

# 1 Seasonal variation in environmental DNA in relation to population 2 size and environmental factors

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## 10 Supplementary Material

### 11 Extraction protocols

12 The glass-microfiber filters were extracted in a fume hood, sterilised with a 10% bleach  
13 solution and UV-light the filter paper was removed from the sealed syringe filter holder using  
14 sterilised wire cutters and sterilised forceps. Once removed the filters were cut into strips  
15 approximately 3mm in width with each filter placed length ways into a separate 1.5 mL  
16 microcentrifuge tube. As a result for the digestion step each sample consisted of two  
17 microcentrifuge tubes, one for each of the two filters. 675 µL of the ATL buffer from the DNeasy  
18 Blood & Tissue kit was added to each tube; it was then vortexed for 15 seconds to mix before  
19 20 µL of Pro K was added and again vortexed. The samples were then incubated on a rotating  
20 block, for 3 hours at 56°C or overnight at 37°C. Following incubation the liquid was then  
21 transferred to a fresh microcentrifuge tube; the two digestion reactions for a sample were  
22 combined at this stage. 200 µL of AL buffer and 200 µL of ice cold absolute ethanol was added  
23 to each tube and vortexed for 15 seconds to facilitate DNA precipitation. 650 µL of the  
24 extraction solution was transferred into a DNeasy Blood and Tissue kit Mini spin column and

25 centrifuged at 8000rpm for one minute with the flow through discarded. This was then repeated  
26 using the same mini spin column until the entire sample had been passed through. DNA  
27 extraction continued as per the DNeasy Blood and Tissue kit manufacturers' protocol, eluting  
28 into 200 µL of the elution buffer.

29 Extraction from ethanol precipitation samples was undertaken using a modified protocol from  
30 Biggs (*et al.*)<sup>1</sup>. The mixture was centrifuged at 10,020g, (8500rpm) for 35 minutes and the  
31 supernatant discarded. The remainder of the extraction protocol was conducted as per Biggs  
32 (*et al.*)<sup>1</sup>.

### 33 [References](#)

34 1. Biggs, J. *et al.* Using eDNA to develop a national citizen science-based monitoring  
35 programme for the great crested newt (*Triturus cristatus*). *Biol. Conserv.* **183**, 19–28  
36 (2015).

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