

Kent Academic Repository

Full text document (pdf)

Citation for published version

Loo, Ruey Leng (2018) Characterization of metabolic responses to healthy diets and the association with blood pressure: application to the Optimal Macronutrient Intake Trial for Heart Health (OmniHeart), a Randomized Control Study. Characterization of metabolic responses to healthy diets and the association with blood pressure: application to the Optimal Macronutrient Intake Trial for Heart

DOI

Link to record in KAR

<http://kar.kent.ac.uk/65733/>

Document Version

Author's Accepted Manuscript

Copyright & reuse

Content in the Kent Academic Repository is made available for research purposes. Unless otherwise stated all content is protected by copyright and in the absence of an open licence (eg Creative Commons), permissions for further reuse of content should be sought from the publisher, author or other copyright holder.

Versions of research

The version in the Kent Academic Repository may differ from the final published version.

Users are advised to check <http://kar.kent.ac.uk> for the status of the paper. **Users should always cite the published version of record.**

Enquiries

For any further enquiries regarding the licence status of this document, please contact:

researchsupport@kent.ac.uk

If you believe this document infringes copyright then please contact the KAR admin team with the take-down information provided at <http://kar.kent.ac.uk/contact.html>

Characterization of metabolic responses to healthy diets and the association with blood pressure: application to the Optimal Macronutrient Intake Trial for Heart Health (OmniHeart), a Randomized Control Study

Author names:

Ruey Leng Loo*, Xin Zou, Lawrence J Appel, Jeremy K Nicholson and Elaine Holmes

Affiliations:

Medway Metabonomics Research Group, Medway School of Pharmacy, Universities of Kent and Greenwich, Chatham Maritime, Kent, UK (RLL, XZ).

Ministry of Education Key Laboratory of Systems Biomedicine, Shanghai Centre for Systems Biomedicine, Shanghai Jiao Tong University, Shanghai, 200240, China (XZ)

Department of Medicine, Johns Hopkins University School of Medicine, Baltimore, Maryland (LJA).

Welch Center for Prevention, Epidemiology, and Clinical Research, Johns Hopkins University, Baltimore, Maryland (LJA).

Division of Computational and Systems Medicine, Department of Surgery and Cancer, Imperial College London, UK (JKN and EH).

MRC-HPA Centre for Environment and Health, Imperial College London, UK (JKN and EH).

Authors' last name: Loo, Zou, Appel, Nicholson and Holmes

Disclaimers: None

Address corresponding to RL Loo, Medway School of Pharmacy, Anson Building, Chatham Maritime, Kent, ME4 4TB. Telephone: +44 (0)1634 202948. Email: r.loo@kent.ac.uk

Source of funding: This work was supported by Medical Research Council, New Investigator Grant Award (G1002151). X Zou is also supported by Natural Science Foundation of Shanghai (16ZR1417900) and Shanghai Pujiang Talent Fund (16PJ1405200). E Holmes and JK Nicholson also acknowledge the support from Biotechnology and Biological Sciences Research Council (PS8813) and National Institute for Health Research (PSA809).

Short running title: Metabolic response to diets and the link with BP

Abbreviation: 1D, one-dimensional; CVDs, cardiovascular diseases; BP, blood pressure; CI, confidence interval; DASH, Dietary Approaches to Stop Hypertension; HDR, homogenous dietary response; HDL, high-density lipoprotein cholesterol; LDL, low-density lipoprotein cholesterol; mOPLS-DA, multilevel orthogonal partial least squares discriminant analysis; NAD, nicotinamide adenine dinucleotide; NMR, nuclear magnetic resonance; OmniHeart, Optimal Macronutrient Intake Trial for Heart Health; OmniCarb, OmniHeart carbohydrate rich diet; OmniMFA, OmniHeart monounsaturated fat rich diet; OmniProt, OmniHeart protein rich diet; SD, standard deviation; SHOCSY, statistical homogeneous cluster spectroscopy; TSP, sodium 3-trimethylsilyl-(2,2,3,3-²H₄)-1-propionate; VDR, variable dietary response.

Clinical Trial Registration: The original OmniHeart intervention study is registered at www.clinicaltrials.gov as NCT00051350 and metabolomics study is registered at www.clinicaltrials.gov as

1 ABSTRACT

2 **Background:** Inter-individual variation in the response to diet is common but the underlying
3 mechanism for such variation is unclear.

4 **Objective:** The objective of this study was to use a metabolic profiling approach to identify
5 a panel of urinary metabolites representing individuals demonstrating typical
6 (homogeneous) metabolic responses to healthy diets, and subsequently to define the
7 association of these metabolites with improvement of risk factors for cardiovascular
8 diseases (CVD).

9 **Design:** 24-h urine samples from 158 participants, with pre-hypertension and stage 1
10 hypertension collected at baseline and following the consumption of a carbohydrate-rich, a
11 protein-rich and a monounsaturated fat-rich healthy diet (6-weeks per diet) in a randomized,
12 crossover study, were analyzed by proton (^1H) nuclear magnetic resonance (NMR)
13 spectroscopy. Urinary metabolite profiles were interrogated to identify typical and variable
14 responses to each diet. We quantified the differences in absolute excretion of metabolites
15 distinguishing between dietary comparisons within the typical response groups and
16 established their associations with CVD risk factors using linear regression.

17 **Results:** Globally all three diets induced a similar pattern of change in the urinary metabolic
18 profiles for the majority of participants (60.1%). Diet-dependent metabolic variation was not
19 significantly associated with total cholesterol or low density lipoprotein cholesterol levels.
20 However, blood pressure (BP) was found to be significantly associated with six urinary
21 metabolites reflecting: dietary intake (proline-betaine [inverse], carnitine [direct]); gut
22 microbial co-metabolites (hippurate [direct], 4-cresyl sulfate [inverse], phenylacetylglutamine
23 [inverse]), and tryptophan metabolism (N-methyl-2-pyridone-5-carboxamide [inverse]). A
24 dampened clinical response was observed in some individuals with variable metabolic

25 responses, which could be attributed to non-adherence to diet (up to 25.3%), variation in gut
26 microbiome activity (7.6%) or a combination of both (7.0%).

27 **Conclusion:** These data indicate inter-individual variations in BP in response to dietary
28 change and highlight the potential influence of the gut microbiome in mediating this
29 relationship. This approach provides a framework for stratification of individuals undergoing
30 dietary management.

31

32 **Keywords:** diets; gut microbiome; hypertension; metabolic profiling, metabonomic,
33 metabolomic; and personalized health care.

34

35 INTRODUCTION

36 Of the total global deaths, approximately half are attributed to cardiovascular
37 diseases (CVDs), with elevated BP being a key risk factor (1). Genome-wide association
38 studies have identified common genetic variants associated with high BP (2) but these only
39 account for a small proportion of the population variance in BP and do not take lifestyle
40 factors such as physical inactivity or unhealthy diet into account. CVD remains the leading
41 cause of mortality for non-communicable diseases worldwide, even though the adoption of
42 healthy dietary patterns such as those promoted by Dietary Approaches to Stop
43 Hypertension (DASH) (3), Optimal Macronutrient Intake Trial for Heart Health
44 (OmniHeart) (4) and Mediterranean diets (5) have unequivocally been shown to reduce
45 CVD risk. Humans demonstrate substantial variation in response to dietary intervention,
46 partially attributable to genetic heterogeneity (6, 7). For example, the apolipoprotein A-IV
47 protein modulates cholesterol lowering responses to high fat diets (8, 9). However,
48 supporting evidence for genetic influence on variable dietary responses remains conflicting
49 (10) and modifiable factors such as changes in body weight (11, 12), or variation in the
50 composition of the gut-microbiome (13) and virome (14), have been implicated in variation
51 in dietary responses.

52 Metabolic phenotyping technologies provide a framework for investigating the
53 influences of environmental and lifestyle factors on disease risk and have been successfully
54 applied to investigate chronic diseases e.g. diabetes (15). Systematic modulation of
55 metabolism in response to food intake (16) has been reported and the impact of diet in a
56 range of pathological conditions, including gastrointestinal cancer risk, has been assessed
57 (17). Building on methodological approaches developed for characterizing inter-individual
58 variation in response to drug toxicity/therapies (18), we propose to demonstrate the
59 feasibility of identifying inter-individual variation in clinical response to three different

60 healthy diets, using a ^1H NMR based metabolic phenotyping approach and establish the
61 impact of this variation on CVD risk,. We hypothesized that dietary change from a typical
62 American diet to a healthy diet or between different healthy diets would result in typical
63 changes in the urinary metabolic phenotypes for the majority of individuals, herein
64 considered as homogenous dietary response (HDR) group. We ascertained that a minority
65 of individuals demonstrated atypical dietary responses, herein referred to as variable
66 (heterogenous/non-uniform) dietary responders (VDR). We further hypothesized that these
67 specific urinary dietary response phenotypes would be associated with BP. Variation in
68 diet-specific biomarkers will further enhance our understanding of the link between
69 variation in dietary response and the aetiopathogenesis of hypertension.

70

71 **METHODS**

72 **OmniHeart Study design**

73 The OmniHeart Study (N=163) was a randomized, controlled, three period cross-
74 over feeding study aiming to assess the effects of three healthy diets on BP and lipid
75 profiles (19). The key findings and the study design of OmniHeart Study have been
76 previously published (4, 19). Briefly, all three OmniHeart diets had a similar nutrient
77 composition to the established healthy DASH diet but varied in macronutrient composition.
78 The Omniheart carbohydrate-rich diet (OmniCarb diet) provided 58% kcals from
79 carbohydrate, 15% from protein and 27% from fat; the remaining two diets, replaced 10%
80 of calories from carbohydrate with either protein, predominantly obtained from vegetable
81 sources (OmniProt diet), or unsaturated fats, predominantly derived from monounsaturated
82 fat (OmniMFA diet). Participants were randomly assigned to one of six possible orders of
83 administration of the three diets, each intervention period lasting for 6-weeks. During each
84 intervention period, the participants were requested to only consume food prepared in the

85 diet kitchen and were allowed to consume up to 2 alcoholic beverages and 3 non-caloric
86 caffeinated beverages per day as part of the trial. Their main meal was consumed on-site on
87 weekdays and all other meals were eaten at home. Participants completed a diary in which
88 they indicated whether they had complied with the study food protocol during the feeding
89 periods. During the screening visits and washout periods (at least 2 weeks), participants
90 consumed their own food. The Willett food frequency questionnaire (20), administered by
91 certified staff as a means to describe the usual food intake of participants during screening
92 visits indicated participants consume a typical American diet at the outset of the study;
93 corresponding to high intake of saturated fat, excessive refined sugar and salt with low
94 intake of fruit, vegetables and omega-3-fat.

95 A total of 163 men and women, aged between 30 to 80 years from the Baltimore and
96 Boston areas, with pre-hypertension (systolic BP of 120 to 139 mmHg and/or diastolic BP of
97 80 to 89 mmHg) or stage 1 hypertension (systolic BP of 140 to 159 mm Hg and/or diastolic
98 BP of 90 to 99 mm Hg) and without diabetes or prior CVD were recruited to the study. The
99 minimum detectable, between-diet differences for primary (systolic BP) and secondary
100 (diastolic BP, low density lipoprotein cholesterol [LDL], high density lipoprotein cholesterol
101 [HDL], triglyceride and total cholesterol) variables in the full cohort (n=160) and in
102 subgroups (n=80 and 70) were at 80% and 90% power (2-sided alpha, p=0.05).
103 The sample size of the trial (n=160) was selected because it provided adequate power to
104 detect between-diet differences in the primary outcome variables that have public health
105 significance, both overall and in subgroups. Specifically, the minimum detectable effect size
106 for systolic BP was < 3 mmHg even in subgroups that comprised only 40 % (n=64) of
107 participants. One individual completed just one dietary intervention period, and four
108 individuals completed two intervention periods. The remaining 158 completed all three
109 dietary interventions, provided four 24-h urine collections and supplied anthropometric and

110 sociodemographic metrics on CVD (**Supplemental Figure 1**). These four 24-h urine
111 collections corresponded to the baseline screening visit and one at the end of each of the three
112 6-week dietary interventions. NMR urine spectra for these 158 individuals were used for the
113 analyses presented here. During the last 10 days of each dietary intervention period, a fasting
114 blood specimen was obtained to measure lipid levels. BP was measured on 5 days by trained
115 staff using the OMRON 907 device for those requiring a normal or large adult cuff, after
116 participants had been seated for at least 5 mins. The reported BP was based on the average of
117 nine BP measurements taken at screening visits and 15 measurements taken at the last five
118 visits of each feeding period. Body weight for all participants was maintained within 2% of
119 their baseline throughout the study period by adjusting caloric levels each week-day.
120 Baseline socio-demographic and anthropometric characteristics were obtained for each
121 participant. Institutional ethics committee approval was obtained for each site and all
122 participants provided written informed consent.

123

124 **NMR based metabolic phenotyping and data processing**

125 Urine specimens were analyzed by 600 MHz ^1H NMR spectroscopy using a Bruker
126 NMR spectrometer (Bruker Biospin, Rheinstetten, Germany) according to a standard protocol
127 (21) in our London metabolic phenotyping laboratory. Urine specimens were allowed to thaw
128 at room temperature and centrifuged at 12,000g for 5 mins to remove particulates. For each
129 specimen, 500 μL of urine was mixed with 250 μL of phosphate buffer solution at pH 7.4 ± 0.1 .
130 The resulting mixtures were left to stand for 10 mins and then further centrifuged as before.
131 A total volume of 500 μL of the supernatant was added to 50 μL of sodium 3-trimethylsilyl-
132 (2,2,3,3- $^2\text{H}_4$)-1-propionate (TSP) in Deuterium Oxide, giving a final concentration of 1mM.
133 This solution was transferred to a 5mm NMR tube. The prepared urine specimens were
134 placed in the auto-sampler, analyzed in a simple randomized order generated by computer. A

135 one-dimensional (1D) pulse sequence with a water saturation method (recycle delay – 90° – t₁ –
136 90° – t_m – 90° – acquisition) was used to acquire standard ¹H NMR spectra of urine. The spectra
137 were acquired with 64K data points and 128 scans over a spectral width of 12kHz. The
138 recycle delay was set to 2s with a mixing time (t_m) of 100ms and a t₁ of 20μs, providing an
139 acquisition time of approximately 2.72s. All ¹H NMR spectra were phased, baseline
140 corrected, and manually referenced to sodium 3-trimethylsilyl-(2,2,3,3-²H₄)-1-propionate
141 (TSP) at δ 0 with Topspin software (version 2.1, Bruker Biospin) prior to multiplication by an
142 exponential weighting function corresponding to a line broadening of 0.3Hz. The spectral
143 regions containing the water (δ 4.5 to 5.05) and urea (δ 5.5 to 6.5) resonances, as well as the
144 extreme ends (<δ 0.7 and > δ 9.5) of the spectra that contain minimal metabolic information,
145 were removed. Initial analysis showed that the signal arising from the –CH₂ and –CH₃ group
146 of the creatinine peaks dominated the analysis due to the high concentration of creatinine
147 compared to other metabolites. Since there was no statistical difference in the clinical
148 creatinine measurements at screening visit and at the end of each study period based on Jaffé
149 reaction measurement (p>0.5 for all comparison between each diet and the baseline), we
150 removed the creatinine regions containing the peaks at δ 3.035-3.062 and δ 4.052-4.075 from
151 all subsequent analysis. A total of 23,998 NMR data variables, at a full resolution (0.0003
152 ppm), were then normalized by a probabilistic quotient method (22) using the median
153 spectrum of the whole dataset as a reference and subsequently scaled to unit-variance.

154

155 **Data analysis**

156 We applied Statistical HOmogeneous Cluster SpectroscopY (SHOCSY) (23) to the
157 processed and normalized spectroscopic data. SHOCSY is a variant of statistical
158 spectroscopic techniques such as the Subset Optimization by Reference Matching (STORM)
159 (24) and Statistical TOtal Correlation SpectroscopY (STOCSY) (25). SHOCSY involves

160 clustering of the spectral data based on the similarity/dissimilarity of the spectral features
161 followed by the association of clusters to different dietary groups using an enrichment test.
162 The application of SHOCSY enables identification of the groups of spectra showing
163 uniform/homogeneous urinary metabolic responses (HDR) and those showing variation from
164 the coherent metabolic response (VDR) following the consumption of different OmniHeart
165 diets. Due to the nature of cross-over study design, we employed multilevel orthogonal partial
166 least square-discriminant analysis (mOPLSDA) (26, 27), which incorporates the variation
167 between and within participants in the dataset to optimize visualization of dietary response, in
168 conjunction with SHOCSY. We performed this in a pairwise fashion, comparing the urinary
169 spectral data from the screening visit (reflecting a basal dietary pattern) with those from the
170 end of each dietary intervention and modelled this separately for the urinary spectral data
171 corresponding to a HDR (3 models, 1 per diet) and those representing a VDR (3 models).
172 Thus, each subgroup was compared to its own baseline. We also performed comparison
173 between different OmniHeart diets and separately for the HDR (3 models) and VDR (3
174 models) groups creating a total of 12 different mOPLSDA models, **Supplemental Table 1**.
175 Each mOPLSDA comparison was validated using a seven-fold cross-validation procedure.
176 The model statistics, Q^2Y_{hat} (28) is defined as the proportion of variance in the data predicted
177 by the mOPLSDA model and is therefore a measure of the robustness of the model. In
178 addition, permutation testing was performed by randomly assigning classes to the samples
179 and remodeling repeatedly for 100 times. The Q^2Y_{hat} statistic for the real model was then
180 compared to the null hypothesis distribution obtained from the permuted Q^2Y_{hat} t values and
181 was considered significant when the p-value of the real Q^2Y_{hat} was <0.05 on those permuted
182 values.

183 The three criteria used to identify discriminatory metabolites were: i) P-values of the
184 correlations between the spectral variable and the mOPLS-DA scores vector should be $<$

185 1.85×10^{-6} (corresponding to $p < 0.05$ after Sidák correction); ii) a variable loading coefficient
186 strength, $r^2 > 0.3$ as defined in Zou et al (23); and iii) the stability of the NMR variables,
187 whereby a data point was considered significant when flanked by two NMR spectral variables
188 conforming to criteria i) and ii). For peaks that were free from spectral overlap, the 24-h
189 urinary excretion of each discriminatory metabolite was quantified by integration of the NMR
190 signal intensities. Since we found no significant difference in the excretion of creatinine
191 between different OmniHeart diets and the typical American diet ($P > 0.5$), the absolute
192 excretion of each discriminatory metabolite was normalized to the corresponding 24-h urinary
193 creatinine excretion (in mmol/L). The difference in absolute excretion of each discriminatory
194 metabolite was determined for the comparison of each dietary intervention with baseline or
195 between different OmniHeart dietary interventions. The association between the differences
196 in absolute excretion of each discriminatory metabolite and changes in CVD risk factors
197 (systolic and diastolic BP, LDL, total cholesterol) was established using linear regression for
198 HDR groups. In addition, known covariates for hypertension including urinary excretion of
199 sodium, potassium, calcium and phosphate, were also established for HDR and VDR groups
200 for the comparison between baseline and each OmniHeart diet. The statistical significance of
201 these covariates was adjusted by Bonferroni correction (0.05 divided by number of
202 comparisons) to account for multiple testing. All analyses were performed using in-house
203 software written in Matlab (version 2012a, MathWorks, Natick, MA).

204

205 **Identification of discriminatory metabolites**

206 The discriminatory metabolites found to be significantly influenced by the healthy dietary
207 interventions were confirmed by in-house and published database (29) references and
208 authenticated by spiking in standard compounds purchased from Sigma Aldrich. These
209 compounds included: N-methyl-2-pyridone-5-carboxamide, 4-hydroxyphenylacetic acid,

210 carnitine, creatine, dimethylglycine, S-methyl-L-cystiene-S-oxide, N-methyl nicotinic acid, N-
211 methyl nicotinamide, proline-betaine and hippurate. For the remaining urinary metabolites
212 where they were not available commercially, identification was achieved using further
213 analytical methods such as two dimensional NMR experiments, solid phase extraction
214 chromatography experiments coupled with NMR, ultra-performance liquid chromatography
215 coupled to mass spectroscopy, and statistical analysis such as Subset Optimization by Reference
216 Matching (STORM) (24) as well as using published databases and/or literature.

217

218 **RESULTS**

219 **Individuals show variation in urinary metabolic phenotypes to OmniHeart diets**

220 Participants' demographics and changes in CVD risk factors following each
221 OmniHeart diet are provided in **Table 1**. Each diet elicited a range of clinical responses
222 over the six-week study, in terms of reduction of CVD risk factors, which was reflected in
223 the urinary metabolome. Inter-individual differences in dietary response were observed;
224 the majority of the participants showed a HDR to all of the OmniHeart diets when
225 compared with the baseline profile: 71.5% (N=113) for OmniProt, 80.4% (N=127) for
226 OmniMFA and 86.7% (N=137) for OmniCarb. The remaining individuals who did not
227 demonstrate a 'typical' response to a given diet were grouped into the VDR class: N=45 for
228 OmniProt, N=31 for OmnMFA, and N=21 for OmniCarb. A similar modelling strategy was
229 applied to compare between pairs of OmniHeart diets. We found > 70% participants
230 showed consistent metabolic differences between diets, **Supplemental Table 1**.

231

232 **OmniHeart diets show distinctive urinary metabolic phenotypes**

233 Each of the three OmniHeart diets was associated with a distinct metabolic
234 phenotype in the majority of participants (the HDR group). For the OmniHeart-baseline

235 comparisons, the discriminatory metabolites were predominantly related to: i) dietary intake
236 - increased excretion of proline-betaine, N-acetyl-S-methyl-L-cysteine sulfoxide, S-methyl-
237 L-cysteine-S-oxide, creatine, and carnitine; ii) tryptophan-nicotinamide-adenine
238 dinucleotide (NAD) degradation - reduced excretion of N-methyl-2-pyridone-5-
239 carboxamide and N-methyl nicotinamide, and increased excretion of N-methyl nicotinic
240 acid; and iii) gut microbial-mammalian metabolism - increased excretion of hippurate and
241 dimethylglycine, and reduced excretion of 4-hydroxyphenylacetic acid, **Supplemental**
242 **Table 2**. Compared to the baseline profiles, proline-betaine was the only metabolite
243 uniformly increased in the urinary phenotypes of HDR groups across all three diets,
244 consistent with increased citrus fruit consumption (30). Increased excretion of carnitine
245 and creatine in the OmniProt diet reflected the increase in protein intake (31).

246 Additional pairwise comparisons ($P < 10^{-5}$) between different OmniHeart diets
247 further indicated that each diet was associated with a distinct metabolic phenotype. The
248 HDR group of the OmniProt diet was generally characterized by higher excretion of urinary
249 creatine; N- methyl-2-pyridone-5-carboxamide and two gut microbial mammalian co-
250 metabolites, phenylacetylglutamine and 4-cresyl sulfate compared to the other two
251 OmniHeart diets; whilst the HDR group for the OmniCarb diet consistently showed higher
252 excretion of hippurate and guanodinoacetate (**Supplemental Table 3 and 4**). The
253 differences in the markers for dietary intake of cruciferous vegetables (S-methyl-L-
254 cystiene-S-oxide and N-acetyl-S-methyl-L-cysteine sulfoxide) (32) and markers for citrus
255 fruit intake (proline-betaine) (30) observed when comparing urine of OmniHeart diets with
256 the baseline profiles, were generally not observed for pairwise comparisons between the
257 OmniHeart diets since all three diets included higher proportions of fruit/vegetables than
258 the baseline.

259

260 **Urinary metabolites significantly associated with BP**

261 We quantified ten discriminatory metabolites altered in response to one or more
262 OmniHeart diets and assessed their associations with BP and lipid profiles using the HDR
263 groups only. Although no significant associations were found between dietary phenotypes and
264 LDL or total cholesterol, we found significant associations between two of these food related
265 metabolites with BP. Proline-betaine was inversely associated with systolic and diastolic BP
266 for OmniCarb and OmniMFA diets when compared to baseline ($P < 0.05$, **Table 2**). A similar
267 trend was observed for the OmniProt diet although it was not statistically significant. A direct
268 association was found between systolic BP and carnitine for the OmniProt diet when
269 compared to baseline ($P < 0.05$). We found three metabolites related to host-gut microbial
270 pathways that were significantly associated with BP (hippurate, phenylacetylglutamine and 4-
271 cresyl sulfate). Hippurate showed a direct association with systolic BP ($P < 0.001$) and
272 diastolic BP ($P < 0.01$) levels for the OmniCarb diet compared to baseline, whereas 4-cresyl
273 sulfate and phenylacetylglutamine (distal colonic microbial metabolites of tyrosine and
274 phenylalanine, respectively) were inversely associated with BP for the comparison between
275 OmniMFA and OmniProt diets. N-methyl-2-pyridone-5-carboxamide (tryptophan-NAD
276 metabolite) was also found to be inversely associated with systolic and diastolic BP levels for
277 the OmniCarb-baseline comparison ($P < 0.05$). These data demonstrate healthy diets can elicit
278 coherent changes in the urinary metabolic phenotypes for the majority of individuals and that
279 some of these metabolites are either directly or inversely associated with BP.

280

281 **Urinary metabolic phenotypes can identify non-adherence to diets**

282 The urinary spectral data for the VDR groups for each of the OmniHeart diets typically
283 produced fewer dietary-specific discriminatory metabolites than the HDR groups
284 (**Supplemental Tables 2 and 3**). The VDR groups also showed discordance in the levels of

285 proline-betaine and hippurate when compared to the HDR groups. Since increased
286 consumption of citrus fruits was a feature of all dietary interventions, we therefore classified
287 individuals with a lower level of proline-betaine (a direct marker of citrus fruit intake) (33,
288 34), as non-adherent to these diets on the assumption that this was generally indicative of
289 dietary behavior. We found the majority of participants in the VDR groups excreted lower
290 24-h urinary concentrations of proline-betaine when compared to the HDR groups. Fifteen of
291 the 21 individuals (71.4%) from the OmniCarb-VDR group showed a 24-h urinary excretion
292 of less than 95% confidence interval (CI) obtained for proline-betaine excretion of the
293 OmniCarb-HDR group. A similar trend was observed for the OmniMFA-VDR (21/31,
294 67.7%) and OmniProt-VDR (35/45, 77.8%) groups. The overall estimation of non-adherence
295 to each diet was: 9.5% (n=15) for the OmniCarb, 13.3% (n=21) for the OmniMFA and 22.2%
296 (n=35) for the OmniProt diet. Despite sub-classification of VDR groups as adherent or non-
297 adherent, contrasting patterns remained in the VDR and HDR groups, as exemplified for
298 hippurate (a gut microbial co-metabolite of dietary phenols), where increased excretion of
299 hippurate was characteristic for the HDR but not either of the VDR (diet adherent or non-
300 adherent) subgroups for OmniCarb. Differential metabolite patterns were also observed for
301 different subgroups within the OmniMFA (**Figure 1**).

302

303 **Urinary metabolic variation reflects inter-individual differences in clinical responses**

304 Discarding the non-adherent VDR group, we assessed the effect of each diet, stratified
305 by the HDR versus adherent-VDR, on urinary electrolyte concentrations. We found
306 significant overall changes in mean urinary sodium (decrease) and mean urinary potassium
307 (increase) in the HDR groups for all OmniHeart diets when compared to baseline values
308 (**Supplemental Table 5**). The mean changes in urinary electrolytes were of slightly greater
309 magnitude when considering the subset of pre-hypertensive individuals within the HDR

310 groups for sodium: -31.3mmol/day (OmniCarb), -44.9mmol/day (OmniMFA), and -
311 35.9mmol/day (OmniProt); and potassium 26.4 mmol/day (OmniCarb), 28.4 mmol/day
312 (OmniMFA) and 24.7 mmol/day (OmniProt), $P<0.001$ (data not shown). This general trend
313 in mean urinary sodium and potassium levels was apparent for the adherent-VDR groups but
314 the changes from baseline level were insignificant. With regard to the inter-comparison
315 between OmniHeart diets, no systematic differences were observed in the electrolyte levels
316 with the exception of higher urinary sodium and phosphate levels being characteristic of the
317 OmniProt-HDR when compared to the OmniMFA-HDR group ($P<0.01$, data not shown). No
318 systematic differences in electrolytes were expected as micronutrients such as potassium,
319 sodium, calcium and magnesium were indexed to the energy level from the diet for each
320 participant (19).

321 We also investigated the changes in CVD risk factors post-diet and found a significant
322 ($P<10^{-10}$) reduction in all HDR diet groups when compared to the baseline for systolic and
323 diastolic BP, LDL and total cholesterol. Additionally, the reduction in serum triglyceride
324 concentrations was significant for the OmniProt-HDR group; and HDL for the OmniCarb-
325 HDR and OmniProt-HDR groups, $P<0.05$, (**Figure 2**). High risk individuals such as those
326 who were hypertensive or those with non-optimal lipid profiles in the HDR groups showed
327 greater reduction in these CVD risk factors than low risk individuals (**Supplemental Figure**
328 **2**). For all the VDR groups, a dampened reduction in CVD risk factors was generally
329 observed when compared to the corresponding HDR comparator groups (**Figure 2**). A
330 significant ($P<0.05$) reduction in systolic and diastolic BP was observed in both the adherent-
331 and non-adherent-OmniMFA-VDR and the non-adherent-OmniProt-VDR groups; whilst the
332 adherent- and non-adherent-OmniProt-VDR groups also generally showed significant
333 reductions for LDL, HDL and total cholesterol although the magnitude of the change in CVD
334 risk factors was generally more variable than that observed for the corresponding HDR

335 groups. The observed lack of dietary-induced clinical benefit in the adherent-VDR groups
336 may be partially due to the reduced sample size ($N < 10$) following stratification of the cohort.
337 In addition to the observation that HDR groups of all three OmniHeart diets generally elicited
338 a reduction in CVD risk factors when compared to typical American diets, we also found the
339 HDR-OmniProt group generally showed a larger overall reduction in the CVD risk factors
340 when compared to the HDR-OmniMFA and HDR-OmniCarb groups (**Supplemental Figure**
341 **3**).

342

343 **Stratification of individual response based on urinary metabolic phenotypes**

344 From a cohort of 158 individuals, who partook in all three dietary interventions, we
345 were able to stratify individuals according to diet-response specific urinary phenotypes;
346 corresponding to those who demonstrated: HDR to all three diets ($N=95$, 60.1%; Group 1);
347 HDR to two diets but VDR to one diet ($N=35$, 22.2%; Group 2); HDR to only one diet but
348 VDR to two diets ($N=22$, 13.9%; Group 3); non-adherent-VDR to all three diets ($N=4$, 2.5%;
349 Group 4); and mix of non-adherent- and adherent-VDR to all three diets ($N=2$, 1.3%; Group
350 5). Moreover, we were able to further sub-stratify individuals in the VDR groups that
351 demonstrated a dampened clinical response into those participants that were: a) adherent to
352 diets but showed differences in metabolic phenotypes from the majority of participants
353 (including gut-microbial co-metabolites; $N=12$, 7.6%); b) non-adherent to one or more diet
354 ($N=40$, 25.3%); or c) a combination of the two ($N=11$, 7.0%), **Table 3**. We found that
355 individuals consistently classified as HDR for all three OmniHeart diets generally manifested
356 a greater reduction in CVD risk factors than those that were classified as HDR for just one or
357 two of the OmniHeart (**Supplemental Figure 4**).

358

359 **DISCUSSION**

360 We show that the majority, but not all, of the participants responded similarly in terms
361 of their expressed metabolic phenotype to a particular diet and that each of the three diets had
362 a distinct effect on the metabolism. However, regardless of the macronutrient differences
363 between the three OmniHeart diets and the diet-specific impact on the metabolic profile, the
364 majority of participants (60.1%), demonstrated post-diet improvement in clinical risk factors
365 for CVD. We applied an agnostic multivariate statistical tool to identify participants who
366 showed a coherent biochemical response (HDR) to each of the diet and sub-divided the
367 dataset into high- and low-risk individuals based on their BP status or lipid profiles. Although
368 both groups demonstrated a coherent biochemical response irrespective of the CVD risk status
369 the high-risk groups generally demonstrated a larger reduction in CVD risk factors than low-
370 risk individuals. Our results thus demonstrate that manipulation of dietary macronutrient
371 content, without alteration of caloric intake and body weight, can elicit coherent changes in
372 metabolic profiles and contribute to beneficial effects on both BP levels and lipid profiles

373 Notably, we identified two gut microbial-host co-metabolites associated with BP:
374 phenylacetylglutamine and 4-cresyl sulfate, deriving from phenylalanine and tyrosine,
375 respectively, resulting from bacterial putrefaction of protein in the distal colon. The gut
376 microbiota, in particular Firmicutes and Bacteroidetes, can adapt to dietary changes and
377 induce changes in host metabolism (35): an increase of Firmicutes to Bacteroidetes ratio has
378 been demonstrated in spontaneous hypertensive rats (36). Other researchers have
379 manipulated gut microbiota balance via probiotic administration with consequent beneficial
380 effects on BP levels (37). More recently, blood levels of phenylacetylglutamine were found
381 to be strongly anti-correlated with BP, consistent with our results, and with carotid-femoral
382 pulse-wave velocity, a measure of aortic stiffness (38). Although 4-cresyl sulfate has never
383 been formally linked to BP, its dietary excretion has been shown to be highly correlated with
384 that of phenylacetylglutamine (16).

385 The association between gut-microbial co-metabolites and BP is further evidenced in
386 the direct association we found between BP and hippurate, originating from the conversion of
387 benzoic acid by gut microflora via the shikimate pathway (39). In contrast to our results,
388 hypertensive rats showed an anti-correlation between hippurate and BP (40) but interpolation
389 from animal data to human must be performed with care due to the differences in the gut
390 microbiome between species. An inverse association between excretion of hippurate and BP
391 has been reported in humans but this association was not significant after adjusting for body-
392 mass-index, alcohol intake, and urinary excretion of sodium and potassium (41). A controlled
393 feeding study by Wu et al showed that changes in the gut microbiome occurred within 24-h of
394 initiating a change in diet (35) and that body-mass-index and weight loss can also influence
395 the gut-microbiome composition. However, in our dietary intervention study, all participants
396 consumed a consistent healthy dietary pattern for 6 weeks and maintained their body weight,
397 with micronutrients being indexed to the energy level of their diets. Our data, therefore,
398 suggest modulation of diets can affect gut microbiome activity and that this may lead to a
399 direct effect on BP regulation.

400 We observed an inverse association of N-methyl-2-pyridone-5-carboxamide
401 (tryptophan-NAD metabolite) and BP. Bartus et al showed that ingestion of 1-
402 methylnicotinamide in hypertriglyceridemic rats resulted in an increase of 1-
403 methylnicotinamide and its metabolites such as N-methyl-2-pyridone-5-carboxamide and
404 found that ingestion of 1-methylnicotinamide in both the diabetic and hypertriglyceridemia
405 rats can ameliorate the nitric oxide dependent vasodilation, a surrogate marker for
406 atherosclerosis (42). Others have found that 1-methylnicotinamide demonstrates anti-
407 thrombotic activity (43). Our findings further support the beneficial impact of N-methyl-2-
408 pyridone-5-carboxamide on CVD health. We suggest, the tryptophan-NAD pathway may
409 offer a new target for pharmacological treatment of hypertension.

410 We also confirmed the association of dietary markers with BP including: a direct
411 association between BP and carnitine (a marker for protein ingestion); and an inverse
412 association with proline-betaine (citrus fruit ingestion). Our results are consistent with
413 previous studies linking hypertension with blood concentration of carnitine (44) and
414 variations in BP following carnitine treatment in rats (45). Similarly our results support the
415 previously postulated benefit of citrus fruit intake in reduction of BP (34). Specifically for the
416 OmniProt diet, despite the increased excretion of carnitine, a marker which was linked to
417 higher BP, overall beneficial reductions in CVD risk factors (both BP levels and lipid
418 profiles) was elicited and these benefits persisted for those who were considered as typical
419 (HDR) as well as variable (VDR) responders. The specific mechanisms for this remain
420 unclear although it may be hypothesized that the altered large-bowel microbiome following
421 protein rich dietary intervention may play a significant role.

422 We investigated our data stratified by responders (HDR groups) and non-responders
423 (VDR groups) to ascertain whether the lack of demonstrated response was purely due to poor
424 adherence to diet. We used a marker of citrus fruits, proline-betaine, as a proxy for dietary
425 adherence to OmniHeart diets, as participants were given citrus fruits as part of their diets.
426 Using the level of proline-betaine excretion at <95% CI of the HDR groups as a cutoff, we
427 estimated non-adherence contributed to the dampened clinical responses for 9.5% to 22.2% of
428 the participants, depending on the type of OmniHeart diet. These non-adherence values are
429 considerably higher than the <5% non-adherence estimated from the self-reported data from
430 this study (4) and provided an additional objective measure to the mean urine urea nitrogen
431 measurements, reflecting protein intake, which was highest on the protein rich diet. Our
432 modeling strategy thus provided an objective method for classification of individuals in the
433 VDR groups as non-adherent to each of the OmniHeart diets. The remaining discrepancy in
434 metabolic response in individuals showing good dietary adherence was mainly attributable to

435 variation in the excretion of gut microbial metabolites (7.6%). These results are consistent
436 with findings from a recent study by Zeevi et al (46) who showed inter-individual differences
437 in glycaemic response to foods and that this was correlated with differences in the
438 composition of the microbiome.

439 As a feeding study, this study has several strengths including: the provision of all meals
440 to participants where their body weights were held constant throughout the feeding periods,
441 thereby removing the confounding effect of weight loss; the inclusion of 24-h urine collection;
442 and the randomized cross-over design all add rigor to the study. Further, we have included
443 individuals from high CVD risk groups such as African American (~50%) and pre-hypertensive
444 patients (~80%), which strengthens the general applicability of our stratification pipeline,
445 although we recognize large proportion of our participants were either overweight or obese and
446 therefore not reflective of the general population. However, this reflects the higher incidence
447 of obesity among the African American. Since, by design, participants' weight remained the
448 same throughout the study, our models were not adjusted for body-mass-index. We also did
449 not adjust for socioeconomic status based on previous findings in a large scale cross sectional
450 study, which demonstrated that the inverse association with BP was explained mostly by dietary
451 differences (47).

452 Our study represents one of the largest dietary interventions of its kind where many
453 prior nutritional metabolic phenotyping studies have typically involved a small number of
454 participants (N< 25) (48, 49). In this study, we used food frequency questionnaires to describe
455 participants' food intake during the screening visit (baseline) and this information was used to
456 estimate the average intended food intake to maintain the participants' body weight throughout
457 the isocaloric feeding periods. However, one limitation was that we were unable to perform
458 more detailed analysis on individual dietary components and the dose-response relationship
459 with BP. An additional limitation of the current study was the use of NMR spectroscopy as the

460 sole method of metabolic profiling. Although the robustness of the technique is advantageous
461 for generating high quality data, mass spectrometry would offer better sensitivity and selectivity
462 and may have identified further candidate biomarkers relating to BP. Nonetheless, we were
463 able to uncover a number of biomarkers related to BP and these biomarkers were structurally
464 authenticated.

465 In this global profiling study, we opted to use urine as our choice of biofluid as urine
466 contains rich source of information encompassing the influence of dietary and gut microbiota.
467 We and others (41, 50) have successfully identified urinary discriminatory metabolites related
468 to BP. However, future studies should validate our findings by the use of urine specimens
469 collected from independent epidemiological studies. Further to validating the candidate
470 biomarkers related to dietary modulation of BP, a series of in vivo studies to establish causality
471 would be necessary. For example, Menni et al have shown a possible causal relationship
472 between hexadecanedioate with BP using rodent models (51).

473 Our strategy illustrates the feasibility of adopting a rational stratification approach for
474 diabetologists/cardiologists/dieticians to identify individuals' non-adherence to diets and to
475 optimize clinical responses to therapy. Extending this concept, we can envisage that further
476 characterization of inter-individual responses to healthy diets as determined by an individual's
477 phenotypic patterns and further determining an individual's longitudinal phenotypic stability
478 prior to a healthy dietary intervention would need to be developed for the identification of latent
479 sub-phenotypes. This may confer a public health benefit with potential to provide a personalized
480 approach to dietary recommendations aimed at optimizing prevention of CVD and related
481 disorders.

482 In conclusion, variation in metabolic phenotypes in response to specific healthy diets
483 may hold clues as to the mechanisms underlying inter-individual variations in response to
484 dietary modulation and points the potential importance of the gut microbiome in accounting for

485 differences in dietary response and the subsequent impact on BP. The workflow presented here
486 provides a clinically actionable framework to develop tailored dietary interventions designed
487 to reduce BP and other CVD risk factors.

488

489 **Acknowledgments:** We thank Miss T. Yap for her contribution to the sample preparation for
490 NMR analyses. This manuscript was prepared using OmniHeart research materials obtained
491 from the NHLBI Biologic Specimens and Data Repository Information Coordinating Centre
492 and does not necessarily reflect the opinions or views of the NHLBI. The authors would like
493 to thank the Imperial-National Institute for Health Research (NIHR) Clinical Phenome
494 Centre, which is supported by the NIHR Imperial Biomedical Research Centre based at
495 Imperial College Healthcare National Health Service (NHS) Trust and Imperial College
496 London. The views expressed are those of the author(s) and not necessarily those of the NHS,
497 the NIHR, or the Department of Health.

498

499 **Conflict of Interest:** None

500

501 **Authors' contributions:**

502 RLL: designed metabolic profiling research;

503 LJA designed OmniHeart research;

504 RLL and XZ: conducted the research;

505 RLL and XZ analyzed data;

506 RLL, EH and XZ: wrote the manuscript;

507 RLL had primary responsibility for final content;

508 EH and JKN facilitated access to MRC-NIHR National Phenome Centre and related work;

509 RLL, EH and JKN conducted metabolite identification.

510 All authors reviewed and approved the manuscript.

511

512 **Data and materials availability:** The OmniHeart study description together with the study

513 protocol and associated metadata are available from the Biologic Specimen and Data

514 Repository Information Coordinating Center (BioLINCC) at

515 https://biolincc.nhlbi.nih.gov/static/studies/omniheart/MOP.pdf?link_time=2017-07-

516 02_01:45:33.646682.

REFERENCES

1. Global Disease Burden Risk Factor Collaborators. Global, regional, and national levels of maternal mortality, 1990-2015: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet* 2016;388(10053):1775-812.
2. Ehret GB. Genome-wide association studies: contribution of genomics to understanding blood pressure and essential hypertension. *Curr Hypertens Rep* 2010;12(1):17-25.
3. Appel LJ, Moore TJ, Obarzanek E, Vollmer WM, Svetkey LP, Sacks FM, Bray GA, Vogt TM, Cutler JA, Windhauser MM, et al. A clinical trial of the effects of dietary patterns on blood pressure. DASH Collaborative Research Group. *N Engl J Med* 1997;336(16):1117-24.
4. Appel LJ, Sacks FM, Carey VJ, Obarzanek E, Swain JF, Miller ER, Conlin PR, Erlinger TP, Rosner BA, Laranjo NM, et al. Effects of protein, monounsaturated fat, and carbohydrate intake on blood pressure and serum lipids: results of the OmniHeart randomized trial. *Jama* 2005;294(19):2455-64.
5. Sleiman D, Al-Badri MR, Azar ST. Effect of mediterranean diet in diabetes control and cardiovascular risk modification: a systematic review. *Frontiers in public health* 2015;3:69.
6. Katan MB, Beynen AC, de Vries JH, Nobels A. Existence of consistent hypo- and hyperresponders to dietary cholesterol in man. *Am J Epidemiol* 1986;123(2):221-34.
7. Jacobs DR, Jr., Anderson JT, Hannan P, Keys A, Blackburn H. Variability in individual serum cholesterol response to change in diet. *Arteriosclerosis* 1983;3(4):349-56.
8. Mata P, Ordovas JM, Lopez-Miranda J, Lichtenstein AH, Clevidence B, Judd JT, Schaefer EJ. ApoA-IV phenotype affects diet-induced plasma LDL cholesterol lowering. *Arteriosclerosis and thrombosis : a journal of vascular biology* 1994;14(6):884-91.
9. McCombs RJ, Marcadis DE, Ellis J, Weinberg RB. Attenuated hypercholesterolemic response to a high-cholesterol diet in subjects heterozygous for the apolipoprotein A-IV-2 allele. *N Engl J Med* 1994;331(11):706-10.
10. Masson LF, McNeill G, Avenell A. Genetic variation and the lipid response to dietary intervention: a systematic review. *Am J Clin Nutr* 2003;77(5):1098-111.
11. Denke MA. Review of human studies evaluating individual dietary responsiveness in patients with hypercholesterolemia. *Am J Clin Nutr* 1995;62(2):471S-7S.
12. Denke MA, Adams-Huet B, Nguyen AT. Individual cholesterol variation in response to a margarine- or butter-based diet: A study in families. *JAMA* 2000;284(21):2740-7.
13. Faith JJ, McNulty NP, Rey FE, Gordon JI. Predicting a human gut microbiota's response to diet in gnotobiotic mice. *Science* 2011;333(6038):101-4.
14. Minot S, Sinha R, Chen J, Li H, Keilbaugh SA, Wu GD, Lewis JD, Bushman FD. The human gut virome: inter-individual variation and dynamic response to diet. *Genome research* 2011;21(10):1616-25.
15. Pelantova H, Buganova M, Holubova M, Sediva B, Zemenova J, Sykora D, Kavalkova P, Haluzik M, Zelezna B, Maletinska L, et al. Urinary metabolomic profiling in mice with diet-induced obesity and type 2 diabetes mellitus after treatment with metformin, vildagliptin and their combination. *Molecular and cellular endocrinology* 2016;431:88-100.
16. Heinzmann SS, Merrifield CA, Rezzi S, Kochhar S, Lindon JC, Holmes E, Nicholson JK. Stability and robustness of human metabolic phenotypes in response to sequential food challenges. *J Proteome Res* 2012;11(2):643-55.
17. O'Keefe SJ, Li JV, Lahti L, Ou J, Carbonero F, Mohammed K, Posma JM, Kinross J, Wahl E, Ruder E, et al. Fat, fibre and cancer risk in African Americans and rural Africans. *Nature communications* 2015;6:6342.
18. Everett JR, Loo RL, Pullen FS. Pharmacometabonomics and personalized medicine. *Ann Clin Biochem* 2013;50(Pt 6):523-45.
19. Carey VJ, Bishop L, Charleston J, Conlin P, Erlinger T, Laranjo N, McCarron P, Miller E, Rosner B, Swain J, et al. Rationale and design of the Optimal Macro-Nutrient Intake Heart Trial to Prevent Heart Disease (OMNI-Heart). *Clin Trials* 2005;2(6):529-37.
20. Rimm EB, Giovannucci EL, Stampfer MJ, Colditz GA, Litin LB, Willett WC. Reproducibility and validity of an expanded self-administered semiquantitative food frequency questionnaire among male health professionals. *Am J Epidemiol* 1992;135(10):1114-26; discussion 27-36.
21. Beckonert O, Keun HC, Ebbels TM, Bundy J, Holmes E, Lindon JC, Nicholson JK. Metabolic profiling, metabolomic and metabonomic procedures for NMR spectroscopy of urine, plasma, serum and tissue extracts. *Nat Protoc* 2007;2(11):2692-703.
22. Dieterle F, Ross A, Schlotterbeck G, Senn H. Probabilistic quotient normalization as robust method to account for dilution of complex biological mixtures. Application in ¹H NMR metabonomics. *Anal Chem* 2006;78(13):4281-90.

23. Zou X, Holmes E, Nicholson JK, Loo RL. Statistical HOMogeneous Cluster Spectroscopy (SHOCSY): an optimized statistical approach for clustering of (1)H NMR spectral data to reduce interference and enhance robust biomarkers selection. *Anal Chem* 2014;86(11):5308-15.
24. Posma JM, Garcia-Perez I, De Iorio M, Lindon JC, Elliott P, Holmes E, Ebbels TM, Nicholson JK. Subset Optimization by Reference Matching (STORM): An Optimized Statistical Approach for Recovery of Metabolic Biomarker Structural Information from (1)H NMR Spectra of Biofluids. *Anal Chem* 2012;84(24):10694-701.
25. Cloarec O, Dumas ME, Craig A, Barton RH, Trygg J, Hudson J, Blancher C, Gauguier D, Lindon JC, Holmes E, et al. Statistical total correlation spectroscopy: an exploratory approach for latent biomarker identification from metabolic ¹H NMR data sets. *Anal Chem* 2005;77(5):1282-9.
26. Westerhuis JA, van Velzen EJ, Hoefsloot HC, Smilde AK. Multivariate paired data analysis: multilevel PLSDA versus OPLSDA. *Metabolomics* 2010;6(1):119-28.
27. van Velzen EJ, Westerhuis JA, van Duynhoven JP, van Dorsten FA, Hoefsloot HC, Jacobs DM, Smit S, Draijer R, Kroner CI, Smilde AK. Multilevel data analysis of a crossover designed human nutritional intervention study. *J Proteome Res* 2008;7(10):4483-91.
28. Trygg J, Wold S. O2-PLS, a two-block (X-Y) latent variable regression (LVR) method with an integral OSC filter. *Journal of Chemometrics* 2003;17(1):53-64.
29. Wishart DS, Tzur D, Knox C, Eisner R, Guo AC, Young N, Cheng D, Jewell K, Arndt D, Sawhney S, et al. HMDB: the Human Metabolome Database. *Nucleic Acids Res* 2007;35(Database issue):D521-6.
30. Heinzmann SS, Brown IJ, Chan Q, Bictash M, Dumas ME, Kochhar S, Stamler J, Holmes E, Elliott P, Nicholson JK. Metabolic profiling strategy for discovery of nutritional biomarkers: proline betaine as a marker of citrus consumption. *Am J Clin Nutr* 2010;92(2):436-43.
31. Stella C, Beckwith-Hall B, Cloarec O, Holmes E, Lindon JC, Powell J, vanderOuderaa F, Bingham S, Cross AJ, Nicholson JK. Susceptibility of Human Metabolic Phenotypes to Dietary Modulation. *J Proteome Res* 2006;5(10):2780-8.
32. Edmands WM, Beckonert OP, Stella C, Campbell A, Lake BG, Lindon JC, Holmes E, Gooderham NJ. Identification of human urinary biomarkers of cruciferous vegetable consumption by metabonomic profiling. *J Proteome Res* 2011;10(10):4513-21.
33. May DH, Navarro SL, Ruczinski I, Hogan J, Ogata Y, Schwarz Y, Levy L, Holzman T, McIntosh MW, Lampe JW. Metabolomic profiling of urine: response to a randomised, controlled feeding study of select fruits and vegetables, and application to an observational study. *Br J Nutr* 2013;110(10):1760-70.
34. Lloyd AJ, Beckmann M, Fave G, Mathers JC, Draper J. Proline betaine and its biotransformation products in fasting urine samples are potential biomarkers of habitual citrus fruit consumption. *Br J Nutr* 2011;106(6):812-24.
35. Wu GD, Chen J, Hoffmann C, Bittinger K, Chen YY, Keilbaugh SA, Bewtra M, Knights D, Walters WA, Knight R, et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science* 2011;334(6052):105-8. doi: 10.1126/science.1208344.
36. Yang T, Santisteban MM, Rodriguez V, Li E, Ahmari N, Carvajal JM, Zadeh M, Gong M, Qi Y, Zubevcic J, et al. Gut dysbiosis is linked to hypertension. *Hypertension* 2015;65(6):1331-40.
37. Khalesi S, Sun J, Buys N, Jayasinghe R. Effect of probiotics on blood pressure: a systematic review and meta-analysis of randomized, controlled trials. *Hypertension* 2014;64(4):897-903.
38. Menni C, Mangino M, Cecelja M, Psatha M, Brosnan MJ, Trimmer J, Mohney RP, Chowienczyk P, Padmanabhan S, Spector TD, et al. Metabolomic study of carotid-femoral pulse-wave velocity in women. *J Hypertens* 2015;33(4):791-6; discussion 6.
39. Pero RW. Health consequences of catabolic synthesis of hippuric acid in humans. *Curr Clin Pharmacol* 2010;5(1):67-73. doi: CCP-003 [pii].
40. Akira K, Masu S, Imachi M, Mitome H, Hashimoto T. A metabonomic study of biochemical changes characteristic of genetically hypertensive rats based on (1)H NMR spectroscopic urinalysis. *Hypertension research : official journal of the Japanese Society of Hypertension* 2012;35(4):404-12.
41. Holmes E, Loo RL, Stamler J, Bictash M, Yap IKS, Chan Q, Ebbels T, De Iorio M, Brown IJ, Veselkov KA, et al. Human metabolic phenotype diversity and its association with diet and blood pressure. *Nature* 2008;453(7193):396-400.
42. Bartus M, Lomnicka M, Kostogrysb RB, Kazmierczak P, Watala C, Slominska EM, Smolenski RT, Pisulewski PM, Adamus J, Gebicki J, et al. 1-Methylnicotinamide (MNA) prevents endothelial dysfunction in hypertriglyceridemic and diabetic rats. *Pharmacol Rep* 2008;60(1):127-38.
43. Chlopicki S, Swies J, Mogielnicki A, Buczko W, Bartus M, Lomnicka M, Adamus J, Gebicki J. 1-Methylnicotinamide (MNA), a primary metabolite of nicotinamide, exerts anti-thrombotic activity mediated by a cyclooxygenase-2/prostacyclin pathway. *British journal of pharmacology* 2007;152(2):230-9.

44. Mels CM, Schutte AE, Erasmus E, Huisman HW, Schutte R, Fourie CM, Kruger R, Van Rooyen JM, Smith W, Malan NT, et al. L-carnitine and long-chain acylcarnitines are positively correlated with ambulatory blood pressure in humans: the SABPA study. *Lipids* 2013;48(1):63-73.
45. Rauchova H, Dobesova Z, Drahotka Z, Zicha J, Kunes J. The effect of chronic L-carnitine treatment on blood pressure and plasma lipids in spontaneously hypertensive rats. *European journal of pharmacology* 1998;342(2-3):235-9.
46. Zeevi D, Korem T, Zmora N, Israeli D, Rothschild D, Weinberger A, Ben-Yacov O, Lador D, Avnit-Sagi T, Lotan-Pompan M, et al. Personalized Nutrition by Prediction of Glycemic Responses. *Cell* 2015;163(5):1079-94.
47. Stamler J, Elliott P, Appel L, Chan Q, Buzzard M, Dennis B, Dyer AR, Elmer P, Greenland P, Jones D, et al. Higher blood pressure in middle-aged American adults with less education-role of multiple dietary factors: the INTERMAP study. *J Hum Hypertens* 2003;17(9):655-775.
48. Bondia-Pons I, Canellas N, Abete I, Rodriguez MA, Perez-Cornago A, Navas-Carretero S, Zulet MA, Correig X, Martinez JA. Nutri-metabolomics: subtle serum metabolic differences in healthy subjects by NMR-based metabolomics after a short-term nutritional intervention with two tomato sauces. *Omics : a journal of integrative biology* 2013;17(12):611-8.
49. Lai S, Molfino A, Coppola B, De Leo S, Tommasi V, Galani A, Migliaccio S, Greco EA, Gnerre Musto T, Muscaritoli M. Effect of personalized dietary intervention on nutritional, metabolic and vascular indices in patients with chronic kidney disease. *European review for medical and pharmacological sciences* 2015;19(18):3351-9.
50. Hanson M, Zahradka P, Taylor CG, Aliani M. Identification of urinary metabolites with potential blood pressure-lowering effects in lentil-fed spontaneously hypertensive rats. *European journal of nutrition* 2016.
51. Menni C, Graham D, Kastenmuller G, Alharbi NH, Alsanosi SM, McBride M, Mangino M, Titcombe P, Shin SY, Psatha M, et al. Metabolomic identification of a novel pathway of blood pressure regulation involving hexadecanedioate. *Hypertension* 2015;66(2):422-9.

Table 1: Characteristics of participants completed all three OmniHeart diets (N = 158).

Characteristics		P values
Age, mean (SD)	53.1 (10.8)	
Ethnicity, N (%)		
African American	86 (54.4%)	
Non-African American	72 (45.6%)	
Gender, N (%)		
Male	88 (55.7%)	
Female	70 (44.3%)	
Hypertension, N (%)		
Pre-hypertension	127 (80.4%)	
Hypertension	31 (19.6%)	
Obesity status, N (%)		
Normal range	32 (20.3%)	
Overweight	53 (33.5%)	
Obese	73 (46.2%)	
Smoking, N (%)		
Current	18 (11.4%)	
Former	42 (26.6%)	
Never	98 (62%)	
Alcohol intake		
No alcohol, N (%)	88 (56%)	
Serving per week among drinker, mean \pm SD	4.17 \pm 3.5	
Education, N (%)		
\leq high school	32 (20.3%)	
Some college	53 (33.5%)	
College graduate	73 (46.2%)	
Mean changes of SBP from baseline (95% CI), mmHg		
OmniCarb diet	-8.0 (-9.4, -6.6)	‡
OmniMFA diet	-9.4 (-10.7, -8.1)	‡
OmniProt diet	-9.4 (-10.8, -8.1)	‡
Mean changes of DBP from baseline (95% CI), mmHg		
OmniCarb diet	-4.1 (-4.9, -3.3)	‡
OmniMFA diet	-4.9 (-5.7, -4.1)	‡
OmniProt diet	-5.3 (-6.1, -4.4)	‡
Mean changes of LDL from baseline (95% CI), mg/dL		
OmniCarb diet	-11.6 (-14.6, -8.6)	‡
OmniMFA diet	-13.2 (-16.5, -9.9)	‡
OmniProt diet	-14.4 (-17.7, -11.1)	‡
Mean changes of HDL from baseline (95% CI), mg/dL		
OmniCarb diet	-1.5 (-2.6, -0.3)	*
OmniMFA diet	-0.4 (-1.4, 0.6)	
OmniProt diet	-2.7 (-3.7, -1.7)	†
Mean changes of Triglyceride from baseline (95% CI), mg/dL		
OmniCarb diet	-0.2 (-9.1, 8.7)	
OmniMFA diet	-9.7 (-17.9, -1.5)	*
OmniProt diet	-16.5 (-25.8, -7.3)	*

Mean changes of total cholesterol from baseline (95% CI), mg/dL		
OmniCarb diet	-12.5 (-15.8, -9.1)	‡
OmniMFA diet	-15.6 (-19.2, -11.9)	‡
OmniProt diet	-20.2 (-23.7, -16.69)	‡

Abbreviations: N, number of individuals; SD, standard deviation; CI, confidence interval;

SBP, systolic blood pressure; DBP, diastolic blood pressure; HDL, high-density lipoprotein cholesterol; LDL, low-density lipoprotein cholesterol.

T-test comparison between baseline clinical data and after each dietary intervention: *

$p < 0.05$; † $p < 10^{-5}$; ‡ $p < 10^{-10}$.

Table 2: Estimated mean differences in CVD risk factors. The systolic and diastolic BP, LDL and total cholesterol mean differences per 2 standard deviation (SD) increase in absolute excretion for the comparison between baseline and post OmniHeart diets; and between different OmniHeart diets for the HDR groups.

Urinary metabolites	2SD excretion (mmol/L)	SBP (mmHg)	DBP (mmHg)	LDL (mg/dL)	Total Cholesterol (mg/dL)
Homogeneous dietary responder for OmniCarb diet vs Baseline (N=137)					
proline-betaine	1.25	-4.10 (-2.90) [†]	-1.77 (-2.15) [*]	-3.90 (-1.26)	-3.94 (-1.17)
Hippurate	3.47	6.14 (4.64) [‡]	2.27 (2.79) [†]	-1.47 (-0.48)	2.70 (0.80)
N-methyl-2-pyridone-5-carboxamide	0.21	-3.03 (-2.24) [*]	-1.77 (-2.19) [*]	2.22 (0.75)	3.49 (1.11)
N-methyl nicotinic acid	0.27	-0.20 (-0.14)	0.59 (0.71)	-1.94 (0.64)	-0.25 (-0.07)
N-methyl nicotinamide	0.03	-0.86 (-0.60)	-0.75 (-0.97)	0.60 (0.20)	-0.55 (0.17)
Homogeneous dietary responders for OmniMFA vs Baseline (N=127)					
proline-betaine	0.87	-3.53 (-2.76) [†]	-1.73 (-2.20) [*]	2.41 (0.73)	1.87 (0.48)
Homogeneous dietary responders for OmniProt vs Baseline (N=113)					
proline-betaine	0.74	-1.16 (0.74)	0.14 (0.16)	1.31 (0.33)	-1.42 (-0.33)
Carnitine	0.29	3.11 (1.99) [*]	1.13 (1.38)	0.45 (0.11)	1.03 (0.24)
Creatine	1.62	1.54 (1.05)	-1.19 (-0.24)	-4.34 (-1.11)	-6.87 (-1.65)
Homogeneous dietary responders for OmniCarb vs OmniMFA (N=113)					
Guanodinoacetate	0.95	0.29 (0.35)	0.06 (0.09)	-0.51 (-0.21)	2.70 (0.94)
Homogeneous dietary responders for OmniCarb vs OmniProt (N=134)					
Phenylacetylglutamine	0.70	-0.89 (-0.96)	-0.93 (-1.42)	-2.82 (-1.18)	-0.44 (-0.15)
4-cresyl sulfate	0.27	0.06 (0.07)	0.73 (1.11)	-0.91 (-0.79)	1.02 (0.35)
Homogeneous dietary responders for OmniMFA vs OmniProt (N=118)					

Phenylacetylglutamine	0.68	-1.7 (-1.93)	-1.89 (-3.32) [†]	0.31 (0.13)	-0.33 (-0.14)
4-cresyl sulfate	0.30	-2.68 (-3.05) [†]	-2.15 (-3.74) [†]	0.73 (0.31)	-0.27 (-0.11)

Abbreviations: N, number of individuals; SD, standard deviation; CI, confidence interval; SBP, systolic blood pressure; DBP, diastolic blood pressure; LDL, low-density lipoprotein cholesterol. Key: * p<0.05; [†] p<0.01; [‡] p<0.001

The correlation between changes of metabolites and CVD factors were evaluated by linear regression. 2SD excretion of each urinary metabolite was calculated by the absolute differences between dietary comparisons. Numbers in parenthesis are Z-scores, i.e. regression coefficient divided by standard error ($|Z\text{-score}| \geq 1.96$, p<0.05; ≥ 2.58 , p<0.01; ≥ 3.89 , p<0.001). NMR chemical shifts (multiplicity) used for quantification: proline-betaine, $\delta 3.11$ (singlet); hippurate, $\delta 7.64$ (triplet); N-methyl-2-pyridone-5-carboxamide, $\delta 6.67$ (doublet); N-methyl nicotinic acid, $\delta 4.44$ (singlet); N-methyl nicotinamide, $\delta 8.89$ (triplet); carnitine, $\delta 3.23$ (singlet); creatine, $\delta 3.93$ (singlet); guanodinoacetate, $\delta 3.80$ (singlet); phenylacetylglutamine, $\delta 7.43$ (triplet); 4-cresyl sulfate, $\delta 2.35$ (singlet).

Table 3: Stratification by urinary phenotypes. Individuals were stratified based on diet-specific urinary phenotypes.

Summary of sub-phenotypes	N	%
Group 1: HDR to all three diets	95	60.1%
Group 2: HDR to two diets but VDR to one diet	35	22.2%
a) Non-adherent-VDR to the other diet	25	15.8%
b) Adherent-VDR to the other diet	10	6.3%
Group 3: HDR to one diet but VDR to two diets	22	13.9%
a) Non-adherent-VDR to the other two diets	11	7.0%
b) Adherent-VDR to the other two diets	2	1.2%
c) Mixed response – non-adherence-VDR to one diet and adherence-VDR to the other diet	9	5.7%
Group 4: Non-adherence-VDR to all three diets	4	2.5%
Group 5: Mix of non-adherence and adherence to all three diets	2	1.3%

Figure 1: The observed mean differences in excretion for hippurate between homogeneous (HDR), adherent- and non-adherent variable dietary response (VDR) groups when OmniHeart diets and their corresponding baseline spectra were compared. Open squares, OmniCarb-HDR (N=137); light-grey closed squares, adherent- OmniCarb-VDR (N=6); dark-grey closed square, non-adherent-OmniCarb-VDR (N=15); open circle, OmniMFA-HDR (N=127); light-grey closed circle, adherent- OmniMFA-VDR (N=10); dark-grey closed circle, non-adherent-OmniMFA-VDR (N=21); open triangle, OmniProt-HDR (N=113); light-grey closed triangle, adherent- OmniProt-VDR (N=10); dark-grey closed triangle, non-adherent-OmnProt-VDR (N=35). Error bars indicate 95% confidence interval. Significant t-test comparison between baseline and post OmniHeart diets: * $p < 0.05$; † $p < 10^{-5}$; ‡ $p < 10^{-10}$.

Figure 2: Key observations for changes in the cardiovascular disease risk factors showing differences in homogeneous and variable dietary response groups for the comparisons between each OmniHeart diet and baseline corresponding to the changes in: (A) systolic BP; (B) diastolic BP; (C) LDL; (D) HDL; (E) triglycerides; (F) total cholesterol. Open squares, OmniCarb-HDR (N=137); light-grey closed squares, adherent- OmniCarb-VDR (N=6); dark-grey closed square, non-adherent-OmniCarb-VDR (N=15); open circle, OmniMFA-HDR (N=127); light-grey closed circle, adherent- OmniMFA-VDR (N=10); dark-grey closed circle, non-adherent-OmniMFA-VDR (N=21); open triangle, OmniProt-HDR (N=113); light-grey closed triangle, adherent- OmniProt-VDR (N=10); dark-grey closed triangle, non-adherent-OmnProt-VDR (N=35). Error bars indicate 95% confidence interval. Missing data include: LDL (N=2 for OmniMFA-VDR, OmniProt-VDR, OmniMFA-HDR, OmniProt-HDR and N=3 for OmniCarb-HDR); HDL (N=1 for OmniMFA-VDR and OmniProt-VDR); triglycerides (N=1 for OmniMFA-VDR and OmniProt-VDR); and total cholesterol (N=1 for OmniMFA-

VDR and OmniProt-VDR). Significant t-test comparison between baseline and post OmniHeart diets: * $p < 0.05$; † $p < 10^{-5}$; ‡ $p < 10^{-10}$.