We report a protein-observe $^{19}$F NMR-based ligand titration binding study of human PDI b’x with Δ-somatostatin that also emphasises the need to optimise recombinant protein fluorination when using 5- or 6-fluoroindole. This study highlights a recombinant preference for 5-fluoroindole over 6-fluoroindole; most likely due to the influence of fluorene atomic packing within the folded protein structure. Fluorination affords a single $^{19}$F resonance probe to follow displacement of the protein x-linker as ligand is titrated and provides a dissociation constant of 23 ± 4 μM.

Protein disulphide isomerase (PDI) is a key enzyme responsible for the formation of native disulphide bonds in proteins that enter the secretory pathway of eukaryotic cells. PDI is a multifunctional protein able to catalyse the oxidation and isomerisation of disulphide bonds, as well as to bind to unfolded proteins and act as a molecular chaperone. The isomerisation of incorrectly paired cysteine residues is often a rate-limiting step on the folding pathway of disulphide bond-containing proteins both in vitro and in vivo. The ability of PDI to combine redox and molecular chaperone-like activities allows it to bind to partly structured folding intermediates and to catalyse simultaneously protein folding and associated native disulphide bond formation. PDI is the archetype of a large family of ER-resident PDI-like proteins.

PDI contains four thioredoxin-like domains, two of which – like thioredoxin itself – have redox-active catalytic sites (a and a’) and two of which do not (b and b’). The domain order is a-b-b’-x-a’-c, where x is a 19 residue linker between the b’ and a’ domains and c is a C-terminal acidic tail containing the KDEL ER-retention signal. The a and a’ domains are responsible for the redox activity of PDI while the b’ domain has been shown to be essential for ligand binding. The b’ domain binds both small and large peptide ligands, although large ligands also require the a and a’ domains to contribute to the overall binding interaction. The b’ domain has been shown to contribute to the substrate specificity of PDI and is required for disulphide isomerisation reactions in protein substrates.

The nature of substrate binding involving the b’ domain continues to be a subject of interest and any information regarding the role of b’ and x is extremely useful. Recent studies have highlighted the b’ domain of human PDI (hPDI) to be structurally connected and influenced by x, the linker that connects the b’ and a’ domains. The b’x construct has been shown to be receptor to ligand binding and has also been shown to moderate homodimerisation in b’x and bb’.

Data supported the previously published crystal structure studies of a b’x mutant that confirms b’ contains the thioredoxin-fold with x occluding the ligand binding site as mapped using NMR chemical shifts. As a result, it is thought that b’x exists in two conformational states with the x-linker ‘capping’ or ‘uncapping’ the ligand binding site and that ‘uncapping’ provided opportunity for homodimerisation. Limited proteolysis, using wild-type and mutant proteins to promote or retard ‘capping’, confirmed the x-linker operates similarly in full-length hPDI. Homodimerisation in hPDI is a mechanism by which the binding site is hidden from substrates, and has been proposed as a potential regulation mechanism for the protein in vivo.

hPDI bb’x and bb’ have been studied using $^1$H, $^{15}$N-HSQC chemical shift mapping to provide estimated dissociation constants ($K_d$) with peptide ligands Δ-somatostatin (AGSKNFWKFTFS) and mastoparan (INLKALAALAKKIL) with these studies approximating the $K_d$ in the range of 0.1–1.0 mM. Despite these investigations, the role of the x-linker remains a point of conjecture because backbone resonances in the linker itself were notoriously difficult to observe and track in the $^1$H, $^{15}$N-HSQC ligand binding studies mentioned above due to chemical shift line broadening. Therefore, we present this concise study of x within b’x using $^{19}$F NMR to follow the displacement of the ‘capped’ form of the linker when a known peptide ligand is titrated against constant
protein concentration. Switching to $^{19}$F NMR provides an optimum detection window of the x-linker to observe both ligand binding and displacement of the linker from the binding site. This is facilitated by the fact that hPDI b’x contains only a single tryptophan residue that resides at position 347 within the x-linker and positioned directly above the reported ligand binding site as shown in Fig. 1. In addition, we also describe the rationale behind our choice of indole fluorination that provides optimal expression.

All NMR datasets were acquired at 298 K using a Bruker Avance III 14.1 T (600 MHz $^1$H) NMR spectrometer equipped with a 5 mm QCI-F cryoprobe. Data were processed using Bruker Topspin 3.1 software and referenced using the position of the $^1$H$_2$O, relative gyromagnetic ratios (for $^{15}$N) or using trifluoroacetic acid (for $^{19}$F). Bacterial expression and purification of all proteins were identical to that reported previously for hPDI b’x with additional steps only to permit fluorination in accordance with the recently published protocol by Crowley and co-workers. For convenience, the entire growth protocol and purification can be found in the ESI.† The method utilises a standard bacteria minimal growth media with 60 mg L$^{-1}$ of indole, 5-fluoroindole or 6-fluoroindole added 15 minutes prior to induction with IPTG (isopropyl β-D-1-thiogalactopyranoside). All indoles were added to the growth media from a 200× dimethylsulfoxide (DMSO) stock solutions. An SDS-PAGE gel illustrating the effect of each indole on expression of hPDI b’x is shown in Fig. 2.

The inclusion of indole using DMSO as a stock carrier liquid did not hinder induction of the protein and the addition of 5-fluoroindole did also not alter the protein yield. From a minimal growth medium both indole and 5-fluoroindole provided yields of 6–8 mg L$^{-1}$ of purified protein. However, Fig. 2 highlights the effect of using 6-fluoroindole where the yield of recombinant b’x was found to be consistently lower than for indole and 5-fluoroindole with an estimated yield of ca. 0.5 mg L$^{-1}$.

Differences in fluoroindole overexpression are likely be related to the positioning of the fluorine in the indole ring and the SDS-PAGE gel in Fig. 2 highlights the importance of utilising fluorination that supports correct folding of a stable protein. This hypothesis was further corroborated using mass spectrometry (Bruker MicroTOF-Q) where 5-F-Trp incorporation was found to be >80% but 6-F-Trp incorporation was ca. 55% (see ESI Fig. 2†) confirming that 6-F-Trp incorporation was less successful. In an attempt to understand the effect of fluorination, Fig. 3 uses the ‘capped’ b’x I272A structure with in silico modified tryptophans to highlight the structural effects of 5-fluoro- (5-F-Trp: Fig. 3a) or 6-fluorotryptophan (6-F-Trp: Fig. 3b) labelling.

Close inspection of these structures suggests that when fluorine is present as 6-F-Trp, the fluorine is packed against residues F223, I284, F287 and F288 (see ESI Fig. 3a†). These four residues were emphasised as providing key interactions between b’ and the x-linker in the crystal structure. In contrast, inspection of the structure where fluorine is present using 5-F-Trp confirms that this structure experiences fewer critical interactions because the fluorine atom resides in a pocket above the ligand-binding site (Fig. 3a and ESI Fig. 3b†). The structural stability difference expected for 5-F-Trp versus 6-F-Trp is a likely explanation for the difference in expression yields when using 5-fluoro- and 6-fluoroindole. It could be
suggested that 6-fluoroindole may have compromised cell metabolism resulting in the loss of production. However, observed growth rates were similar regardless of indole utilised and close inspection of post-expression gel bands in Fig. 2 between 20–100 kDa supports comparable background protein levels across all samples. This observation would support that 6-fluoroindole does not adversely affect tryptophan synthase because many of these background proteins must also require tryptophan. In addition, we can confirm from other studies that 6-fluoroindole can be used to successfully produce other proteins with 6-F-Trp (unpublished results). However, these data do highlight the need to carefully consider the choice of fluorination sites when producing labelled proteins. The structural integrity of wild-type b’x expressed in the presence and absence of 5-F-Trp was assessed using 1H, 15N-HSQC data obtained from samples grown in minimal media containing a single nitrogen source of 0.6 mg L⁻¹ U₁₅N ammonium sulphate. 1H, 15N-HSQC b’x and 5-F-Trp-b’x spectra are shown in Fig. 4. The spectra overlay extremely well and suggest both samples have the same fold and structural arrangement, which is confirmed via a minimal chemical shift map analysis of the data (ESI Fig. 4†). Furthermore, these data are also comparable to previously published 1H, 15N-HSQC spectra of wild-type and x-linker ‘capping’ mutants of b’x.4,11

The degree of fluorination was estimated by comparing the relative ratios of the indole NH peaks in the 1H, 15N-HSQC spectra of b’x and 5-F-Trp-b’x and was found to vary between 80–90% depending on the preparation. This is in good agreement with the mass spectrometry data shown in ESI Fig. 2.† 19F NMR spectra of 0.3 mM hPDI 5-F-Trp-b’x were obtained with increasing Δ-somatostatin peptide concentration and are shown in Fig. 5. Each 1D 19F NMR experiment was acquired over 1024 scans using a relaxation delay of 2 s and an acquisition time of 0.321 s. The data obtained in Fig. 5 demonstrates the utility of the QCI-F cryoprobe and the ability to create a dataset within half the time required to acquire a 1H, 15N-HSQC dataset. This is especially important because b’x is prone to dimerise over time and the monomeric state in the predominantly ‘capped’ x conformation is required for this measurement. It is for this reason that a spectrum of a 48-hour ‘aged’ b’x sample is also shown in Fig. 5 where both ‘capped’ and ‘uncapped’ forms can be distinguished by 19F NMR. Using a fresh, purified sample enabled the titration of Δ-somatostatin to be monitored through a single 19F resonance that reports on the x-linker status as the peak tracks from the ‘capped’ toward the ‘uncapped’ state. It is interesting to note the reduction in 19F resonance line width as ligand is added; this supports the displacement of the x-linker in b’x as ligand binds to the protein. Once the linker is displaced, it will display increased mobility compared to the bulk protein, which manifests as an increase in the 19F T₂ relaxation time and subsequent reduction in line width. This reduction in linewidth associated with x-linker displacement has also been observed in 1H, 15N HSQC spectra.4,11 The change in 19F chemical shift with ligand concentration from Fig. 5 can be plotted as shown in Fig. 6 and fitted using the well-documented equation below:14,15 where Δ is the observed change in chemical shift, Δ₀ is the maximum shift and [L] and [P] are ligand and protein concentration respectively.

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\Delta = \Delta_0 \left( \frac{K_d + [L] + [P]}{2[P]} \right) - \sqrt{\left( (K_d + [L] + [P])^2 - 4[P][L] \right) / 2[P]}
\]

Solving the equation to fit the curve in Fig. 6 produces a dissociation constant for hPDI 5-F-Trp-b’x with Δ-somatostatin of 23 ± 4 μM. This value is marginally lower than that obtained from earlier studies that report K_d is in the 1.0–0.1 mM range and suggests the affinity of 5-F-Trp-b’x for Δ-somatostatin is higher than for the wild-type b’x protein. This likely to be due to a small but significant destabilisation of ‘capped’ species caused by fluorination that promotes displacement of the x-linker and exposure of the binding site. Fluorine substitutions

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**Fig. 4** 1H, 15N-HSQC spectra of hPDI b’x (a) and 5-F-Trp-b’x (b) from 0.25 mM protein samples. Data were acquired for each spectrum as 2048 x 256 points over 45 minutes.

**Fig. 5** 19F NMR spectra of 0.3 mM 5-F-Trp-b’x with increasing concentration of Δ-somatostatin peptide (Δ-som). 19F NMR spectra of 48 h ‘aged’ 5-F-trp-b’x shows 19F resonances reporting on ‘capped’ and ‘uncapped’ x-linker and 5-fluoroindole are also shown.
in proteins and peptides have been known to stabilise and drive hydrophobic interactions and the ligand-binding process across many PDIs including hPDI b’x is considered as hydrophobically driven.

If fluorine mediated hydrophobicity increased the strength of binding of the x-linker region to b’ then one might expect ‘uncapping’ to be more unfavourable with the consequence that ligand affinity is reduced. This does not appear to be the case.

A third explanation could be that the ligand experiences favourable hydrophobic interactions with the ‘uncapped’ x-linker; this would manifest as increased affinity and a lower $K_d$. These observations suggest the smaller $K_d$ is driven by destabilisation of the ‘capped’ state and/or the ‘uncapped’ state experiences a hydrophobically driven binding event that is stabilised by fluorination. The latter hypothesis proposes a potential role for the x-linker in ligand binding and such a process has been suggested from analysis of recent crystal structures of yeast and hPDI as well as in silico modelling and SAXS analysis of PDI from Humicola insolens.

**Conclusions**

$^{19}$F labelling using 5-fluoroindole to produce 5-fluorotryptophan for protein NMR spectroscopy is straightforward and successful. In addition, 5-fluoro-1-tryptophan is approximately 50-times more expensive than 5-fluoroindole per gram, thus making fluorindole-based labelling economical. However, our experience draws attention to the care required to find the optimum fluorindole for the system to be studied. hPDI b’x was best served using 5-fluoroindole but it is worth noting that 4-fluoro-, 5-fluoro- and 6-fluoroindoles are all available from general chemical suppliers. As a result, we recommend small-scale expression tests with different fluorindoles to discover the optimum labelling strategy. Our results also suggest that interrogation of a known structure can help identify the optimum fluoroindole through analysis of close-quarter atomic interactions.

Ultimately, $^{19}$F NMR has provided a useful shift in the NMR timescales of detection that was a challenge regarding x-linker observation using $^1$H, $^{13}$N HSQCs. Fluorine NMR enables direct observation of the tryptophan side chain in b’x and the $^{19}$F chemical shift change can be used to study ligand binding and for identification of the ‘capped’ and ‘uncapped’ conformations of the protein. $^{19}$F NMR has also confirmed that the x-linker is displaced upon binding as the 5-F-Trp chemical shift tracks from the ‘capped’ to ‘uncapped’ state.

The data provides an excellent fit and a $K_d$ of 23 µM for $\Delta$-somatostatin binding to 5-F-b’x and this value is marginally lower than 0.1–1 mM reported for the wild-type protein. We suggest this was most likely due to destabilisation of ‘capped’ 5-F-b’x, stabilisation of protein–peptide complex or a combination of both factors. Ultimately, this approach can be expanded further to study multiple PDI domain constructs and other PDI family members to monitor structure-function of this isomerase and folding chaperone.

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