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King, B F and Townsend-Nicholson, A and Wildman, Scott S.P. and Thomas, T and Spyer, K M and Burnstock, G (2000) Coexpression of rat P2X2 and P2X6 subunits in *Xenopus* oocytes. *The Journal of neuroscience : the official journal of the Society for Neuroscience*, 20 (13). pp. 4871-4877.

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Coexpression of Rat P2X₂ and P2X₆ Subunits in *Xenopus* Oocytes

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Transcripts for P2X₂ and P2X₆ subunits are present in rat CNS and frequently colocalize in the same brainstem nuclei. When rat P2X₂ (rP2X₂) and rat P2X₆ (rP2X₆) receptors were expressed individually in *Xenopus* oocytes and studied under voltage-clamp conditions, only homomeric rP2X₂ receptors were fully functional and gave rise to large inward currents (2–3 μ A) to extracellular ATP. Coexpression of rP2X₂ and rP2X₆ subunits in *Xenopus* oocytes resulted in a heteromeric rP2X_{2/6} receptor, which showed a significantly different phenotype from the wild-type rP2X₂ receptor. Differences included reduction in agonist potencies and, in some cases (e.g., Ap₄A), significant loss of agonist activity. ATP-evoked inward currents were biphasic at the heteromeric rP2X_{2/6} receptor, particularly when Zn²⁺ ions were present or extracellular pH was lowered. The pH range

was narrower for H⁺ enhancement of ATP responses at the heteromeric rP2X_{2/6} receptor. Also, H⁺ ions inhibited ATP responses at low pH levels (<pH 6.3). The pH-dependent blocking activity of suramin was changed at this heteromeric receptor, although the potentiating effect of Zn²⁺ on ATP responses was unchanged. Thus, the rP2X_{2/6} receptor is a functionally modified P2X₂-like receptor with a distinct pattern of pH modulation of ATP activation and suramin blockade. Although homomeric P2X₆ receptors function poorly, the P2X₆ subunit can contribute to functional heteromeric P2X channels and may influence the phenotype of native P2X receptors in those cells in which it is expressed.

Key words: P2X receptor; ionotropic receptor; heteromer; ATP; purinergic; oocyte

P2X receptors are ligand-gated cation channels that when activated by extracellular ATP mediate fast excitation in various cells, including central and peripheral neurons (Burnstock, 1997). Neuronal P2X receptors show considerable differences in their sensitivity to naturally occurring agonists, P2 receptor antagonists, and allosteric modulators and, furthermore, show differences in kinetics of receptor activation and inactivation (Khakh et al., 1995; King, 1998). Such diversity in the operational profiles of ATP-gated ion channels may be attributable to the subunit composition of native P2X receptors, because other classes of ionotropic receptors show differing phenotypes that depend on subunit composition (Barnard et al., 1998). Seven P2X receptor subunits (P2X_{1–7}) have been cloned, each of which is believed to form functional homomeric assemblies (Buell et al., 1996). They can also coassemble with other P2X subunits to form heteromeric P2X receptors of three, or possibly four, protein subunits per ATP-gated ion channel (Kim et al., 1997; Nicke et al., 1998; Torres et al., 1999). Three functional heteromeric P2X receptors have been reported: P2X_{2/3} (Lewis et al., 1995; Radford et al., 1997), P2X_{4/6} (Lê et al., 1998), and P2X_{1/5} (Torres et al., 1998;

Haines et al., 1999; Lê et al., 1999). Heteromeric channels composed of splice variants of the same P2X subunit (e.g., mP2X₄ and mP2X_{4a}) can also generate a different phenotypic form of the wild-type P2X receptor (Townsend-Nicholson et al., 1999).

The potential for heteropolymerization among P2X_{1–7} receptor subunits was recently investigated using coimmunoprecipitation procedures (Torres et al., 1999). For P2X subunits concentrated in the CNS (namely P2X₂, P2X₄, and P2X₆) (Collo et al., 1996), epitope-tagged P2X₂ and P2X₆ subunits or P2X₄ and P2X₆ subunits (but not P2X₂ and P2X₄ subunits) were shown to form immunopositive heteromeric assemblies. The functional properties of heteromeric P2X_{4/6} receptors have been established (Lê et al., 1998), but not yet the phenotype of heteromeric P2X_{2/6} receptors. The result of P2X₂ and P2X₆ subunit coexpression is of considerable interest because of (1) the distinct pH modulation of ATP responses at the homomeric P2X₂ receptor (King, 1998), (2) a growing belief that the P2X₆ subunit might only contribute to functional channels when other P2X subunits are present (Torres et al., 1999), and (3) the recent identification of a pH-modulated ATP receptor in those nuclei of rat brainstem where P2X₂ and P2X₆ transcripts have been detected (Thomas et al., 1999; Thomas and Spyer, 2000).

Thus, it was of interest to examine the contribution of the P2X₆ subunit, when coexpressed with the pH-modulated P2X₂ subunit, to the operational profile of the resultant heteromeric P2X_{2/6} receptor expressed in defolliculated *Xenopus* oocytes. Differences in the ways heteromeric P2X_{2/6} and homomeric P2X₂ receptors respond to nucleotidic agonists, suramin, pH, and Zn²⁺ ions were investigated in the oocyte expression system. The results establish the P2X_{2/6} receptor as the fourth example of a heteropolymeric ATP-gated ion channel that, in this case, possesses a pattern of pH modulation of ATP responses distinct from other known homomeric and heteromeric P2X receptors.

Received Dec. 2, 1999; revised April 10, 2000; accepted April 19, 2000.

This work was supported by grants from British Heart Foundation, Biotechnology and Biological Sciences Research Council (BBSRC, UK), and Roche Bioscience (Palo Alto, CA). We acknowledge Dr. David Julius (University of California San Francisco) and Dr. Gary Buell (Ares Sereno, Geneva) for the gifts of cDNAs encoding rP2X₂ and rP2X₆ subunits.

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MATERIALS AND METHODS

Oocyte preparation. *Xenopus laevis* frogs were killed by immersion in a lethal dose in Tricaine (0.4% w/v, in tap water) and then decapitated, and ovarian lobes were removed by blunt dissection. *Xenopus* oocytes (stages V and VI) were defolliculated by a two-step process involving (1) collagenase treatment (Type IA, 2 mg/ml in Ca²⁺-free Ringer's solution, for 2–3 hr) and (2) stripping away the follicle cell layer with fine forceps. Defolliculated oocytes do not possess native P1 and P2 receptors (King et al., 1996a,b) and are largely devoid of ecto-ATPases (Ziganshin et al., 1995). Oocytes were stored in Barth's solution (pH 7.5, at 4°C) containing (in mM): NaCl 110, KCl 1, NaHCO₃ 2.4, Tris HCl 7.5, Ca(NO₃)₂ 0.33, CaCl₂ 0.41, MgSO₄ 0.82, supplemented with gentamycin sulfate, 50 μg/l. Cells were injected cytosolically with cRNA for rP2X₂ (40 nl, 0.002 μg/μl) or rP2X₆ (40 nl, 1 μg/μl) or both rP2X₂ and rP2X₆ (40 nl of each) and incubated for 48 hr at 18°C in Barth's solution. Thereafter, injected oocytes were kept at 4°C for up to 12 d until they were used in electrophysiological experiments.

Electrophysiology. Membrane currents were recorded from cRNA-injected oocytes using a twin-electrode voltage-clamp amplifier (Axoclamp 2A). The holding potential (V_h) was -50 mV, unless stated otherwise. The voltage-recording and current-recording microelectrodes (1–5 MΩ tip resistance) were filled with 3.0 M KCl. Oocytes were placed in an electrophysiological chamber (volume, 0.5 ml) and superfused with Ringer's solution (5 ml/min, at 18°C) containing (in mM): NaCl 110, KCl 2.5, HEPES 5, CaCl₂ 1.8, adjusted to pH 7.5. Extracellular pH (pH_e) was adjusted with HCl (1.0N) or NaOH (1.0N) to reach the desired level. Electrophysiological data were stored on magnetic tape using a DAT recorder (Sony 1000ES) and displayed using a pen recorder (Gould 2200S).

Drug solutions. ATP and other nucleotides were prepared in Ringer's solution, and the pH of stock solutions was readjusted to the desired level. Agonists were superfused, at the concentrations given in the text, by a gravity-feed continuous flow system allowing the rapid addition and washout of drugs. ATP was added for 120 sec or until the current reached a peak, then washed off with Ringer's solution for a period of 5 min. Where used, antagonists were applied for 5 min before and during the application of agonists.

Agonist responses were normalized to the maximum inward current (I_{max}) evoked by ATP at pH 7.5, including agonist responses recorded at lower pH levels. At pH 7.5, maximum responses were evoked by 300–1000 μM ATP. The agonist concentration required to evoke 50% of the maximum response (EC_{50}) was taken from Hill plots, using the transform $\log(I/I_{max} - I)$, where I is the peak current evoked by each concentration of ATP.

The potentiating effects of extracellular Zn²⁺ ions on agonist activity were investigated in two ways. Zn²⁺ ions were either applied simultaneously with ATP or added to the Ringer's solution for 5 min before ATP was applied (with Zn²⁺ present).

Statistics and graphs. Data are presented as mean ± SEM of four to seven sets of data from different oocyte batches. Concentration–response curves and inhibition curves were fitted by nonlinear regression analysis using Prism v2.0 (GraphPad). Significant differences were determined by unpaired Student's *t* test or one-way ANOVA followed by Dunnett's *post hoc* test, again using Prism v2.0 (GraphPad).

Drugs and reagents. All common salts and reagents were AnalaR grade (Aldrich Chemicals, Poole, UK). ATP and ATPαS were purchased from Boehringer (Mannheim, Germany). 2-Methylthio ATP (2-MeSATP) was obtained from RBI (Natick, MA), and other nucleotides [ATPγS, ADP, AMP, adenosine, UTP, UDP, UMP, uridine, CTP, GTP, ITP, diadenosine polyphosphates (Ap_nA; $n = 2-6$), α,β-meATP, β,γ-meATP, and 2'- and 3'-O-(4-benzoyl-benzoyl)ATP (BzATP)] came from Sigma (Poole, UK). Suramin was a gift from Bayer (Newbury, UK).

RESULTS

ATP responses of P2X receptors

In initial experiments, the functionality of homomeric rP2X₂ and rP2X₆ receptors expressed in *Xenopus* oocytes was tested against a near-saturating concentration of ATP (100 μM), according to available pharmacological data on homomeric P2X receptors (King, 1998). At a holding potential of -50 mV, ATP-activated rP2X₂ receptors produced fast-activating and slowly inactivating inward currents (1993 ± 147 nA; $n = 6$) (Fig. 1A,C). rP2X₆

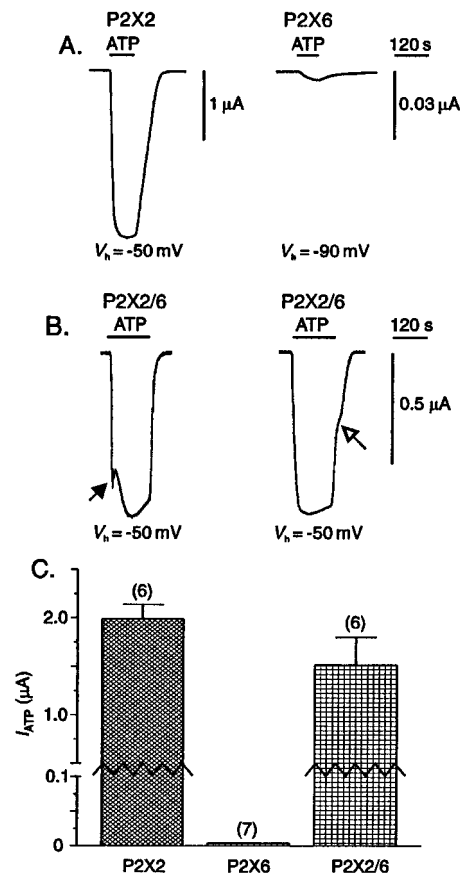


Figure 1. Expression of homomeric and heteromeric P2X receptors. *A*, Whole-cell inward currents by homomeric rP2X₂ and rP2X₆ receptors activated by a near-saturating ATP concentration (100 μM, for 60 sec), at the given holding potentials (V_h). *B*, Whole-cell inward currents by ATP-activated heteromeric rP2X_{2/6} receptors. ATP responses were often biphasic, showing a transient component (filled arrow) followed by a sustained current. The deactivation of inward current occasionally showed two phases of current decay (open arrow). *C*, Averaged whole-cell inward currents by homomeric rP2X₂, rP2X₆, and heteromeric rP2X_{2/6} receptors activated by ATP (100 μM). The y-axis of the histogram has been truncated to help reveal the small responses by rP2X₆ receptors. Data are expressed as mean ± SEM for six to seven cells per determination.

receptors failed to respond to ATP at a holding potential -50 mV, but where increased to -90 mV, the agonist did evoke low-amplitude slowly activating inward currents (4.57 ± 1.31 nA, $n = 7$) (Fig. 1A,C). Control (water-injected) oocytes failed to respond to ATP, at either -50 or -90 mV.

In further experiments, coexpression of rP2X₂ and rP2X₆ subunits resulted in fast-activating and slowly inactivating inward currents (1516 ± 286 nA, $n = 6$) (Fig. 1B,C), which were broadly similar in their time course to the ATP responses produced by homomeric rP2X₂ receptors. However, ATP-activated heteromeric rP2X_{2/6} receptors uniquely showed biphasic (transient and sustained) components to the evoked inward currents (Fig. 1B, closed arrow). Such biphasic responses were seen in all cRNA-injected oocytes tested ($n = 175$), although the amplitude of each component of biphasic currents was variable from response to response. Furthermore, evoked responses would change in an unpredictable manner from biphasic to monophasic currents (and back again) over several successive ATP applications. However, the reproducibility of biphasic inward currents by rP2X_{2/6} recep-

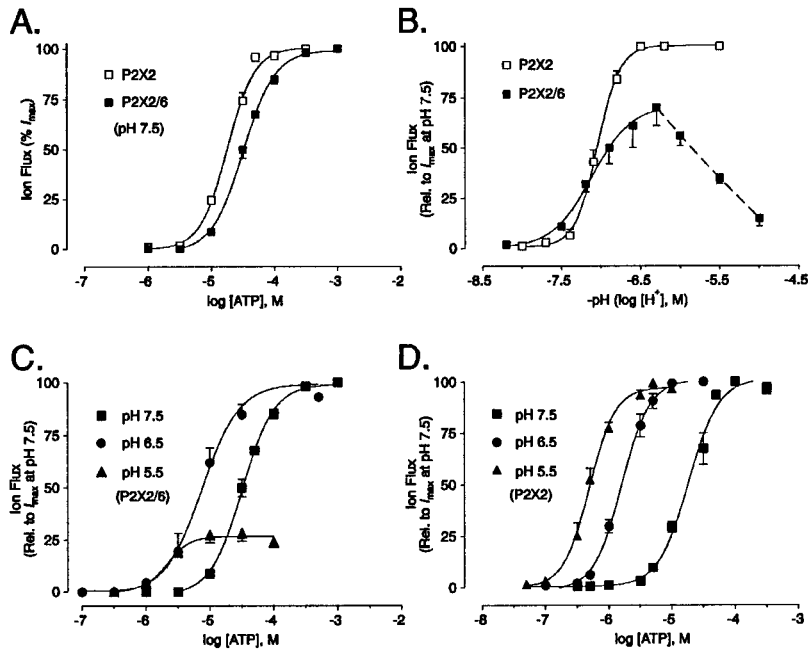


Figure 2. ATP activity at homomeric and heteromeric P2X receptors. *A*, C–R relationship for ATP-activated inward currents at rP2X₂ and rP2X_{2/6} receptors, at pH 7.5. *B*, The relationship between the amplitude of ATP responses (rP2X₂, 3 μ M; rP2X_{2/6}, 10 μ M; each producing 5% of the maximum response) and the extracellular pH level (range, pH 8.3–5.0) at homomeric and heteromeric P2X receptors. *C*, The C–R curves for ATP activation of rP2X_{2/6} receptors at the pH_e levels indicated. ATP efficacy was markedly reduced at pH 5.5. *D*, The C–R curves for ATP activation of rP2X₂ receptors. ATP efficacy was not altered at pH 5.5. Curves were fitted by the Hill equation in *A–D* (solid lines) and by a single exponential function in *B* (dashed line). Data given as mean \pm SEM for four to six cells per curve.

tors was enhanced when pH_e was lowered or Zn²⁺ ions were present in the bathing solution (see Fig. 5). Deactivation of rP2X_{2/6} receptors frequently comprised two phases of current decay (Fig. 1*B*, open arrow). Neither biphasic inward currents nor biphasic current decays were seen at rP2X₂ receptors.

The concentration–response (C–R) relationship was studied for ATP responses at rP2X₂ and rP2X_{2/6} receptors, at pH 7.5 (Fig. 2*A*). ATP was more potent (approximately twofold) at rP2X₂ receptors (EC₅₀, 18 \pm 2.1 μ M; n_H = 2.0 \pm 0.2) than rP2X_{2/6} receptors (EC₅₀, 32 \pm 1.6 μ M; n_H = 1.7 \pm 0.2) (p < 0.05, unpaired t test). Since the potency of agonists at rP2X₂ receptors is strongly affected by pH_e, the above differences in ATP activity could potentially be attributed to incorrect pH_e measurements. However, rP2X₂ and rP2X_{2/6} receptors responded in different ways to changes in pH_e (Fig. 2*B*). The amplitude of ATP responses at rP2X₂ receptors increased over the range of pH 8.0 to 6.3 and was maintained at lower pH_e levels (up to pH 5.0). ATP responses at rP2X_{2/6} receptors initially increased in size over the range of pH 8.0 to 6.3, then decreased in amplitude as pH_e levels were lowered further. The pK_a value for the potentiating phase of the H⁺ effect was 7.04 \pm 0.05 (n = 4) at P2X_{2/6} receptors, a value not significantly different from that of P2X₂ receptors (7.05 \pm 0.05; n = 4). However, the slopes of the curves describing the potentiating H⁺ effect were significantly different (P2X_{2/6}, 1.83 \pm 0.31; P2X₂, 3.04 \pm 0.22; p < 0.05).

The C–R relationship for ATP was reexamined at different pH_e levels for rP2X_{2/6} receptors. ATP potency was increased fourfold at pH 6.5 and 15-fold at pH 5.5 (Table 1; see EC₅₀ values). The maximum response to ATP was unchanged at pH 6.5, but agonist efficacy was significantly reduced (by 76 \pm 3%) at pH 5.5 (Fig. 2*C*). At rP2X₂ receptors, acidification of the bathing solution shifted the ATP C–R curve to the left without a reduction in the maximum (Fig. 2*D*). ATP potency was increased 12-fold at pH 6.5 and 30-fold at pH 5.5 at rP2X₂ receptors (Table 1; see EC₅₀ values). The effects of lowering pH_e were reversed on restoration to pH 7.5 for both rP2X₂ and rP2X_{2/6} receptors.

Agonist activity at P2X receptors

ATP, ATP α S, ATP γ S, and 2-meSATP are known to be full agonists at rP2X₂ receptors (King et al., 1997), and consequently their ability to activate rP2X_{2/6} receptors was investigated. Each nucleotide (30 μ M) elicited large, slowly inactivating inward currents at rP2X_{2/6} receptors, with an apparent potency order of (estimated EC₅₀ value) ATP (29.9 μ M) = ATP γ S (30.8 μ M) > 2-MeSATP (34.8 μ M) > ATP α S (40.6 μ M) (Fig. 3*A,C*). BzATP was a weak agonist at rP2X_{2/6} receptors (EC₅₀, 399 μ M) (Fig. 3*A,C*). P2X_{2/6} receptors did not respond to ADP, AMP, adenosine, UTP, UDP, UMP, uridine, CTP, GTP, ITP, α,β -meATP, and β,γ -meATP (each tested at 30 and 100 μ M) (data not shown). Of the diadenosine polyphosphates tested (Ap_nA, n = 2–6), Ap₄A alone showed activity but proved to be a weak agonist (EC₅₀, >1 mM) (Fig. 3*B,C*). This weak activity contrasted with results from rP2X₂ receptors, at which Ap₄A is a full agonist (EC₅₀, 15.2 μ M) (Pintor et al., 1996).

Suramin blockade at P2X receptors

Suramin is an effective antagonist at rP2X₂ receptors, at which its potency is enhanced when pH_e levels are lowered (King et al., 1997). Similar results were obtained for P2X_{2/6} receptors, with suramin reducing ATP responses in a concentration-dependent

Table 1. Effect of extracellular pH on ATP potency

pH _e	rP2X ₂ receptor	rP2X _{2/6} receptor
pH 7.5	16.2 \pm 1.4	32.0 \pm 1.6*
	(1.8 \pm 0.2)	(1.7 \pm 0.2)
pH 6.5	1.3 \pm 0.2	7.5 \pm 1.1*
	(2.0 \pm 0.2)	(1.5 \pm 0.3)
pH 5.5	0.55 \pm 0.04	2.2 \pm 0.7*
	(2.1 \pm 0.3)	(2.5 \pm 0.3)

EC₅₀ values (μ M) and Hill slopes (n_H, in brackets) for ATP activation of rP2X₂ and rP2X_{2/6} receptors at the given extracellular pH (pH_e) levels are shown. At each pH_e level tested, EC₅₀ values were significantly different (* p < 0.05, by unpaired t test). Data are expressed as mean \pm SEM (n = 4).

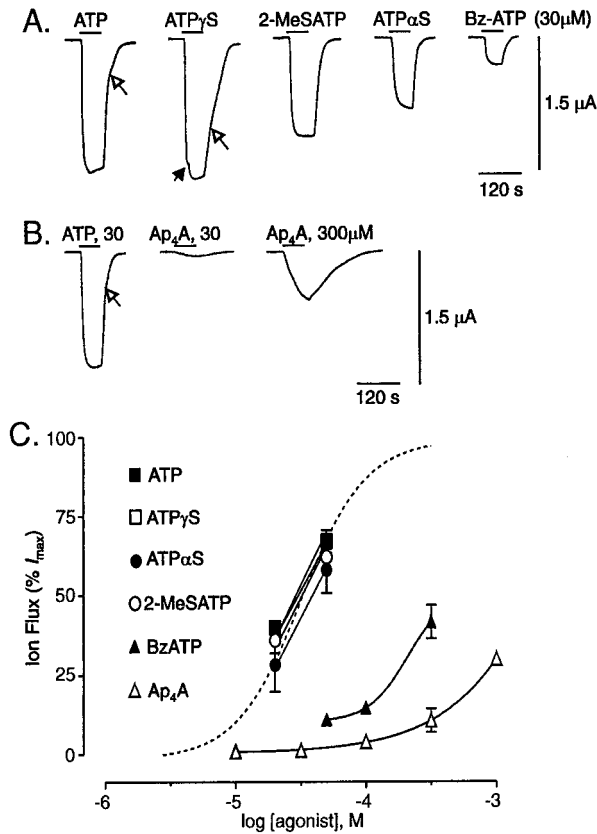


Figure 3. Nucleotide activation of rP2X_{2/6} receptors. In *A*, whole-cell inward currents at the heteromeric rP2X_{2/6} receptor were evoked by ATP, ATP γ S, 2-MeSATP, ATP α S, and BzATP (30 μ M), each of which is a known agonist of rP2X₂ receptors (King et al., 1997). In *B*, the rP2X_{2/6} receptor was activated weakly by Ap₄A (30 and 300 μ M), and the kinetics of activation and deactivation were considerably slower than ATP responses. *C*, C–R relationship for agonist activation of rP2X_{2/6} receptors at pH 7.5. Estimates of EC₅₀ values (micromolar concentration) were made using the “2 + 2 assay” method of Arunlakshana and Schild (1959); ATP, 29.9 \pm 1.9; ATP γ S, 30.8 \pm 2.9; ATP α S, 40.6 \pm 8.0; 2-MeSATP, 34.8 \pm 5.1; BzATP, 399 \pm 66; Ap₄A, >1000 (n = 4–6). The dashed line shows the position of the full C–R curve for ATP (redrawn from Fig. 2*A*). Open and filled arrows (in *A* and *B*) draw attention to biphasic components of receptor activation and deactivation. Data are given as mean \pm SEM for four to six cells per determination.

manner and its potency enhanced with acidification of the bathing solution (Fig. 4*A,B*). At pH 7.5, suramin was equipotent at rP2X₂ and rP2X_{2/6} receptors (Table 2; see IC₅₀ values). Differences in blocking activity were only observed at lower pH_e levels where, at pH 6.5, the inhibition curve for suramin was biphasic for rP2X_{2/6} receptors and monophasic for rP2X₂ receptors (Fig. 4*C*). Comparison of IC₅₀ values at pH 6.5 revealed that activity indices for each of the two phases of P2X_{2/6} receptor blockade was significantly different (p < 0.05, unpaired t test) compared with the IC₅₀ value for rP2X₂ receptors (Table 2). The blocking activity of suramin at rP2X₂ and rP2X_{2/6} receptors was reversed on washout, at all pH_e levels studied.

Actions of Zn²⁺ ions at P2X receptors

Extracellular Zn²⁺ is known to potentiate ATP responses at P2X₂ receptors (Wildman et al., 1998), although the degree of potentiation depends on whether Zn²⁺ is applied before, or simultaneously with, the agonist. When applied 5 min before ATP, Zn²⁺ ions (1–30 μ M) progressively increased ATP re-

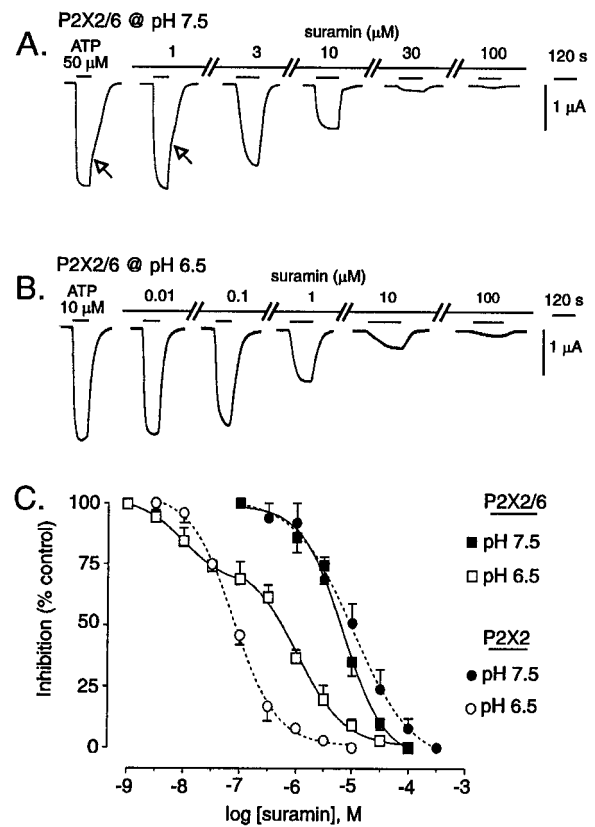


Figure 4. Suramin antagonism of rP2X_{2/6} receptors. Shown is antagonism of ATP responses (V_h = –50 mV) at heteromeric rP2X_{2/6} receptors by suramin at pH 7.5 (*A*) and pH 6.5 (*B*). Suramin was effective at micromolar concentrations at pH 7.5, but the concentration range for suramin blockade was extended at pH 6.5. *C*, Inhibition curves for suramin blockade of ATP responses at rP2X₂ and rP2X_{2/6} receptors at the given pH levels. At pH 6.5, the inhibition curve for rP2X_{2/6} was fitted best by a biphasic curve. IC₅₀ values are given in Table 2. Open arrows draw attention to biphasic current decays. Data are expressed as mean \pm SEM for four to eight cells per curve. The biphasic curve for rP2X_{2/6} was constructed from eight sets of data, using the results from the first two log₁₀ units of concentration (suramin, 0.001–0.01 μ M) to represent the first component of the inhibition curve.

sponses at rP2X_{2/6} receptors, by 6- to 14-fold (averaging 9.82 \pm 2.29, n = 6), whereas higher concentrations (30–300 μ M) progressively decreased and abolished ATP responses in a concentration-dependent manner (Fig. 5*A*). The potentiating and inhibitory effects were reversed on washout. Zn²⁺ preincubation also affected ATP responses by clearly increasing the incidence of biphasic inward currents, a phenomenon also seen when pH_e levels were lowered (Fig. 5*A,C*). Where applied simultaneously with ATP, Zn²⁺ ions (1–300 μ M) only caused a concentration-dependent increase (8- to 25-fold; averaging 16.35 \pm 4.28, n = 5) in the amplitude of ATP responses at P2X_{2/6} receptors (Fig. 5*B*). Without Zn²⁺ preincubation, the above inhibitory Zn²⁺ effect was not seen, and the incidence of biphasic ATP responses was inconsistent and infrequent for each oocyte tested. EC₅₀ values for the potentiating effects of Zn²⁺ ions at rP2X₂ and rP2X_{2/6} receptors were similar (Fig. 5, see legend).

DISCUSSION

In the present study, expression of homomeric rP2X₆ receptors in defolliculated *Xenopus* oocytes resulted in functional P2X receptors that, even under heightened conditions for channel activa-

Table 2. Blockade by suramin of P2X receptors

pH _e	rP2X ₂ receptor	rP2X _{2/6} receptor
pH 7.5	10.4 ± 1.2	6.06 ± 1.22
	(−0.83 ± 0.12)	(−1.19 ± 0.14)
pH 6.5	0.078 ± 0.005	0.013 ± 0.003 (<i>I</i> ₁)*
	(−1.19 ± 0.08)	(−1.28 ± 0.15)
		1.61 ± 0.28 (<i>I</i> ₂)*
		(−0.96 ± 0.16)

IC₅₀ values (μM) and Hill slopes (n_H, in brackets) for suramin blockade of rP2X₂ and rP2X_{2/6} receptors at the given extracellular pH (pH_e) levels are shown. The inhibition curve for suramin blockade of rP2X_{2/6} receptors was biphasic at pH 6.5, showing high-affinity (*I*₁) and low-affinity (*I*₂) components of blockade of ATP responses (Fig. 4C). The IC₅₀ values of rP2X₂ and rP2X_{2/6} receptors, at pH 6.5, were significantly different (**p* < 0.05, by unpaired *t* test). Data are expressed as mean ± SEM (*n* = 4 for rP2X₂ at pH 7.5 and 6.5, and for rP2X_{2/6} at pH 7.5; *n* = 8 for rP2X_{2/6} at pH 6.5).

tion, only managed to produce low-amplitude responses. Such weak ATP responses were attributed to the activation of just a small number of functional rP2X₆ receptors, because defolliculated oocytes do not possess native P1 or P2 receptors to complicate the analysis of agonist actions (King et al., 1996a,b). Homomeric rP2X₆ receptors have thus far been reported to function well in human embryonic kidney (HEK) 293 cells (Buell et al., 1996; Collo et al., 1996), to be silent in HEK 293 cells (Torres et al., 1999), or not to function at all in *Xenopus* oocytes (Soto et al., 1996; Lê et al., 1998). Our initial experiments thus confirmed that there are difficulties associated with rP2X₆ receptor expression in *Xenopus* oocytes and, in all probability, in other cell systems. It is possible that *Xenopus* oocytes and occasionally HEK 293 cells fail to produce an essential protein necessary to insert P2X₆ subunits into the cell membrane. One plausible candidate for this protein is another P2X subunit, perhaps the P2X₄ subunit, on the grounds that a P2X₄-like cDNA (AF012903) has been isolated from HEK 293 cells (direct submission GenBank by Chang and Chang in 1996) and the P2X₄ protein is present at low levels in these cells (Worthington et al., 1999). The heteromeric rP2X_{4/6} receptor is similar in its functional properties to the operational profile of homomeric rP2X₆ receptors (Lê et al., 1998). Because <5% of HEK 293 cells transfected with rP2X₆ cDNA go on to assemble a functional P2X₆ (or possibly P2X_{4/6}-like) receptor (Collo et al., 1996), the P2X₄ subunit may not be present in all HEK 293 cells.

Where coexpression of P2X₂ and P2X₆ subunits in *Xenopus* oocytes was concerned, our experiments were based on a comparison of the operational profiles of wild-type rP2X₂ receptors and heteromeric rP2X_{2/6} receptors. The rP2X₂ receptor has already been characterized in our laboratory in an extensive survey of agonists, antagonists, and modulators at this ATP-gated ion channel (King et al., 1996c, 1997; Pintor et al., 1996; Wildman et al., 1997, 1998, 1999a,b,c). Torres and colleagues (1999) have demonstrated that epitope-tagged rP2X₂ and rP2X₆ subunits will coprecipitate when expressed in a heterologous expression system. Thus, our present results confirm that functional heteromeric P2X_{2/6} receptors are indeed formed and inserted into the membrane of *Xenopus* oocytes. Several key observations were made on this new heteromeric P2X receptor, particularly (1) the nature of the evoked inward currents, (2) the potency of agonists, and (3) the effect of pH on ATP responses and suramin blockade.

ATP-evoked inward currents at heteromeric rP2X_{2/6} receptors were sometimes biphasic in nature, involving transient and sustained components that varied in amplitude from response to

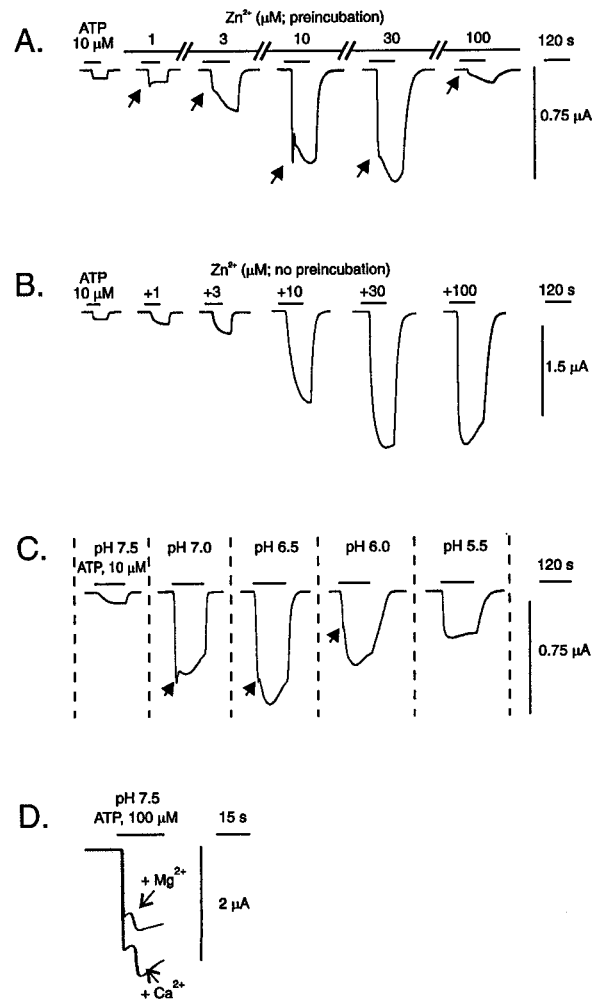


Figure 5. Modulation of ATP responses by Zn²⁺ and H⁺ at rP2X_{2/6} receptors. *A*, Concentration-dependent potentiation and inhibition of agonist-evoked inward currents by extracellular Zn²⁺ (1–100 μM) given 5 min before and during ATP application at rP2X_{2/6} receptors, at pH 7.5. EC₅₀ values (micromolar concentration) for Zn²⁺ potentiation of ATP responses were rP2X_{2/6}, 6.8 ± 1.0 versus rP2X₂, 6.9 ± 1.1 (*n* = 4). *B*, Concentration-dependent potentiation of ATP-evoked inward currents by extracellular Zn²⁺ (1–100 μM) applied simultaneously with the agonist. Under these circumstances, biphasic currents were rarely seen, and the inhibitory action of Zn²⁺ was lost. EC₅₀ values (micromolar concentration) for Zn²⁺ potentiation of ATP responses were rP2X_{2/6}, 8.2 ± 0.5 versus rP2X₂, 11.7 ± 2.8 (*n* = 6). *C*, Concentration-dependent potentiation and inhibition of ATP-evoked inward currents by extracellular H⁺ ions (pH 7.0–5.5) at rP2X_{2/6} receptors. pK_a values (−log₁₀[H⁺] causing 50% potentiation) were rP2X_{2/6}, 7.04 ± 0.05 versus rP2X₂, 7.05 ± 0.05 (*n* = 4). *D*, Paired biphasic inward currents evoked by ATP (100 μM, at pH 7.5) at rP2X_{2/6} receptors with either Mg²⁺ or Ca²⁺ (1.8 mM) present in the bathing solution. Substitution of Ca²⁺ with Mg²⁺ resulted in a reduction of ATP potency [as shown for rP2X₂ receptors (King et al., 1997)] without significantly altering the appearance of biphasic currents. Filled arrows draw attention to transient component of ATP-evoked inward currents (*A*, *C*). Data are expressed as mean ± SEM for four to six cells per determination.

response in the same cell. However, the incidence and reproducibility of biphasic responses in each oocyte varied in an unpredictable manner. The incidence of biphasic currents was greater and reproducibility more consistent when extracellular Zn²⁺ was present (Fig. 5*A*) or extracellular pH was lowered (Fig. 5*C*). Biphasic currents have already been reported at homomeric

rP2X₄ receptors, at which time-dependent changes in channel permeability were observed and shifts in the reversal potential for ATP-evoked currents noted (Khakh et al., 1999; Virginio et al., 1999). The binary permeability properties of rP2X₄ receptors were seen only when extracellular Ca²⁺ levels were lowered or zero Ca²⁺ conditions imposed (Khakh et al., 1999). Therefore, we explored this possibility and found that biphasic responses at heteromeric rP2X_{2/6} receptors were not enhanced when Ca²⁺ was replaced with equimolar Mg²⁺ (Fig. 5D). Ca²⁺-independent binary permeability properties have been reported for homomeric rP2X₂ receptors, although the time- and concentration-dependent changes in permeability do not result in biphasic currents to ATP (Khakh et al., 1999; Virginio et al., 1999). Others have reported, however, that rP2X₂ receptor ion channels do not show significant changes in unitary conductance or reversal potential of whole-cell currents (Ding and Sachs, 1999b). This inconsistency with the P2X₂ receptor is reminiscent of the variability of agonist responses (monophasic and biphasic) at the heteromeric P2X_{2/6} receptor. Currently, there is no satisfactory explanation for biphasic ATP responses at heteromeric P2X_{2/6} receptors.

The potency of ATP was lower at heteromeric rP2X_{2/6} receptors than homomeric rP2X₂ receptors, regardless of the pH level studied (Table 1). Although ATP potency was decreased overall, the rank potency order for mononucleotidic agonists at the heteromeric receptor remained the same as at the rP2X₂ receptor, namely ATP = ATP γ S > 2-MeSATP > ATP α S > BzATP. One significant difference in agonist activity involved the dinucleotide diadenosine tetraphosphate (Ap₄A), which is a full and potent agonist at rP2X₂ receptors (Pintor et al., 1996; Wildman et al., 1999a) and only a weak agonist at rP2X_{2/6} receptors. This difference in Ap₄A activity is potentially important, because this dinucleotide occurs naturally and is released in a Ca²⁺-dependent manner from central synaptosomes in rat brain (Pintor et al., 1992). Therefore, Ap₄A may subserve a transmitter role at homomeric rP2X₂ receptors but not at heteromeric rP2X_{2/6} receptors.

Extracellular pH is known to exert a profound effect on ATP potency at homomeric rP2X₂ receptors (King et al., 1996c, 1997; Stoop et al., 1997; Wildman et al., 1997, 1998, 1999b,c; Stoop and Quayle, 1998; Ding and Sachs, 1999a). A secondary inhibitory effect is observed at very low pH levels (e.g., pH 4.2), at which ATP responses rapidly desensitize, yet recover quickly, if pH_e is reversed to levels above pH 5.0, a phenomenon called “fade and rebound” (Stoop and Quayle, 1998). The heteromeric rP2X_{2/6} receptor showed both the potentiating and inhibitory effects of extracellular H⁺, and the pH ranges for these two separate effects are compressed when compared with rP2X₂ receptors. The inhibitory effect was caused by a reduction in agonist efficacy alone and not a decrease in agonist potency, as evidenced by the lower maximum for the ATP C–R curve at pH 5.5 (Fig. 2C). Because some homomeric P2X receptors (rP2X₁, rP2X₃, rP2X₄, and rP2X₇) also show a reduction in ATP activity when pH_e is lowered (Virginio et al., 1997; Wildman et al., 1999c), it is conceivable that the observed H⁺ inhibitory effect at rP2X_{2/6} receptors is caused as much by an action of H⁺ ions at the rP2X₆ subunit as at the rP2X₂ subunit.

The potency of suramin is progressively enhanced at rP2X₂ receptors as pH is lowered, with blockade occurring at nanomolar concentrations at pH 5.5 (King et al., 1997). The present results now show that this is an attribute shared by heteromeric rP2X_{2/6} receptors, although subtle differences were observed at pH 6.5 for

suramin blockade of homomeric rP2X₂ and heteromeric rP2X_{2/6} receptors. There appeared to be high-affinity (I₁) and low-affinity (I₂) sites for suramin at rP2X_{2/6} receptors, and activity indices for each component failed to match the corresponding IC₅₀ value at rP2X₂ receptors. The precise cause of this unusual effect is as yet unresolved. However, one possibility may involve differences in the subunit composition of heteromeric P2X_{2/6} receptors, if subpopulations of oligomeric assemblies containing different numbers of rP2X₆ subunits were generated. Where shown to be functional, the homomeric rP2X₆ receptor (or even the heteromeric rP2X_{4/6} receptor) has been reported to be relatively insensitive to suramin blockade (Collo et al., 1996; Lê et al., 1998). The suramin insensitivity of the P2X₆ subunit might help contribute to biphasic inhibition curves seen at pH 6.5 with the heteromeric P2X_{2/6} receptor.

The potentiating effect of extracellular Zn²⁺ was not significantly different at rP2X₂ and rP2X_{2/6} receptors. However, one subtle difference was noted when using high concentrations ($\geq 100 \mu\text{M}$) of this transition metal, which appeared to directly activate the heteromeric rP2X_{2/6} receptors without the need for exogenous ATP (data not shown). It is known that *Xenopus* oocytes continuously extrude small amounts of intracellular ATP via a mechanogated transport pathway (Nakamura and Strittmatter, 1996), and consequently the potency of locally released ATP may be sufficiently elevated by Zn²⁺ ions to explain the apparent Zn²⁺-activated inward currents. The subsequent inhibition of ATP responses by high concentrations of Zn²⁺ ions may be caused by a gradual desensitization of the receptor pool by locally released ATP.

In conclusion, the heteromeric P2X_{2/6} receptor possesses a significantly different operational profile from the wild-type P2X₂ receptor. It is of interest to us that rP2X₂ and rP2X₆ transcripts are found in rat brainstem (Collo et al., 1996; Comer et al., 1997) in nuclei with demonstrable pH-dependent chemoreceptive inputs (Thomas et al., 1999). The pH modulation of the homomeric P2X₂ and heteromeric rP2X_{2/6} receptor forms an interesting basis for examining the recently discovered involvement of ATP receptors in the CO₂-evoked (and pH-dependent) changes in central respiratory drive in rat (Thomas et al., 1999; Thomas and Spyer, 2000). At this point in time, however, the present results establish the P2X_{2/6} receptor as the fourth example of a heteropolymeric ATP-gated ion channel, which in this case possesses a pattern of pH modulation of ATP responses distinct from other known homomeric and heteromeric P2X receptors.

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