 **Table S2** Osteocyte density, and estimated stature for younger and older adult males.

Project n	Sex	Age Category (y: 18-34yrs, m: 35-50yrs)	Side	Ot.Dn	Estimated Stature (cm)
157	M	middle-aged	right	1051.28	174.46
158	M	middle-aged	right	1165.38	168.51
159	M	middle-aged	right	1205.13	178.03
160	M	middle-aged	right	1230.77	167.80
161	M	middle-aged	right	1230.77	176.84

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55 **Table S3** Retzius periodicity (RP), and estimated stature in young adult males. Abbreviations:
 56 Low M1, lower first molar; Low M2, lower second molar; Up M1, upper first molar; Up M2,
 57 upper second molar.

Project n	Tooth type	Age Category (y: 18-34yrs)	RP	Estimated stature (cm)
272	Up-M1	Younger adult male	7	175.03
273	Low-M2	Younger adult male	8	172.32
274	Low-M2	Younger adult male	7	165.65
275	Low-M1	Younger adult male	9	176.02
276	Low-M2	Younger adult male	9	176.3
277	Low-M2	Younger adult male	8	166.33
278	Low-M2	Younger adult male	8	166.5
279	Low-M2	Younger adult male	7	179.81
280	Low-M2	Younger adult male	6	169.87
281	Low-M2	Younger adult male	9	161.97
282	Low-M2	Younger adult male	10	160.74
283	Low-M2	Younger adult male	6	182.91
284	Up-M1	Younger adult male	7	175.64
285	Up-M1	Younger adult male	8	172.14
286	Low-M1	Younger adult male	8	169.57
287	Low-M2	Younger adult male	6	180.62
288	LowM2	Younger adult male	10	173.25
289	Low-M1	Younger adult male	12	167.19
290	Low-M2	Younger adult male	8	171.35
291	Up-M1	Younger adult male	10	169.04
292	Low-M1	Younger adult male	8	175.8
293	Up-M1	Younger adult male	11	169.5
294	Low-M2	Younger adult male	6	171.25
295	Low-M1	Younger adult male	9	175.17
296	Up-M2	Younger adult male	9	166.00
297	Up-M2	Younger adult male	7	169.66
298	Low-M2	Younger adult male	7	182.91

For Peer Review Only