

Kent Academic Repository

Aykur, Mehmet, Malatyalı, Erdoğan, Demirel, Filiz, Cömert-Koçak, Burçak, Gentekaki, Eleni, Tsaousis, Anastasios D. and Dogruman-Al, Funda (2024) *Blastocystis:*A mysterious member of the gut microbiome. Microorganisms, 12 (3). ISSN 2076-2607.

Downloaded from

https://kar.kent.ac.uk/105216/ The University of Kent's Academic Repository KAR

The version of record is available from

https://doi.org/10.3390/microorganisms12030461

This document version

Publisher pdf

DOI for this version

Licence for this version

CC BY (Attribution)

Additional information

Versions of research works

Versions of Record

If this version is the version of record, it is the same as the published version available on the publisher's web site. Cite as the published version.

Author Accepted Manuscripts

If this document is identified as the Author Accepted Manuscript it is the version after peer review but before type setting, copy editing or publisher branding. Cite as Surname, Initial. (Year) 'Title of article'. To be published in *Title* of *Journal*, Volume and issue numbers [peer-reviewed accepted version]. Available at: DOI or URL (Accessed: date).

Enquiries

If you have questions about this document contact ResearchSupport@kent.ac.uk. Please include the URL of the record in KAR. If you believe that your, or a third party's rights have been compromised through this document please see our Take Down policy (available from https://www.kent.ac.uk/guides/kar-the-kent-academic-repository#policies).





Review

Blastocystis: A Mysterious Member of the Gut Microbiome

Mehmet Aykur ^{1,*}, Erdoğan Malatyalı ², Filiz Demirel ³, Burçak Cömert-Koçak ⁴, Eleni Gentekaki ⁵, Anastasios D. Tsaousis ⁶ and Funda Dogruman-Al ⁷

- Department of Parasitology, Faculty of Medicine, Tokat Gaziosmanpasa University, Tokat 60030, Türkiye
- Department of Parasitology, Faculty of Medicine, Aydin Adnan Menderes University, Aydin 09010, Türkiye; erdogan.malatyali@adu.edu.tr
- Department of Medical Microbiology, Ankara City Hospital, Health Science University, Ankara 06500, Türkiye; dr.filiz.demirel@gmail.com
- Department of Medical Microbiology, Karadeniz Ereğli State Hospital, Zonguldak 67300, Türkiye; cburcakt@yahoo.com
- Department of Veterinary Medicine, School of Veterinary Medicine, University of Nicosia, Nicosia 2414, Cyprus; gentekaki.e@unic.ac.cy
- Laboratory of Molecular and Evolutionary Parasitology, RAPID Group, School of Biosciences, University of Kent, Canterbury CT2 7NZ, UK; a.tsaousis@kent.ac.uk
- Division of Medical Parasitology, Department of Medical Microbiology, Faculty of Medicine, Gazi University, Ankara 06560, Türkiye; alfunda@gazi.edu.tr
- * Correspondence: mehmetaykur@gmail.com

Abstract: *Blastocystis* is the most common gastrointestinal protist found in humans and animals. Although the clinical significance of *Blastocystis* remains unclear, the organism is increasingly being viewed as a commensal member of the gut microbiome. However, its impact on the microbiome is still being debated. It is unclear whether *Blastocystis* promotes a healthy gut and microbiome directly or whether it is more likely to colonize and persist in a healthy gut environment. In healthy people, *Blastocystis* is frequently associated with increased bacterial diversity and significant differences in the gut microbiome. Based on current knowledge, it is not possible to determine whether differences in the gut microbiome are the cause or result of *Blastocystis* colonization. Although it is possible that some aspects of this eukaryote's role in the intestinal microbiome remain unknown and that its effects vary, possibly due to subtype and intra-subtype variations and immune modulation, more research is needed to characterize these mechanisms in greater detail. This review covers recent findings on the effects of *Blastocystis* in the gut microbiome and immune modulation, its impact on the microbiome in autoimmune diseases, whether *Blastocystis* has a role like bacteria in the gut–brain axis, and its relationship with probiotics.

Keywords: *Blastocystis*; gut microbiome; microbiome modulation; immune modulation; autoimmune disease; gut–brain axis; probiotics

1. Introduction

Blastocystis is one of the most common microbial eukaryotes in the gastrointestinal tracts of humans and animals. Based on small subunit ribosomal RNA (SSUrRNA), the genus is composed of many genetically distinct subtypes (STs) that most likely represent separate species. The current taxonomy of Blastocystis is as follows: the kingdom Sar, the phylum Stramenopiles, the class Bigyra, the order Opalinata, the family Blastocystidae and the genus Blastocystis; species are not applicable [1]. Stramenopiles comprise over 100,000 species distributed across 21 classes. The majority of described species are diatoms, followed by brown algae, chrysophytes, xanthophytes, and oomycetes. However, unlike most other members of Stramenopiles, Blastocystis is neither flagellated nor motile [2]. In this review, the term "Blastocystis colonization" is used to define both the natural and experimental infection of hosts. Blastocystis can grow abundantly in xenic media and can be easily



Citation: Aykur, M.; Malatyalı, E.; Demirel, F.; Cömert-Koçak, B.; Gentekaki, E.; Tsaousis, A.D.; Dogruman-Al, F. *Blastocystis*: A Mysterious Member of the Gut Microbiome. *Microorganisms* **2024**, 12, 461. https://doi.org/10.3390/ microorganisms12030461

Academic Editor: María Teresa Gómez-Muñoz

Received: 15 January 2024 Revised: 19 February 2024 Accepted: 22 February 2024 Published: 24 February 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

Microorganisms **2024**, 12, 461 2 of 19

isolated from fresh fecal samples. However, achieving an axenic culture of *Blastocystis* is a very challenging process [3]. *Blastocystis* is well adapted to the anoxic/microaerophilic gut environment and lacks typical eukaryotic features, including cytochrome-driven mitochondrial electron transport. The organism is usually defined as a strict or obligate anaerobe that encodes genes for oxygen stress, including an alternative oxidase [4] and an SUF mobilization system [5]. Metabolically, *Blastocystis* has a glycolytic pathway whose components are localized in both the cytosol and its mitochondria, and recently, a mitochondrial carrier capable of transporting glycolytic intermediates was discovered, thus bridging the two branches of glycolysis [6].

Blastocystis has a global distribution; however, higher frequencies have been reported in developing countries because of poor hygiene, animal handling, or the fecal contamination of food and water [7,8]. The range of genetic diversity in *Blastocystis* is considerably high, and recently, at least 42 STs were identified from various hosts, relying on small subunit ribosomal RNA gene (SSU rRNA) polymorphisms [9–11]. In fact, one of the most significant current discussions is the number of STs and the identification of novel subtypes [12]. The genome of *Blastocystis* ST7 was the first to be sequenced in 2011, with data from ST1, ST2, ST4, ST6, ST8, and ST9 becoming available later at various stages of annotation [13,14]. Despite sharing common core genes, some important features, including genome sizes, intron numbers, guanine–cytosine (GC) contents, and gene contents, vary among subtypes [15].

The role of *Blastocystis* in the development of gastrointestinal diseases has also been much disputed despite a considerable number of studies [16-18]. Blastocystis infection has been associated with non-specific gastrointestinal symptoms such as abdominal pain, diarrhea, nausea, vomiting, bloating, and anorexia, as well as less frequent dermatological complaints like urticaria and severe itching [2,19–21]. In vitro studies on *Blastocystis* pathogenesis demonstrated that it can attach to the intestinal mucosa, increase intestinal permeability by secreting cysteine proteases, degrade secretory immunoglobulin A (IgA), induce the secretion of inflammatory cytokines such as interleukin-8, and cause the apoptosis of host cells [1,22]. In general, the prevalence of Blastocystis has been reported to be higher in healthy populations compared to individuals with ulcerative colitis (UC) or irritable bowel syndrome (IBS) [23,24]. Blastocystis resides in the human intestine for a long period of time without causing any symptoms, encouraging the question of whether it should be considered a pathogen or a commensal microorganism [25,26]. Nonetheless, the eradication of *Blastocystis* is considered necessary in cases where it is the sole protist agent and the patient's complaints persist [27]. There are limited studies revealing information regarding the actual abundance of Blastocystis in the host. In a study by Poirier et al., the parasite density of Blastocystis-positive samples was evaluated using a qPCR. Blastocystis numbers in hosts were reported to vary between $<10^2$ and $>10^7$ Blastocystis/g fecal sample [28].

The gut microbiome refers to the collection of bacteria, viruses, archaea, and eukaryotes that colonize the gastrointestinal tract, primarily the large intestine. This highly dynamic and complex ecosystem plays a crucial role in maintaining human health and has various physiological functions. It is currently accepted that the human gut microbiome is first acquired and established before or during birth, with the mode of delivery, ethnicity, and host genetics playing roles in its composition [29,30]. In addition, various external factors such as diet, nutritional status, prenatal events, geographical location, antibiotic treatment, and age contribute to establishing the gut microbiome throughout human life [31–35]. The microbiome reaches a "balanced" state with high taxonomic microbial diversity and richness in the following years of life, forming a commensal relationship with the host [36]. The Human Microbiome Project (HMP) and the Metagenomics of the Human Intestinal Tract (MetaHIT) project, as well as the development of novel technologies such as 16S rRNA gene metabarcoding, have improved our understanding [37,38]. The study of the gut microbiome has become a major area of interest in various disciplines. These days, some define the microbiome as a novel multicellular "organ" which interacts closely with

Microorganisms **2024**, 12, 461 3 of 19

its host [39]. The gut microbiome has numerous important functions including digestion, nutrient production, immune system regulation, gut barrier function for pathogens, and the regulation of metabolic activities; therefore, maintaining a healthy and diverse gut microbiome is essential for overall well-being [40,41]. The term "dysbiosis" can be defined as a persistent imbalance in the gut microbial community and can lead to various chronic conditions. Integrative analyses of the gut microbiome in humans and laboratory animals have offered possible relationships with many chronic diseases such as autoimmune disorders, obesity, diabetes, IBS, metabolic syndrome, depression, and allergy [42–47].

Single-celled eukaryotes constitute an important and heterogeneous group within the human intestinal microbiota. A major discussion point revolves around the categorization of these species as pathogenic, commensal, beneficial, or opportunistic pathogens. The well-known gut-related protozoa in humans are Blastocystis, Dientamoeba fragilis, Giardia intestinalis, Entamoeba histolytica, and Cryptosporidium spp. Among these, the last three significantly contribute to acute gastroenteritis and diarrheal diseases on a global scale [48]. However, many intestinal protist species, such as Endolimax nana, Entamoeba polecki, Iodamoeba butschlii, and Chilomastix mesnili, are non-pathogenic and might even be beneficial inhabitants of the gut [49]. Presently, at least eight species of Entamoeba spp. (E. polecki, E. gingivalis, E. chattoni, E. histolytica, E. dispar, E. hartmanni, E. moshkovskii, and E. Bangladeshi) have been identified in human samples, while E. histolytica is the only species with well-established pathogenicity [50]. A metagenomic approach that included samples from different countries revealed higher frequencies of Entamoeba spp., Blastocystis, and some other protozoan genera in healthy individuals [51]. Most retrospective studies have reported conflicting results regarding the roles of D. fragilis and Blastocystis in the development of gastrointestinal diseases [52,53]. Recent investigations on the microbiota have provided novel approaches to understanding the pathogenicity of intestinal protozoa.

There is a growing body of literature that emphasizes the importance of *Blastocystis* in the human gut microbiome [54–57]. Metagenomic studies have shown an association with increased abundances of the phylum Bacillota (syn. Firmicutes) and the class Clostridiales in the gut microbiomes of *Blastocystis*-colonized individuals, as well as a decreased abundance of *Bacteroides* [58,59]. However, *Blastocystis* infection has been linked to gut microbiome imbalance in certain gastrointestinal diseases such as IBS–constipation and inflammatory bowel disease (IBD) [60–62]. In addition, few studies have investigated *Blastocystis* subtype and microbiome interactions [55,57]. In general, *Blastocystis* is a common eukaryote in the intestinal microbiome of healthy humans. Its presence is linked with the high diversity and richness of bacterial communities [57]. However, a systematic understanding of how *Blastocystis* affects the gut microbiome and vice versa is still lacking. The main subjects addressed in this review are *Blastocystis* and gut microbiome modulation, immune modulation, autoimmune diseases, and, finally, the gut–brain axis.

2. Blastocystis and the Gut Microbiome

2.1. The Effect of Blastocystis on Gut Microbiome Modulation

Blastocystis colonization is thought to be related to changes in the gut bacterial microbiome [63]. Recent studies indicate that Blastocystis infection may be associated with alterations in the abundances of both beneficial and harmful intestinal bacteria. Research on the relationship between asymptomatic Blastocystis infection and intestinal bacterial composition is ongoing, although this association still needs to be fully understood [57,64,65]. Behboud et al. have reported that the mean relative abundances of Bifidobacterium and Lactobacillus/Enterococcus (beneficial bacteria) groups and Peptostreptococcus productus and Escherichia coli (harmful bacteria) were upregulated significantly, while the relative abundances of Bacteroides fragilis (B. fragilis) and Enterococcus sp. were downregulated considerably in those with Blastocystis compared to a control group [64]. According to a study by Di Cristanziano et al., in patients with Blastocystis, there was a consistent presence of bacterial genera linked to healthy status, including Eubacterium rectale and Eubacterium

Microorganisms **2024**, 12, 461 4 of 19

coprostanoligenes groups, as well as *Roseburia* and *Succinivibrio*. Nevertheless, their relative abundances were consistently lower compared to the control group [66].

Many studies report that colonization with *Blastocystis* is associated with increased diversity of the human intestinal bacterial microbiota (Figure 1). For instance, Audebert et al. reported a higher abundance of Clostridia and a lower abundance of Enterobacteriaceae in the fecal microbiota of patients colonized with *Blastocystis*, concluding that *Blastocystis* colonization is generally associated with healthy intestinal microbiota [67]. In a study aiming to evaluate the effect of *Blastocystis* on gut microbiota in healthy children, the diversity of intestinal microbiota and the proportion of beneficial bacteria were found to be higher in children colonized with *Blastocystis* than in children not colonized with the organism [68]. These results follow those of Alzate et al., who also found that *Blastocystis* was associated with a significant increase in bacterial richness in children [69].

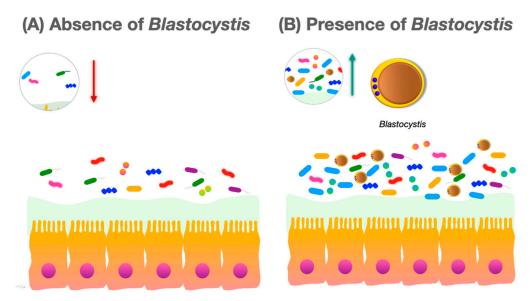


Figure 1. (**A**) In the absence of *Blastocystis*, the gut microbiota species richness and community evenness are lower; (**B**) in the presence of *Blastocystis*, the species richness and community evenness of the gut microbiota increases.

There are limited data in the literature on the relationships between *Blastocystis* STs and the gut microbiota. Blastocystis ST1 is one of the most commonly found STs in humans worldwide [70-72]. Some studies have demonstrated that ST1 has beneficial effects on the host gut microbiome and immune system. Deng et al. showed that colonization with Blastocystis ST1 could increase the levels of Alloprevotella and Akkermansia, which are beneficial bacteria for gut health, in a murine model [73]. Blastocystis ST3, another very common subtype, has been shown to cause an increase in beneficial bacteria such as Bacillota (syn. Firmicutes) and Bacteroidota (syn. Bacteroidetes) in the host gut microbiome, and it has been reported that this may indirectly be beneficial to the host immune response [74]. In a study examining the effect of Blastocystis ST3 colonization in a rat model, no significant influence on bacterial alpha diversity was observed before inducing colitis. However, after colitis induction, higher bacterial diversity was observed in rats with long-term Blastocystis ST3 colonization [63]. Recently, it was also reported that Blastocystis ST4 is beneficial for the gut as it increases the diversity of the gut microbiome [75,76]. Blastocystis ST4 has been found to promote the abundance of groups of bacteria belonging to Akkermansia spp., the family Lachnospiraceae, and the class Clostridia, all of which are considered beneficial to gut health, and to also inhibit the proliferation of Bacteroides spp., Escherichia spp., and Shigella spp. in the intestine, resulting in the alleviation of intestinal inflammation [75,76]. These results reflect those of Deng et al., who also found that colonization with Blastocystis ST4 may modify the intestinal microbiome and increase the accumulation of Th2 and Treg cells in the intestinal mucosa in a mouse model of induced colitis [75]. It has also

Microorganisms **2024**, 12, 461 5 of 19

been demonstrated that while Blastocystis ST4 colonization increases beneficial bacteria, it inhibits the proliferation of *Bacteroides vulgatus*, which is pathogenic for the intestine, when co-incubated with intestinal bacteria [77]. It has been observed that healthy individuals colonized with Blastocystis ST4 have high abundances of bacterial genera such as Sporolactobacillus and Candidatus Carsonella in their gut microbiome, while a reverse correlation was observed with Akkermansia [78,79]. Although ST7 is less common in humans worldwide than other STs of Blastocystis, an ST7 isolate has been reported to have pathogenic properties in in vitro and in vivo studies [62]. In addition, it has been reported that Blastocystis ST7 can disrupt the microbiotic balance in the gut microbiome population, especially by reducing Bifidobacteria longum (B. longum) and Lactobacillus brevis (L. brevis) [62]. Both L. brevis and B. longum have been found to benefit the gut microbiomes of IBS and IBD patients in addition to other beneficial gut microbiome species [80–82]. Even et al. verified that the colonization of Blastocystis has a major impact on the higher-level taxonomic diversity of the gut microbiota. The authors also found that the relative abundances of Ruminococcaceae and Clostridiales were higher in patients colonized with Blastocystis. Interestingly, it was shown that patients with multiple STs had a higher diversity of gut bacteria than those with just one ST [56].

It is considered that the composition of the gut microbiome plays a crucial role in the pathogenesis of certain diseases such as IBD, which is a chronic inflammation of the gastrointestinal tract. Although some studies have reported a high prevalence of *Blastocystis* in patients with IBD, the relationship between the organism and the disease is still controversial [83]. Deng et al. revealed that *Blastocystis* ST7 was associated with a decrease in beneficial bacteria such as *Bifidobacterium* and *Lactobacillus* and could lead to an increase in the severity of colitis in their murine model. They also reported that *Blastocystis* ST4 can decrease the severity of colitis by modulating the gut microbiome [83].

In contrast, Nagel et al. reported no association between *Blastocystis* and the gut microbiome in diarrhea-predominant IBS patients [84]. In another study focusing on cirrhotic patients with hepatic encephalopathy (HE), it was proposed that an inverse association existed between *Blastocystis* and HE severity. The authors reported that the alteration in bacterial diversity and the presence of *Blastocystis* may be significant factors in the pathophysiology of HE, highlighting a need for more research on this subject [85].

2.2. The Effect of Blastocystis on Immune Modulation

The gut microbiome plays an essential role in the health and disease status of the host. It is now known that it contributes significantly to the pathogenesis of autoimmune diseases, with the deterioration of the gut microbiome being linked to the dysregulation of the immune system [86]. The pathogenic potential of *Blastocystis*, its clinical significance, and its potential effects on the host immune system are still debated [57]. Whether *Blastocystis* is pathogenic or non-pathogenic depends on factors such as its interaction with the human gut microbiome, the subtype, and the human immune response regulators or modulators involved [75]. While *Blastocystis* colonizes the human gut and does not cause any infection, this situation can change in the event of a disruption to the immune system or gut microbiome balance [87]. An investigation of the metabolic profiles of *Blastocystis* carriers and non-carriers revealed differential levels of certain amino acids (Ala, Gly, His, Ile, Met, Thr, Try, and Val) in fecal samples collected from individuals from different countries. These amino acids are considered inflammatory biomarkers if their abundance in the gut is increased. The decrease in these amino acids in *Blastocystis*-positive individuals may indicate that the organism assumes an anti-inflammatory role in the intestine [88].

One of the areas of interest in *Blastocystis* research, along with recent work, is the insight into its potential to modulate the host immune system. Research suggests that *Blastocystis* infection can elicit various immune responses, including both pro-inflammatory and anti-inflammatory responses. However, the exact nature and significance of these immune responses are not fully understood [89]. Recent studies have revealed how some

Microorganisms **2024**, 12, 461 6 of 19

Blastocystis subtypes modulate the gut microbiome composition and how this change in the gut microbiome affects the immune response (Figure 2) [87,90,91].

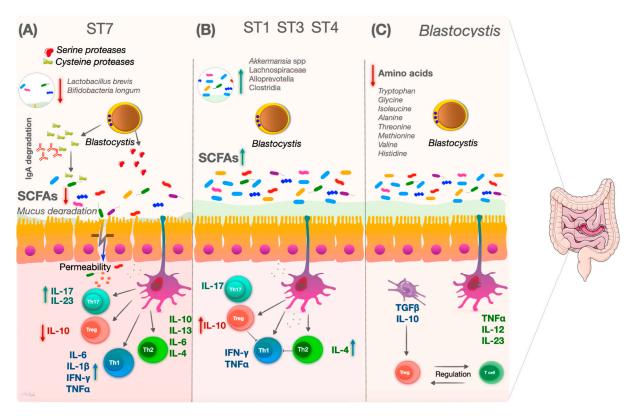


Figure 2. A general graphical overview of the impact of the gut microbiome on the immune response in *Blastocystis* infections and some of its subtypes. (**A**) The gut microbiome associated with *Blastocystis* ST7 can establish a pro-inflammatory environment by interacting with epithelial and dendritic cells (DC). (**B**) *Blastocystis* ST1, ST3, and ST4 increase the diversity of the gut microbiome and promote an anti-inflammatory state in the intestinal mucosa. (**C**) The decrease in some amino acids in the gut microbiome in the presence of *Blastocystis* may provide a balance in immune modulation.

The gut microbial ecosystem is crucial for the modulation and regulation of the immune system [92,93]. Mucin, a thick and sticky glycoprotein, is produced by goblet cells that secrete mucus in the body, especially in the gastrointestinal tract. Cysteine proteases, produced by pathogenic parasites, cause the breakdown of mucin, creating gaps between colon epithelial cells and thus making the invasion of the underlying host tissue possible [94]. The gut microbiome can support the development of T regulatory cells (Tregs) by producing short-chain fatty acids (SCFAs) and regulating Th2 immune responses during parasite infection. More specifically, *Blastocystis* ST4 has been associated with increased abundances of bacteria such as *Akkermansia* spp. and SCFA-producing bacteria associated with increased SCFA production, which can provide energy to goblet cells [49,95]. These data indicate subtype-specific effects of *Blastocystis* on immune modulation. Moreover, regarding host–microbiome interactions, the increase in diverse bacteria in the gut microbiota of *Blastocystis* ST1 and ST4 carriers causes SCFA production, which is important for the immune system overall and its modulation [73,96].

Immunoglobulin A is crucial in the mucosal defense of the gastrointestinal tract as it provides immune protection against microbial pathogens [97]. The release of cysteine protease by *Blastocystis* ST7 and aspartic proteases by ST4 has been shown to mediate the degradation of IgA and subsequently modulate the host immune response [87,98]. It has been demonstrated that *Blastocystis* ST4 cysteine proteases induce the upstream synthesis of interleukin (IL)-8 through the nuclear factor-κB (NF-κB) pathway [99]. An increase in the proinflammatory chemokine IL-8 and granulocyte–macrophage colony-

Microorganisms **2024**, 12, 461 7 of 19

stimulating factor (GM-CSF) in human colon carcinoma cells with a *Blastocystis* ST1 coculture has been reported [2]. In another study involving colonic epithelial cells, *Blastocystis* ST1 modulated the immune system by stimulating IL-8 release [100]. In the presence of colonization by *Blastocystis* ST7, a cascade of mitogen-activated protein kinases (MAPKs), an important signaling pathway in innate immunity, induced the upregulation of the pro-inflammatory cytokines IL-6, IL-1 β , and tumor necrosis factor- α (TNF α) in murine macrophages [57,101]. These cytokines play key roles in initiating and regulating the immune response. Additionally, *Blastocystis* has been found to activate immune cells, such as macrophages and dendritic cells, which are involved in the innate immune response [89].

Numerous studies in recent years have shown that Th1 and Th2 cells play distinct roles in the mediation of immunological responses [102]. Interferon-gamma (IFN- γ), IL-12, IL-2, and TNF- α are primarily secreted by Th1 cells, which additionally regulate cellular immunity. Th2 cells are essential for Th1 differentiation and the Th1 response. Th2 cells play a key role in mediating humoral immunity by primarily producing IL-4, IL-10, IL-13, and IL-6 [103]. Colonization with *Blastocystis* ST1, ST3, and ST4, the most common subtypes of *Blastocystis*, was associated with enhanced potent Th2 and Treg immune responses in a murine model of experimental colitis. Colonization with ST1, ST3, and ST4, has been reported in studies to have a beneficial effect on host health by regulating the gut microbiome composition and adaptive immune responses [63,73,75].

Deng et al. indicated that ST1 colonization could induce Th2 and Treg cell responses in normal, healthy mice [73]. Long-term *Blastocystis* ST3 colonization was reported to modify the appearance of induced colitis in a rat model of intestinal inflammation, whereas short-term colonization had no effect. In addition, it has been suggested that prolonged colonization with *Blastocystis* ST3 may be protective against intestinal inflammation by promoting faster recovery due to a significant decrease in inflammatory markers such as TNF α and IL-1 β [63]. Moreover, *Blastocystis* ST4 induces Th2 immune responses and increases the production of IL-4, IL-5, IL-13, and IL-10, thereby causing the suppression of inflammatory responses in colonic mucosal tissues [76].

The overall immune modulation effects of *Blastocystis* are complex and can vary depending on the specific subtype of the parasite, the host immune status, and other factors. When *Blastocystis*-positive and *Blastocystis*-negative patients with chronic urticaria were compared, patients infected with ST3 and ST2 had a higher abundance of IgE. However, there was no relationship between the *Blastocystis* STs of healthy individuals and patients with chronic urticaria [21]. Furthermore, the clinical implications of these immune responses are still not well understood.

It is worth noting that *Blastocystis* is often found in healthy individuals without any symptoms, suggesting that it may have a commensal symbiotic relationship with the host in certain cases [63,74,104]. However, more research is needed for a better understanding of the immunomodulatory effects of *Blastocystis* and their clinical significance in different individuals.

2.3. The Interaction of Blastocystis and the Gut Microbiome in Autoimmune Diseases

Autoimmune diseases (ADs) occur when cells of the immune system attack the host's cells and tissues, resulting in chronic inflammation. In the last decade, it has become known that environmental factors trigger ADs in genetically predisposed individuals [105]. The gut microbiome, which consists of trillions of microorganisms inhabiting the gastrointestinal tract, plays a critical role in regulating the immune system and maintaining gut health. It has been suggested that imbalances, either an increase or decrease in the specific taxa of the gut microbiome, may contribute to the development of ADs [106]. A disturbed balance in the gut microbiome may be associated not only with intestinal ADs (IBD, IBS, celiac disease, and autoimmune gastritis, etc.) but also with extra-intestinal ADs (multiple sclerosis, rheumatoid arthritis (RA), type 1 diabetes, and systemic lupus erythematosus (SLE)).

Microorganisms 2024, 12, 461 8 of 19

Few studies have investigated the relationship between ADs and the presence of *Blastocystis*. These include ADs such as SLE, RA, spondyloarthritis (SpA), IBD, UC, Crohn's disease (CD), and IBS [107–110]. However, the richness and diversity of the gut microbiome in association with *Blastocystis* and ADs have only been examined in SpA and IBS patients [111].

Spondyloarthritis comprises a group of rheumatic diseases with differential clinical features, such as ankylosing spondylitis (AS), reactive arthritis (ReA), and psoriatic arthritis (PsA), along with inflammatory bowel disease-associated SpA, uveitis, and dermatological and gastroenterological involvement [112]. A prevalent feature in many inflammatory diseases, including SpA, is gut microbial dysbiosis. Patients with SpA showed a decreased fecal abundance of Faecalibacterium prausnitzii and an increase in B. fragilis [113]. Regarding the gut microbiome composition of SpA patients, the main results from a meta-analysis showed increased frequencies of Bacteroidaceae and Enterobacteriaceae in the phylum Pseudomonadota (syn Proteobacteria), while the gut microbiome diversity in the phylum Bacteroidota (syn Bacteroidetes) showed decreases in Bacteroidales and Akkermansia [114]. There is a growing number of studies on the link between the gut microbiome and SpA, and several research investigations have demonstrated that the microbial profiles of SpA patients and healthy people differ [115–117]. Blastocystis-positive SpA patients showed significant increases in Pseudomonadota (syn Proteobacteria), the class Gammaproteobacteria, the family Succinivibrionaceae, and the genus Succinivibrio. However, in Blastocystis-negative SpA patients, there were significant increases in the Bacilli class, the order Lactobacillales, the Lactobacillaceae and Clostridiaceae families, and the genera Lactobacillus and Clostridium [111]. While Blastocystis-positive healthy individuals showed an increased diversity of the gut microbiome, no such increase was noted in the intestinal diversity of SpA patients [111]. These findings highlight the importance of Blastocystis as a typical component of a balanced gut microbiome [25,79].

Irritable bowel syndrome is a common functional gastrointestinal disorder characterized by abdominal pain, discomfort during defecation, and changes in the gut microbiome [118]. Some studies have reported that the gut microbiome of IBS patients had a significantly increased number of bacteria in the families Enterobacteriaceae and Bacteroides compared to healthy controls. Moreover, a significant increase in the family Lactobacillaceae in IBS patients has been reported [119,120]. A review of the relationship between IBS and the gut microbiome revealed that the genera *Faecalibacterium* and *Bifidobacterium* were significantly reduced in IBS patients [121]. A meta-analysis of 13 publications confirmed the lower abundance of *Bifidobacterium* in IBS patients, along with decreased *Lactobacillus* and *F. prausnitzii* [122]. In other studies, the proportion of *Bifidobacterium* in intestinal microbiota decreased in *Blastocystis*-positive individuals with IBS, while a decrease in *F. prausnitzii* in healthy *Blastocystis*-positive individuals was noted [60,123].

Recently, gut microbiome studies have shown greater abundances and higher gut richness of the Clostridia class, the families Ruminococcaceae and Prevotellaceae, and the *Faecalibacterium* and *Roseburia* genera in individual patients colonized with *Blastocystis* [57,67]. However, individuals not colonized with *Blastocystis* exhibited a higher abundance of Bacteroides [74]. Additionally, the increase in Bacteroides in the gut microbiome in people with various diseases, such as celiac disease and colorectal cancer, appears to be associated with low bacterial diversity [124–127]. These studies indicate that individuals colonized with *Blastocystis* have been associated with a richer and more diverse gut microbiome.

The interaction between *Blastocystis*, the gut microbiome, and ADs is a complex and evolving area of research. While some studies suggest possible connections, the mechanisms and clinical significance of these interactions remain unclear. Further research is needed to better understand the role of *Blastocystis* in ADs and its potential impact on gut health and the immune system. In individuals with ADs, the presence or absence of *Blastocystis* may cause changes in the balance of the gut microbiome. In addition, the different subtypes of *Blastocystis* should be considered along with the many factors that contribute to developing ADs.

Microorganisms **2024**, 12, 461 9 of 19

3. Blastocystis and the Gut-Brain Axis

The communication between the brain and the gut microbiome is bidirectional and is termed the "gut microbiome–brain axis". Communication along the gut–brain axis is mediated by various transmission systems, including the enteric nervous system, central nervous system, immune system, and endocrine system [128]. Maintaining a good balance between the gut microbiome and the brain is important for the host [80]. Various biochemical and metabolic processes must occur in order to maintain the gut–brain axis balance [129]. During these processes, signals in the gut microbiome modulate aspects of homeostasis through pathways of communication between the gut and the brain via the vagus nerve, metabolites such as SCFAs, the endocrine system, the immune system, and neurotransmitters such as serotonin, dopamine, acetylcholine, glutamate, γ -aminobutyric acid (GABA), and noradrenaline [130].

Although research has been conducted on the relationship between parasite manipulations and insect parasite interactions with the central nervous system (CNS), there have not been many studies on the interaction of the vertebrate host CNS and parasites [131,132]. The understanding of the interaction of the host CNS and parasites has increased recently with the development of the new and developing field of neuro-parasitology. Parasites can significantly affect the functioning of the host organism, including the immune response and the gut–brain axis, resulting in altered host behavior [133]. *Echinococcus granulosus*-derived ESPs (excretory–secretory products) affect cognitive function and the gut microbiome–brain axis as they have been demonstrated to alleviate dysbiosis and ameliorate cognitive decline in obese mice [134]. Another study revealed that *Hymenolepis diminuta* positively affected the spatial memory and new object recognition of the infected animal [131].

Despite the uncertainty surrounding the parasitic nature of *Blastocystis*, studies such as the above can shed light on the gut-brain axis relation to *Blastocystis* colonization/infection. There have been a limited number of studies showing the mechanisms through which the presence of *Blastocystis* in the intestine might influence the cognitive behavior of the host. In a study conducted by Defaye et al., a possible relationship between Blastocystis infection, colonic hypersensitivity, behavioral disorders, and gut microbiota changes using a rat model was investigated. In the study, animals infected with Blastocystis were associated with colonic hyperresponsiveness, anxiety, and depressive-like behavior [135]. In another study, the transplantation of human Blastocystis strains into mice resulted in changes in cognitive function and prefrontal cortex gene expression [136]. The relationship between Blastocystis ST1-4 and 7, the Bacillota/Bacteroidota (syn Firmicutes/Bacteroidetes) ratio (F/B ratio) of the fecal microbiota, and chronic stress was investigated in a population of Mexican university students. It was observed that colonization with ST4 was associated with a protective role in chronic stress [55]. Individuals colonized with *Blastocystis* ST4 have been associated with a richer and more diverse gut microbiome [79]. The relationship between Blastocystis subtypes, chronic stress, and IBS may need to be balanced by the CNS and gut microbiome [137]. A recent study suggests that Blastocystis may have the ability to influence the host's behavior and mood through the tryptophan synthesis pathway [138].

The gastrointestinal system is a complex and dynamic environment. *Blastocystis* exhibits broad genetic diversity, and the mechanisms and relationships between various subspecies and eubiosis/dysbiosis are being investigated [138–141]. Changes in gut microbiota species and critical metabolite levels in *Blastocystis*-colonized individuals may produce various potent signaling molecules in tryptophan metabolism [138,142]. These molecules may influence the gut microbiome–brain axis by altering tryptophan levels in gastrointestinal and neurological signaling pathways [138,143]. *Blastocystis* may also contribute to the balance of the bidirectional gut–brain axis (Figure 3). *Blastocystis* needs to be further considered as a new and mysterious actor in gut microbiome–brain axis research.

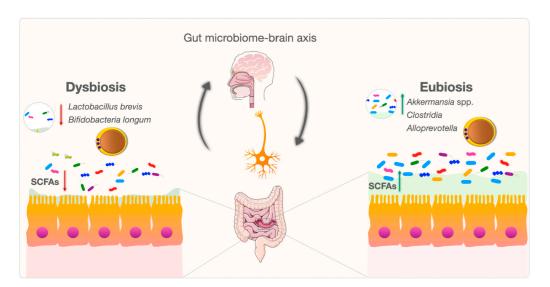


Figure 3. Bidirectional modulation and interaction of the gut microbiome–brain axis between *Blastocystis* and the gut microbiome.

4. Blastocystis and Probiotics

Probiotics are specific microorganisms that have beneficial effects on health. The most commonly used probiotics are specific strains from lactic acid bacterial species, especially *Lactobacillus* strains (*Streptococcus thermophilus*, *Lactococcus lactis*, *Enterococcus faecium*, and others) and *Bifidobacterium* strains and the yeast *Saccharomyces boulardii* (*S. boulardii*). Probiotics can modulate the microbiota and immune response of the host and inhibit the proliferation of parasites, leading to reduced parasitological loads and clinical improvement. Moreover, probiotics can increase the abundance of beneficial bacteria in the microbiota, change the environmental conditions to become less favorable for pathogens, compete with pathogens for nutrients and adhesion sites pathogens, negatively affect pathogens with their useful secretions (i.e., bacteriocins, lactic acid, hydrogen peroxide, etc.), inhibit bacterial toxins, increase mucus secretion, and induce mucosal immunity [144–151]. Although the relationship between probiotics and parasites has been investigated in various studies [144,148,152–158], there are very few reports related to *Blastocystis* and probiotics.

Blastocystis infections can occur in different forms ranging from asymptomatic to severe. Furthermore, the detection of *Blastocystis* in a stool sample does not necessarily mean that treatment is required. Its presence can be associated with infection or colonization whereby *Blastocystis* is a member of the healthy gut microbiome. This variability in outcomes could be due to different subtypes, the immunological response of the host, and gut microbial diversity [25,58,62,63,67,73,77,159]. If treatment is decided upon in the required symptomatic group (gastrointestinal symptoms; dermatological disorders involving acute/chronic urticaria and itching), the first choice is metronidazole. However, in some cases, failure to respond to metronidazole, the development of resistance, reinfection, or drug-related side effects make treatment difficult. Different antibiotics and combinations can be recommended for treatment (trimethoprim/sulfamethoxazole, paromomycin, secnidazole, tinidazole, and ornidazole), but in vitro sensitivity studies are very few, and studies are limited [19,27,160–164]. At this stage, probiotics may be recommended as a sole treatment option or as a support for treatment.

Dinleyici et al. compared therapies with *S. boulardii* and metronidazole in symptomatic children with a *Blastocystis* infection. They assessed clinical and parasitological cures in both study groups. While both metronidazole and *S. boulardii* demonstrated potential beneficial effects in treating *Blastocystis* infection, no statistically significant difference was found between the two treatment groups [165].

Angelici et al. documented a case report of a symptomatic *Blastocystis* infection caused by contaminated water. The patient had an intolerance to nitroimidazole derivatives, so

Microorganisms **2024**, 12, 461 11 of 19

metronidazole could not be used as the treating agent. Initially, a probiotic containing *Lactobacillus* and *Bifidobacterium* was used, but it did not solve the patient's problems. The administration of a different probiotic containing *S. boulardii* resulted in successful treatment [166].

Méabed et al. investigated the therapeutic effect of *S. boulardii* in experimental rats which were infected with the most common subtype of *Blastocystis* (ST3). The authors compared the results of parasitologic reduction, histopathological status, and the level of mRNA expression for the proinflammatory cytokines IL-6, IL-8, TNF- α , and inducible nitric oxide synthase (iNOS) on different therapy groups (*S. boulardii* (live), *S boulardii* (extract), metronidazole, co-therapy (metronidazole + *S. boulardii*), and a placebo). The co-therapy involving metronidazole and *S. boulardii* demonstrated a more favorable effect compared to the other treatments. The live *S. boulardii* had a significant beneficial effect on the local immune response of the colonic mucosa, such as goblet cell hyperplasia, as well as lower levels of proinflammatory cytokines and iNOS [167].

Lepczyńska et al. reported that the lactic acid-producing probiotic bacteria *Lactobacillus rhamnosus* and *Lactococcus lactis* significantly inhibited the growth of *Blastocystis* ST3 on xenic and axenic cultures. In the same study, they also investigated the relationship of *Enterococcus faecium* (which can also be used as a probiotic agent), *E. coli*, *Candida albicans*, and *Candida glabrata* with *Blastocystis* in vitro. Lactic acid-producing bacteria began inhibiting the growth of *Blastocystis* on the second day of the study. In contrast, co-culture with *E. coli* and *E. faecium* initially increased *Blastocystis* in the first two days but started to impede its growth after three days. By the fifth day, both *E. coli* and *E. faecium* demonstrated inhibitory effects on *Blastocystis* growth. The presence of *Candida* species had a limited and statistically insignificant effect on the growth of *Blastocystis*. However, it was indicated that *L. rhamnosus* and *L. lactis* may have the potential to be used as probiotics in *Blastocystis* prophylaxis or as a support for treatment [168].

The possible effects of probiotics on *Blastocystis* along with the type and dose of probiotic used for treatment remain unclear. In addition to the views that probiotics have treatment potential for *Blastocystis*, some studies argue that *Blastocystis* is a member of the healthy microbiota and that some *Blastocystis* subtypes may themselves be used as probiotics in the future [159]. This may also be an intriguing research subject. In the future, more successful results can be achieved with the use of probiotics designed by performing personalized microbiome analyses. Additional extensive studies are needed to achieve a comprehensive understanding.

5. Conclusions and Perspectives

Blastocystis is an important component and potential modulator of the human gut microbiome. This organism modulates the abundance of certain bacterial species and the Bacillota/Bacteroidota ratio. This overall review of recent data provides further support for the hypothesis that Blastocystis is a commensal eukaryote and may be an indicator of a healthy and balanced microbiome. However, these results may be somewhat limited by the study population and the methodology used to analyze the microbiome. Another finding that stands out from the earlier findings is that Blastocystis might have a subtype-dependent effect on the microbiota. An interesting focus in recent *Blastocystis* research is its potential to modulate the immune system. Blastocystis can trigger both pro- and anti-inflammatory cytokines in the host. On the other hand, Blastocystis may have immunomodulatory effects that could dampen the immune response. The metabolites and microbial changes could, in turn, potentially affect the production of neurotransmitters and other signaling molecules, thereby influencing brain function and behavior. While research on the relationship between asymptomatic/symptomatic Blastocystis infection and intestinal bacterial composition is ongoing, it still needs to be fully understood. However, there is an indication that Blastocystis infection may be associated with alterations in both beneficial and harmful intestinal bacteria. Further research on Blastocystis and the microbiome holds great

promise for unravelling the complex host–protist interactions, understanding their clinical significance, and developing novel therapeutic agents such as probiotics.

Author Contributions: Conceptualization, M.A., E.M. and F.D.-A., writing—original draft preparation, M.A., B.C.-K., F.D., E.M. and F.D.-A. writing—review and editing M.A., B.C.-K., F.D., E.M., E.G., A.D.T. and F.D.-A. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Data Availability Statement: Not applicable.

Acknowledgments: The authors acknowledge the COST Action CA21105 *Blastocystis* Under One Health supported by COST (European Cooperation in Science and Technology).

Conflicts of Interest: The authors declare no conflicts of interest.

References

- 1. Stensvold, C.R.; Tan, K.S.W.; Clark, C.G. Blastocystis. Trends Parasitol. 2020, 36, 315–316. [CrossRef]
- 2. Tan, K.S. New insights on classification, identification, and clinical relevance of *Blastocystis* spp. *Clin. Microbiol. Rev.* **2008**, 21, 639–665. [CrossRef]
- 3. Clark, C.G.; Stensvold, C.R. *Blastocystis*: Isolation, Xenic Cultivation, and Cryopreservation. *Curr. Protoc. Microbiol.* **2016**, 43, 20A.1.1–20A.1.8. [CrossRef]
- 4. Tsaousis, A.D.; Hamblin, K.A.; Elliott, C.R.; Young, L.; Rosell-Hidalgo, A.; Gourlay, C.W.; Moore, A.L.; Van der Giezen, M. The human gut colonizer *Blastocystis* respires using complex II and alternative oxidase to buffer transient oxygen fluctuations in the gut. *Front. Cell. Infect. Microbiol.* **2018**, *8*, 371. [CrossRef]
- 5. Tsaousis, A.D.; Ollagnier de Choudens, S.; Gentekaki, E.; Long, S.; Gaston, D.; Stechmann, A.; Vinella, D.; Py, B.; Fontecave, M.; Barras, F. Evolution of Fe/S cluster biogenesis in the anaerobic parasite *Blastocystis*. *Proc. Natl. Acad. Sci. USA* **2012**, 109, 10426–10431. [CrossRef] [PubMed]
- 6. Pyrihová, E.; King, M.S.; King, A.C.; Toleco, M.R.; van der Giezen, M.; Kunji, E.R. A mitochondrial carrier transports glycolytic intermediates to link cytosolic and mitochondrial glycolysis in the human gut parasite *Blastocystis*. *bioRxiv* 2023. [CrossRef]
- 7. Popruk, S.; Pintong, A.-R.; Radomyos, P. Diversity of Blastocystis subtypes in humans. J. Trop. Med. Parasitol. 2013, 36, 88–97.
- 8. El Safadi, D.; Cian, A.; Nourrisson, C.; Pereira, B.; Morelle, C.; Bastien, P.; Bellanger, A.-P.; Botterel, F.; Candolfi, E.; Desoubeaux, G. Prevalence, risk factors for infection and subtype distribution of the intestinal parasite *Blastocystis* sp. from a large-scale multi-center study in France. *BMC Infect. Dis.* **2016**, *16*, 451. [CrossRef] [PubMed]
- 9. Alfellani, M.A.; Jacob, A.S.; Perea, N.O.; Krecek, R.C.; Taner-Mulla, D.; Verweij, J.J.; Levecke, B.; Tannich, E.; Clark, C.G.; Stensvold, C.R. Diversity and distribution of *Blastocystis* sp. subtypes in non-human primates. *Parasitology* **2013**, *140*, 966–971. [CrossRef] [PubMed]
- 10. Santin, M.; Figueiredo, A.; Molokin, A.; George, N.S.; Köster, P.C.; Dashti, A.; González-Barrio, D.; Carmena, D.; Maloney, J.G. Division of *Blastocystis* ST10 into three new subtypes: ST42–ST44. *J. Eukaryot. Microbiol.* **2023**, *71*, e12998.
- 11. Koehler, A.V.; Herath, H.D.; Hall, R.S.; Wilcox, S.; Gasser, R.B. Marked genetic diversity within *Blastocystis* in Australian wildlife revealed using a next generation sequencing–phylogenetic approach. *Int. J. Parasitol. Parasites Wildl.* **2024**, 23, 100902. [CrossRef]
- 12. Stensvold, C.R.; Clark, C.G. Pre-empting Pandora's Box: *Blastocystis* Subtypes Revisited. *Trends Parasitol.* **2020**, *36*, 229–232. [CrossRef]
- 13. Higuera, A.; Salas-Leiva, D.E.; Curtis, B.; Patiño, L.H.; Zhao, D.; Jerlström-Hultqvist, J.; Dlutek, M.; Muñoz, M.; Roger, A.J.; Ramírez, J.D. Draft genomes of *Blastocystis* subtypes from human samples of Colombia. *Parasites Vectors* **2023**, *16*, 52. [CrossRef]
- 14. Denoeud, F.; Roussel, M.; Noel, B.; Wawrzyniak, I.; Da Silva, C.; Diogon, M.; Viscogliosi, E.; Brochier-Armanet, C.; Couloux, A.; Poulain, J. Genome sequence of the stramenopile *Blastocystis*, a human anaerobic parasite. *Genome Biol.* **2011**, *12*, R29. [CrossRef] [PubMed]
- 15. Gentekaki, E.; Curtis, B.A.; Stairs, C.W.; Klimes, V.; Elias, M.; Salas-Leiva, D.E.; Herman, E.K.; Eme, L.; Arias, M.C.; Henrissat, B.; et al. Extreme genome diversity in the hyper-prevalent parasitic eukaryote *Blastocystis*. *PLoS Biol.* **2017**, *15*, e2003769. [CrossRef] [PubMed]
- Robles-Cabrera, M.X.; Maguina, J.L.; Gonzales-Huerta, L.; Panduro-Correa, V.; Damaso-Mata, B.; Pecho-Silva, S.; Navarro-Solsol, A.C.; Rabaan, A.A.; Rodriguez-Morales, A.J.; Arteaga-Livias, K. *Blastocystis* species and Gastrointestinal Symptoms in Peruvian Adults Attended in a Public Hospital. *Infect. Chemother.* 2021, 53, 374–380. [CrossRef] [PubMed]
- 17. Cekin, A.H.; Cekin, Y.; Adakan, Y.; Tasdemir, E.; Koclar, F.G.; Yolcular, B.O. Blastocystosis in patients with gastrointestinal symptoms: A case–control study. *BMC Gastroenterol.* **2012**, 12, 122. [CrossRef]
- 18. Coyle, C.M.; Varughese, J.; Weiss, L.M.; Tanowitz, H.B. *Blastocystis*: To treat or not to treat. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* **2012**, *54*, 105–110. [CrossRef] [PubMed]
- 19. Kurt, O.; Dogruman Al, F.; Tanyuksel, M. Eradication of *Blastocystis* in humans: Really necessary for all? *Parasitol. Int.* **2016**, 65, 797–801. [CrossRef] [PubMed]

20. Bahrami, F.; Babaei, E.; Badirzadeh, A.; Riabi, T.R.; Abdoli, A. *Blastocystis*, urticaria, and skin disorders: Review of the current evidences. *Eur. J. Clin. Microbiol. Infect. Dis.* **2019**, *39*, 1027–1042. [CrossRef] [PubMed]

- 21. Aykur, M.; Camyar, A.; Turk, B.G.; Sin, A.Z.; Dagci, H. Evaluation of association with subtypes and alleles of *Blastocystis* with chronic spontaneous urticaria. *Acta Trop.* **2022**, 231, 106455. [CrossRef]
- 22. Nourrisson, C.; Wawrzyniak, I.; Cian, A.; Livrelli, V.; Viscogliosi, E.; Delbac, F.; Poirier, P. On *Blastocystis* secreted cysteine proteases: A legumain-activated cathepsin B increases paracellular permeability of intestinal Caco-2 cell monolayers. *Parasitology* **2016**, *143*, 1713–1722. [CrossRef]
- 23. Krogsgaard, L.R.; Engsbro, A.L.; Stensvold, C.R.; Nielsen, H.V.; Bytzer, P. The prevalence of intestinal parasites is not greater among individuals with irritable bowel syndrome: A population-based case-control study. *Clin. Gastroenterol. Hepatol.* **2015**, *13*, 507–513.e2. [CrossRef] [PubMed]
- 24. Petersen, A.M.; Stensvold, C.R.; Mirsepasi, H.; Engberg, J.; Friis-Moller, A.; Porsbo, L.J.; Hammerum, A.M.; Nordgaard-Lassen, I.; Nielsen, H.V.; Krogfelt, K.A. Active ulcerative colitis associated with low prevalence of *Blastocystis* and Dientamoeba fragilis infection. *Scand. J. Gastroenterol.* **2013**, *48*, 638–639. [CrossRef]
- 25. Scanlan, P.D.; Stensvold, C.R.; Rajilic-Stojanovic, M.; Heilig, H.G.; De Vos, W.M.; O'Toole, P.W.; Cotter, P.D. The microbial eukaryote *Blastocystis* is a prevalent and diverse member of the healthy human gut microbiota. *FEMS Microbiol. Ecol.* **2014**, 90, 326–330. [CrossRef]
- 26. Parfrey, L.W.; Walters, W.A.; Lauber, C.L.; Clemente, J.C.; Berg-Lyons, D.; Teiling, C.; Kodira, C.; Mohiuddin, M.; Brunelle, J.; Driscoll, M.; et al. Communities of microbial eukaryotes in the mammalian gut within the context of environmental eukaryotic diversity. *Front. Microbiol.* **2014**, *5*, 298. [CrossRef] [PubMed]
- 27. Roberts, T.; Ellis, J.; Harkness, J.; Marriott, D.; Stark, D. Treatment failure in patients with chronic *Blastocystis* infection. *J. Med. Microbiol.* **2014**, *63*, 252–257. [CrossRef]
- 28. Poirier, P.; Wawrzyniak, I.; Albert, A.; El Alaoui, H.; Delbac, F.; Livrelli, V. Development and Evaluation of a Real-Time PCR Assay for Detection and Quantification of *Blastocystis* Parasites in Human Stool Samples: Prospective Study of Patients with Hematological Malignancies. *J. Clin. Microbiol.* **2011**, *49*, 975–983. [CrossRef] [PubMed]
- 29. Cahana, I.; Iraqi, F.A. Impact of host genetics on gut microbiome: Take-home lessons from human and mouse studies. *Anim. Model. Exp. Med.* **2020**, *3*, 229–236. [CrossRef]
- 30. Gupta, V.K.; Paul, S.; Dutta, C. Geography, ethnicity or subsistence-specific variations in human microbiome composition and diversity. *Front. Microbiol.* **2017**, *8*, 1162. [CrossRef]
- 31. Kau, A.L.; Ahern, P.P.; Griffin, N.W.; Goodman, A.L.; Gordon, J.I. Human nutrition, the gut microbiome and the immune system. *Nature* **2011**, 474, 327–336. [CrossRef] [PubMed]
- 32. Yatsunenko, T.; Rey, F.E.; Manary, M.J.; Trehan, I.; Dominguez-Bello, M.G.; Contreras, M.; Magris, M.; Hidalgo, G.; Baldassano, R.N.; Anokhin, A.P.; et al. Human gut microbiome viewed across age and geography. *Nature* **2012**, *486*, 222–227. [CrossRef] [PubMed]
- 33. Thursby, E.; Juge, N. Introduction to the human gut microbiota. Biochem. J. 2017, 474, 1823–1836. [CrossRef] [PubMed]
- 34. Hall, A.B.; Tolonen, A.C.; Xavier, R.J. Human genetic variation and the gut microbiome in disease. *Nat. Rev. Genet.* **2017**, *18*, 690–699. [CrossRef] [PubMed]
- 35. Deidda, G.; Biazzo, M. Gut and Brain: Investigating Physiological and Pathological Interactions between Microbiota and Brain to Gain New Therapeutic Avenues for Brain Diseases. *Front. Neurosci.* **2021**, *15*, 753915. [CrossRef] [PubMed]
- 36. Vemuri, R.; Gundamaraju, R.; Shastri, M.D.; Shukla, S.D.; Kalpurath, K.; Ball, M.; Tristram, S.; Shankar, E.M.; Ahuja, K.; Eri, R. Gut Microbial Changes, Interactions, and Their Implications on Human Lifecycle: An Ageing Perspective. *BioMed Res. Int.* 2018, 2018, 4178607. [CrossRef]
- 37. Gevers, D.; Knight, R.; Petrosino, J.F.; Huang, K.; McGuire, A.L.; Birren, B.W.; Nelson, K.E.; White, O.; Methe, B.A.; Huttenhower, C. The Human Microbiome Project: A community resource for the healthy human microbiome. *PLoS Biol.* **2012**, *10*, e1001377. [CrossRef]
- 38. Ehrlich, S.D.; Consortium, M. MetaHIT: The European Union Project on Metagenomics of the Human Intestinal Tract. In *Metagenomics of the Human Body*; Springer: New York, NY, USA, 2011; pp. 307–316.
- 39. Clarke, S.F.; Murphy, E.F.; O'Sullivan, O.; Lucey, A.J.; Humphreys, M.; Hogan, A.; Hayes, P.; O'Reilly, M.; Jeffery, I.B.; Wood-Martin, R. Exercise and associated dietary extremes impact on gut microbial diversity. *Gut* 2014, 63, 1913–1920. [CrossRef]
- 40. Jandhyala, S.M.; Talukdar, R.; Subramanyam, C.; Vuyyuru, H.; Sasikala, M.; Nageshwar Reddy, D. Role of the normal gut microbiota. *World J. Gastroenterol.* **2015**, *21*, 8787–8803. [CrossRef]
- 41. Zheng, D.; Liwinski, T.; Elinav, E. Interaction between microbiota and immunity in health and disease. *Cell Res.* **2020**, *30*, 492–506. [CrossRef]
- 42. Hills, R.D., Jr.; Pontefract, B.A.; Mishcon, H.R.; Black, C.A.; Sutton, S.C.; Theberge, C.R. Gut Microbiome: Profound Implications for Diet and Disease. *Nutrients* **2019**, *11*, 1613. [CrossRef]
- 43. Zubeldia-Varela, E.; Barker-Tejeda, T.C.; Obeso, D.; Villasenor, A.; Barber, D.; Perez-Gordo, M. Microbiome and Allergy: New Insights and Perspectives. *J. Investig. Allergol. Clin. Immunol.* **2022**, 32, 327–344. [CrossRef] [PubMed]
- 44. Gülden, E.; Wong, F.S.; Wen, L. The gut microbiota and type 1 diabetes. Clin. Immunol. 2015, 159, 143–153. [CrossRef] [PubMed]

45. Jalanka-Tuovinen, J.; Salojärvi, J.; Salonen, A.; Immonen, O.; Garsed, K.; Kelly, F.M.; Zaitoun, A.; Palva, A.; Spiller, R.C.; De Vos, W.M. Faecal microbiota composition and host–microbe cross-talk following gastroenteritis and in postinfectious irritable bowel syndrome. *Gut* 2014, 63, 1737–1745. [CrossRef] [PubMed]

- 46. Perry, R.J.; Peng, L.; Barry, N.A.; Cline, G.W.; Zhang, D.; Cardone, R.L.; Petersen, K.F.; Kibbey, R.G.; Goodman, A.L.; Shulman, G.I. Acetate mediates a microbiome–brain–β-cell axis to promote metabolic syndrome. *Nature* **2016**, *534*, 213–217. [CrossRef] [PubMed]
- 47. Vijay, A.; Valdes, A.M. Role of the gut microbiome in chronic diseases: A narrative review. *Eur. J. Clin. Nutr.* **2022**, *76*, 489–501. [CrossRef] [PubMed]
- 48. Thompson, R.; Ash, A. Molecular epidemiology of Giardia and Cryptosporidium infections. *Infect. Genet. Evol.* **2016**, *40*, 315–323. [CrossRef] [PubMed]
- 49. Dubik, M.; Pilecki, B.; Moeller, J.B. Commensal Intestinal Protozoa—Underestimated Members of the Gut Microbial Community. *Biology* 2022, 11, 1742. [CrossRef] [PubMed]
- 50. Hooshyar, H.; Rostamkhani, P.; Rezaeian, M. An Annotated checklist of the human and animal entamoeba (Amoebida: Endamoebidae) species—A review article. *Iran. J. Parasitol.* **2015**, *10*, 146.
- 51. Lokmer, A.; Cian, A.; Froment, A.; Gantois, N.; Viscogliosi, E.; Chabé, M.; Ségurel, L. Use of shotgun metagenomics for the identification of protozoa in the gut microbiota of healthy individuals from worldwide populations with various industrialization levels. *PLoS ONE* **2019**, *14*, e0211139. [CrossRef]
- 52. Garcia, L.S. Dientamoeba fragilis, One of the Neglected Intestinal Protozoa. J. Clin. Microbiol. 2016, 54, 2243–2250. [CrossRef]
- 53. Shasha, D.; Grupel, D.; Treigerman, O.; Prajgrod, G.; Paran, Y.; Hacham, D.; Ben-Ami, R.; Albukrek, D.; Zacay, G. The clinical significance of Dientamoeba fragilis and *Blastocystis* in human stool—Retrospective cohort study. *Clin. Microbiol. Infect.* **2024**, *30*, 130–136. [CrossRef]
- 54. Olyaiee, A.; Sadeghi, A.; Yadegar, A.; Mirsamadi, E.S.; Mirjalali, H. Gut Microbiota Shifting in Irritable Bowel Syndrome: The Mysterious Role of *Blastocystis* sp. *Front. Med.* **2022**, *9*, 890127. [CrossRef] [PubMed]
- 55. Guangorena-Gomez, J.O.; Lozano, O., II; Rivera-Medina, I.L.; Mendez-Hernandez, A.; Espinosa-Fematt, J.A.; Munoz-Yanez, C. Relationship among *Blastocystis*, the Firmicutes/Bacteroidetes Ratio and Chronic Stress in Mexican University Students. *Curr. Microbiol.* 2022, 79, 72. [CrossRef]
- 56. Even, G.; Lokmer, A.; Rodrigues, J.; Audebert, C.; Viscogliosi, E.; Segurel, L.; Chabe, M. Changes in the Human Gut Microbiota Associated with Colonization by *Blastocystis* sp. and *Entamoeba* spp. in Non-Industrialized Populations. *Front. Cell. Infect. Microbiol.* 2021, 11, 533528. [CrossRef] [PubMed]
- 57. Deng, L.; Wojciech, L.; Gascoigne, N.R.J.; Peng, G.; Tan, K.S.W. New insights into the interactions between *Blastocystis*, the gut microbiota, and host immunity. *PLoS Pathog.* **2021**, *17*, e1009253. [CrossRef]
- 58. Beghini, F.; Pasolli, E.; Truong, T.D.; Putignani, L.; Caccio, S.M.; Segata, N. Large-scale comparative metagenomics of *Blastocystis*, a common member of the human gut microbiome. *ISME J.* **2017**, *11*, 2848–2863. [CrossRef] [PubMed]
- 59. Yanez, C.M.; Hernandez, A.M.; Sandoval, A.M.; Dominguez, M.A.M.; Muniz, S.A.Z.; Gomez, J.O.G. Prevalence of *Blastocystis* and its association with Firmicutes/Bacteroidetes ratio in clinically healthy and metabolically ill subjects. *BMC Microbiol.* **2021**, 21, 339. [CrossRef]
- 60. Nourrisson, C.; Scanzi, J.; Pereira, B.; NkoudMongo, C.; Wawrzyniak, I.; Cian, A.; Viscogliosi, E.; Livrelli, V.; Delbac, F.; Dapoigny, M.; et al. *Blastocystis* is associated with decrease of fecal microbiota protective bacteria: Comparative analysis between patients with irritable bowel syndrome and control subjects. *PLoS ONE* **2014**, *9*, e111868. [CrossRef]
- 61. Shirvani, G.; Fasihi-Harandi, M.; Raiesi, O.; Bazargan, N.; Zahedi, M.J.; Sharifi, I.; Kalantari-Khandani, B.; Nooshadokht, M.; Shabandoust, H.; Mohammadi, M.A.; et al. Prevalence and Molecular Subtyping of *Blastocystis* from Patients with Irritable Bowel Syndrome, Inflammatory Bowel Disease and Chronic Urticaria in Iran. *Acta Parasitol.* **2020**, *65*, 90–96. [CrossRef]
- 62. Yason, J.A.; Liang, Y.R.; Png, C.W.; Zhang, Y.; Tan, K.S.W. Interactions between a pathogenic *Blastocystis* subtype and gut microbiota: In vitro and in vivo studies. *Microbiome* **2019**, 7, 30. [CrossRef]
- 63. Billy, V.; Lhotska, Z.; Jirku, M.; Kadlecova, O.; Frgelecova, L.; Parfrey, L.W.; Pomajbikova, K.J. *Blastocystis* Colonization Alters the Gut Microbiome and, in Some Cases, Promotes Faster Recovery from Induced Colitis. *Front. Microbiol.* **2021**, *12*, 641483. [CrossRef]
- 64. Behboud, S.; Solhjoo, K.; Erfanian, S.; Pirestani, M.; Abdoli, A. Alteration of gut bacteria composition among individuals with asymptomatic *Blastocystis* infection: A case-control study. *Microb. Pathog.* **2022**, *169*, 105639. [CrossRef]
- 65. Nieves-Ramirez, M.E.; Partida-Rodriguez, O.; Laforest-Lapointe, I.; Reynolds, L.A.; Brown, E.M.; Valdez-Salazar, A.; Moran-Silva, P.; Rojas-Velazquez, L.; Morien, E.; Parfrey, L.W.; et al. Asymptomatic Intestinal Colonization with Protist *Blastocystis* Is Strongly Associated with Distinct Microbiome Ecological Patterns. *mSystems* **2018**, 3. [CrossRef] [PubMed]
- 66. Di Cristanziano, V.; Farowski, F.; Berrilli, F.; Santoro, M.; Di Cave, D.; Gle, C.; Daeumer, M.; Thielen, A.; Wirtz, M.; Kaiser, R.; et al. Analysis of Human Gut Microbiota Composition Associated to the Presence of Commensal and Pathogen Microorganisms in Cote d'Ivoire. *Microorganisms* 2021, 9, 1763. [CrossRef]
- 67. Audebert, C.; Even, G.; Cian, A.; The Blastocystis Investigation Group; Loywick, A.; Merlin, S.; Viscogliosi, E.; Chabe, M. Colonization with the enteric protozoa *Blastocystis* is associated with increased diversity of human gut bacterial microbiota. *Sci. Rep.* 2016, 6, 25255. [CrossRef] [PubMed]

68. Kodio, A.; Coulibaly, D.; Kone, A.K.; Konate, S.; Doumbo, S.; Guindo, A.; Bittar, F.; Gouriet, F.; Raoult, D.; Thera, M.A.; et al. *Blastocystis* Colonization Is Associated with Increased Diversity and Altered Gut Bacterial Communities in Healthy Malian Children. *Microorganisms* 2019, 7, 649. [CrossRef] [PubMed]

- 69. Alzate, J.F.; Toro-Londono, M.; Cabarcas, F.; Garcia-Montoya, G.; Galvan-Diaz, A. Contrasting microbiota profiles observed in children carrying either *Blastocystis* spp. or the commensal amoebas *Entamoeba coli* or *Endolimax nana*. *Sci. Rep.* **2020**, *10*, 15354. [CrossRef] [PubMed]
- 70. Asghari, A.; Hassanipour, S.; Hatam, G. Comparative molecular prevalence and subtypes distribution of *Blastocystis* sp. a potentially zoonotic infection isolated from symptomatic and asymptomatic patients in Iran: A systematic review and meta-analysis. *Acta Parasitol.* **2021**, *66*, 745–759. [CrossRef] [PubMed]
- 71. Ramirez, J.D.; Sanchez, A.; Hernandez, C.; Florez, C.; Bernal, M.C.; Giraldo, J.C.; Reyes, P.; Lopez, M.C.; Garcia, L.; Cooper, P.J.; et al. Geographic distribution of human *Blastocystis* subtypes in South America. *Infect. Genet. Evol. J. Mol. Epidemiol. Evol. Genet. Infect. Dis.* 2016, 41, 32–35. [CrossRef]
- 72. Yowang, A.; Tsaousis, A.D.; Chumphonsuk, T.; Thongsin, N.; Kullawong, N.; Popluechai, S.; Gentekaki, E. High diversity of *Blastocystis* subtypes isolated from asymptomatic adults living in Chiang Rai, Thailand. *Infect. Genet. Evol. J. Mol. Epidemiol. Evol. Genet. Infect. Dis.* 2018, 65, 270–275. [CrossRef] [PubMed]
- 73. Deng, L.; Wojciech, L.; Png, C.W.; Kioh, Y.Q.D.; Ng, G.C.; Chan, E.C.Y.; Zhang, Y.; Gascoigne, N.R.J.; Tan, K.S.W. Colonization with ubiquitous protist *Blastocystis* ST1 ameliorates DSS-induced colitis and promotes beneficial microbiota and immune outcomes. *NPJ Biofilms Microbiomes* **2023**, *9*, 22. [CrossRef] [PubMed]
- 74. Andersen, L.O.; Bonde, I.; Nielsen, H.B.; Stensvold, C.R. A retrospective metagenomics approach to studying *Blastocystis*. *FEMS Microbiol*. *Ecol.* **2015**, 91, fiv072. [CrossRef] [PubMed]
- 75. Deng, L.; Wojciech, L.; Png, C.W.; Koh, E.Y.; Aung, T.T.; Kioh, D.Y.Q.; Chan, E.C.Y.; Malleret, B.; Zhang, Y.; Peng, G.; et al. Experimental colonization with *Blastocystis* ST4 is associated with protective immune responses and modulation of gut microbiome in a DSS-induced colitis mouse model. *Cell. Mol. Life Sci.* 2022, 79, 245. [CrossRef] [PubMed]
- 76. Feranmi, F. Blastocystis subtype 4 linked to gut microbiota stability. Lancet Microbe 2022, 3, e564. [CrossRef] [PubMed]
- 77. Deng, L.; Tan, K.S.W. Interactions between *Blastocystis* subtype ST4 and gut microbiota in vitro. *Parasites Vectors* **2022**, *15*, 80. [CrossRef]
- 78. Forsell, J.; Bengtsson-Palme, J.; Angelin, M.; Johansson, A.; Evengard, B.; Granlund, M. The relation between *Blastocystis* and the intestinal microbiota in Swedish travellers. *BMC Microbiol*. **2017**, *17*, 231. [CrossRef]
- 79. Tito, R.Y.; Chaffron, S.; Caenepeel, C.; Lima-Mendez, G.; Wang, J.; Vieira-Silva, S.; Falony, G.; Hildebrand, F.; Darzi, Y.; Rymenans, L.; et al. Population-level analysis of *Blastocystis* subtype prevalence and variation in the human gut microbiota. *Gut* **2019**, *68*, 1180–1189. [CrossRef]
- 80. Hillestad, E.M.R.; van der Meeren, A.; Nagaraja, B.H.; Bjorsvik, B.R.; Haleem, N.; Benitez-Paez, A.; Sanz, Y.; Hausken, T.; Lied, G.A.; Lundervold, A.; et al. Gut bless you: The microbiota-gut-brain axis in irritable bowel syndrome. *World J. Gastroenterol.* **2022**, 28, 412–431. [CrossRef]
- 81. Sugahara, H.; Odamaki, T.; Fukuda, S.; Kato, T.; Xiao, J.Z.; Abe, F.; Kikuchi, J.; Ohno, H. Probiotic *Bifidobacterium longum* alters gut luminal metabolism through modification of the gut microbial community. *Sci. Rep.* **2015**, *5*, 13548. [CrossRef]
- 82. Walter, J. Ecological role of lactobacilli in the gastrointestinal tract: Implications for fundamental and biomedical research. *Appl. Environ. Microbiol.* **2008**, 74, 4985–4996. [CrossRef] [PubMed]
- 83. Deng, L.; Wojciech, L.; Png, C.W.; Kioh, D.Y.Q.; Gu, Y.; Aung, T.T.; Malleret, B.; Chan, E.C.Y.; Peng, G.; Zhang, Y. Colonization with two different *Blastocystis* subtypes in DSS-induced colitis mice is associated with strikingly different microbiome and pathological features. *Theranostics* **2023**, *13*, 1165. [CrossRef] [PubMed]
- 84. Nagel, R.; Traub, R.J.; Allcock, R.J.; Kwan, M.M.; Bielefeldt-Ohmann, H. Comparison of faecal microbiota in *Blastocystis*-positive and *Blastocystis*-negative irritable bowel syndrome patients. *Microbiome* **2016**, *4*, 47. [CrossRef] [PubMed]
- 85. Yildiz, S.; Dogan, I.; Dogruman-Al, F.; Nalbantoglu, U.; Ustek, D.; Sarzhanov, F.; Yildirim, S. Association of Enteric Protist *Blastocystis* spp. and Gut Microbiota with Hepatic Encephalopathy. *J. Gastrointest. Liver Dis.* **2016**, 25, 489–497. [CrossRef] [PubMed]
- 86. Gallo, A.; Passaro, G.; Gasbarrini, A.; Landolfi, R.; Montalto, M. Modulation of microbiota as treatment for intestinal inflammatory disorders: An uptodate. *World J. Gastroenterol.* **2016**, 22, 7186–7202. [CrossRef] [PubMed]
- 87. Rojas-Velázquez, L.; Morán, P.; Serrano-Vázquez, A.; Portillo-Bobadilla, T.; González, E.; Pérez-Juárez, H.; Hernández, E.; Partida-Rodríguez, O.; Nieves-Ramírez, M.; Padilla, A.; et al. The regulatory function of *Blastocystis* spp. on the immune inflammatory response in the gut microbiome. *Front. Cell. Infect. Microbiol.* **2022**, *12*, 967724. [CrossRef] [PubMed]
- 88. Betts, E.L.; Newton, J.M.; Thompson, G.S.; Sarzhanov, F.; Jinatham, V.; Kim, M.-J.; Popluechai, S.; Dogruman-Al, F.; Won, E.-J.; Gentekaki, E. Metabolic fluctuations in the human stool obtained from *Blastocystis* carriers and non-carriers. *Metabolites* **2021**, *11*, 883. [CrossRef]
- 89. Tan, K.S.W.; Mirza, H. *Blastocystis*—Host Interactions: Insights from In Vitro Model Systems. In *Blastocystis: Pathogen or Passenger?*An Evaluation of 101 Years of Research; Mehlhorn, H., Tan, K.S.W., Yoshikawa, H., Eds.; Springer: Berlin/Heidelberg, Germany, 2012; pp. 51–63. [CrossRef]

90. Partida-Rodriguez, O.; Serrano-Vazquez, A.; Nieves-Ramirez, M.E.; Moran, P.; Rojas, L.; Portillo, T.; Gonzalez, E.; Hernandez, E.; Finlay, B.B.; Ximenez, C. Human Intestinal Microbiota: Interaction between Parasites and the Host Immune Response. *Arch. Med. Res.* **2017**, *48*, 690–700. [CrossRef]

- 91. Reynolds, L.A.; Finlay, B.B.; Maizels, R.M. Cohabitation in the Intestine: Interactions among Helminth Parasites, Bacterial Microbiota, and Host Immunity. *J. Immunol.* **2015**, 195, 4059–4066. [CrossRef]
- 92. Huttenhower, C.; Gevers, D.; Knight, R.; Abubucker, S.; Badger, J.H.; Chinwalla, A.T.; Creasy, H.H.; Earl, A.M.; FitzGerald, M.G.; Fulton, R.S.; et al. Structure, function and diversity of the healthy human microbiome. *Nature* **2012**, 486, 207–214. [CrossRef]
- 93. Ahrodia, T.; Das, S.; Bakshi, S.; Das, B. Structure, functions, and diversity of the healthy human microbiome. *Prog. Mol. Biol. Transl.* **2022**, *191*, 53–82. [CrossRef]
- 94. Belzer, C.; Chia, L.W.; Aalvink, S.; Chamlagain, B.; Piironen, V.; Knol, J.; de Vos, W.M. Microbial metabolic networks at the mucus layer lead to diet-independent butyrate and vitamin B₁₂ production by intestinal symbionts. *Mbio* **2017**, *8*, e00770-17. [CrossRef] [PubMed]
- 95. Llinás-Caballero, K.; Caraballo, L. Helminths and bacterial microbiota: The interactions of two of humans' "old friends". *Int. J. Mol. Sci.* 2022, 23, 13358. [CrossRef] [PubMed]
- 96. Schonfeld, P.; Wojtczak, L. Short- and medium-chain fatty acids in energy metabolism: The cellular perspective. *J. Lipid Res.* **2016**, 57, 943–954. [CrossRef]
- 97. Woof, J.M.; Kerr, M.A. The function of immunoglobulin A in immunity. *J. Pathol. J. Pathol. Soc. Great Br. Irel.* **2006**, 208, 270–282. [CrossRef]
- 98. Puthia, M.K.; Vaithilingam, A.; Lu, J.; Tan, K.S. Degradation of human secretory immunoglobulin A by *Blastocystis*. *Parasitol*. *Res.* **2005**, 97, 386–389. [CrossRef]
- 99. Puthia, M.K.; Lu, J.; Tan, K.S. *Blastocystis* ratti contains cysteine proteases that mediate interleukin-8 response from human intestinal epithelial cells in an NF-κB-dependent manner. *Eukaryot*. *Cell* **2008**, 7, 435–443. [CrossRef]
- 100. Long, H.; Handschack, A.; König, W.; Ambrosch, A. *Blastocystis* hominis modulates immune responses and cytokine release in colonic epithelial cells. *Parasitol. Res.* **2001**, *87*, 1029–1030. [CrossRef]
- 101. Lim, M.X.; Png, C.W.; Tay, C.Y.; Teo, J.D.; Jiao, H.; Lehming, N.; Tan, K.S.; Zhang, Y. Differential regulation of proinflammatory cytokine expression by mitogen-activated protein kinases in macrophages in response to intestinal parasite infection. *Infect. Immun.* 2014, 82, 4789–4801. [CrossRef]
- 102. Baumgart, D.C.; Carding, S.R. Inflammatory bowel disease: Cause and immunobiology. Lancet 2007, 369, 1627–1640. [CrossRef]
- 103. Chen, J.; Zhang, Y.; Deng, Z. Imbalanced shift of cytokine expression between T helper 1 and T helper 2 (Th1/Th2) in intestinal mucosa of patients with post-infectious irritable bowel syndrome. *BMC Gastroenterol.* **2012**, *12*, 91. [CrossRef]
- 104. Lukeš, J.; Stensvold, C.R.; Jirků-Pomajbíková, K.; Wegener Parfrey, L. Are human intestinal eukaryotes beneficial or commensals? *PLoS Pathog.* **2015**, *11*, e1005039. [CrossRef]
- 105. Shaheen, W.A.; Quraishi, M.N.; Iqbal, T.H. Gut microbiome and autoimmune disorders. *Clin. Exp. Immunol.* **2022**, 209, 161–174. [CrossRef]
- 106. De Luca, F.; Shoenfeld, Y. The microbiome in autoimmune diseases. Clin. Exp. Immunol. 2019, 195, 74–85. [CrossRef]
- 107. Mahmoud, A.M.; Ismail, K.A.; Khalifa, O.M.; Abdel-wahab, M.M.; Hagag, H.M.; Mahmoud, M.K. Molecular Identification of *Blastocystis* hominis Isolates in Patients with Autoimmune Diseases. *Appl. Microbiol.* **2023**, *3*, 417–428. [CrossRef]
- 108. Keshawy, M.M.; Alabbassy, M.M. Systemic Lupus Erythematosus and Irritable Bowel Syndrome: Is *Blastocystis* Hominis the Missing Piece of the Puzzle. *Suez Canal Univ. Med. J.* **2018**, 21, 88–98. [CrossRef]
- 109. Jimenez-Balderas, F.; Camargo-Coronel, A.; Gargia-Jaimes, J.; Zonana-Nacach, A.; Alcantara-Anguianoi, I.; Carrillo-Becerril, L.; Tapia-Romero, R.; Gonzalez, T.; Villalobos-Gomez, F.; Martinez-Hernandez, S. A study on parasites in Mexican rheumatic disease patients. *J. Egypt. Soc. Parasitol.* **2012**, *42*, 271–280. [CrossRef] [PubMed]
- 110. Alamlih, L.; Abufaied, M.; Al-Allaf, A.-W. Classical reactive arthritis (ReA) due to *Blastocystis* infection: A very unusual trigger. *Rheumatology* **2019**, *58*, kez108.019. [CrossRef]
- 111. Nieto-Clavijo, C.; Morales, L.; Marquez-Ortíz, R.A.; Romero-Sánchez, C.; Ramos-Casallas, A.; Escobar-Perez, J.; Bautista-Molano, W.; Bello-Gualtero, J.M.; Chaparro-Olaya, J. Differential gut microbiome in spondyloarthritis patients associated to *Blastocystis* colonization. *Sci. Rep.* 2023, *13*, 13480. [CrossRef] [PubMed]
- 112. Bakland, G.; Nossent, H.C. Epidemiology of spondyloarthritis: A review. Curr. Rheumatol. Rep. 2013, 15, 351. [CrossRef] [PubMed]
- 113. Stoll, M.L.; Weiss, P.F.; Weiss, J.E.; Nigrovic, P.A.; Edelheit, B.S.; Bridges, S.L., Jr.; Danila, M.I.; Spencer, C.H.; Punaro, M.G.; Schikler, K.; et al. Age and fecal microbial strain-specific differences in patients with spondyloarthritis. *Arthritis Res. Ther.* **2018**, 20, 14. [CrossRef] [PubMed]
- 114. Wang, L.; Wang, Y.; Zhang, P.; Song, C.; Pan, F.; Li, G.; Peng, L.; Yang, Y.; Wei, Z.; Huang, F. Gut microbiota changes in patients with spondyloarthritis: A systematic review. *Semin. Arthritis Rheum.* **2022**, *52*, 151925. [CrossRef] [PubMed]
- 115. Tito, R.Y.; Cypers, H.; Joossens, M.; Varkas, G.; Van Praet, L.; Glorieus, E.; Van den Bosch, F.; De Vos, M.; Raes, J.; Elewaut, D. Brief Report: Dialister as a Microbial Marker of Disease Activity in Spondyloarthritis. *Arthritis Rheumatol.* **2017**, *69*, 114–121. [CrossRef] [PubMed]
- 116. Breban, M.; Tap, J.; Leboime, A.; Said-Nahal, R.; Langella, P.; Chiocchia, G.; Furet, J.P.; Sokol, H. Faecal microbiota study reveals specific dysbiosis in spondyloarthritis. *Ann. Rheum. Dis.* **2017**, *76*, 1614–1622. [CrossRef] [PubMed]

117. Wen, C.; Zheng, Z.; Shao, T.; Liu, L.; Xie, Z.; Le Chatelier, E.; He, Z.; Zhong, W.; Fan, Y.; Zhang, L.; et al. Quantitative metagenomics reveals unique gut microbiome biomarkers in ankylosing spondylitis. *Genome Biol.* **2017**, *18*, 142. [CrossRef]

- 118. Shaikh, S.D.; Sun, N.; Canakis, A.; Park, W.Y.; Weber, H.C. Irritable Bowel Syndrome and the Gut Microbiome: A Comprehensive Review. *J. Clin. Med.* **2023**, *12*, 2558. [CrossRef] [PubMed]
- 119. Chung, C.S.; Chang, P.F.; Liao, C.H.; Lee, T.H.; Chen, Y.; Lee, Y.C.; Wu, M.S.; Wang, H.P.; Ni, Y.H. Differences of microbiota in small bowel and faeces between irritable bowel syndrome patients and healthy subjects. *Scand. J. Gastroenterol.* **2016**, *51*, 410–419. [CrossRef] [PubMed]
- 120. Carroll, I.M.; Ringel-Kulka, T.; Siddle, J.P.; Ringel, Y. Alterations in composition and diversity of the intestinal microbiota in patients with diarrhea-predominant irritable bowel syndrome. *Neurogastroenterol. Motil.* **2012**, 24, 521-e248. [CrossRef]
- 121. Pittayanon, R.; Lau, J.T.; Yuan, Y.; Leontiadis, G.I.; Tse, F.; Surette, M.; Moayyedi, P. Gut Microbiota in Patients with Irritable Bowel Syndrome—A Systematic Review. *Gastroenterology* **2019**, *157*, 97–108. [CrossRef]
- 122. Liu, H.N.; Wu, H.; Chen, Y.Z.; Chen, Y.J.; Shen, X.Z.; Liu, T.T. Altered molecular signature of intestinal microbiota in irritable bowel syndrome patients compared with healthy controls: A systematic review and meta-analysis. *Dig. Liver Dis.* **2017**, *49*, 331–337. [CrossRef]
- 123. Longstreth, G.F.; Thompson, W.G.; Chey, W.D.; Houghton, L.A.; Mearin, F.; Spiller, R.C. Functional bowel disorders. *Gastroenterology* **2006**, *130*, 1480–1491. [CrossRef]
- 124. Costea, P.I.; Hildebrand, F.; Arumugam, M.; Backhed, F.; Blaser, M.J.; Bushman, F.D.; de Vos, W.M.; Ehrlich, S.D.; Fraser, C.M.; Hattori, M.; et al. Enterotypes in the landscape of gut microbial community composition. *Nat. Microbiol.* **2018**, *3*, 8–16. [CrossRef]
- 125. Ou, J.; Carbonero, F.; Zoetendal, E.G.; DeLany, J.P.; Wang, M.; Newton, K.; Gaskins, H.R.; O'Keefe, S.J. Diet, microbiota, and microbial metabolites in colon cancer risk in rural Africans and African Americans. *Am. J. Clin. Nutr.* **2013**, *98*, 111–120. [CrossRef]
- 126. Sobhani, I.; Tap, J.; Roudot-Thoraval, F.; Roperch, J.P.; Letulle, S.; Langella, P.; Corthier, G.; Tran Van Nhieu, J.; Furet, J.P. Microbial dysbiosis in colorectal cancer (CRC) patients. *PLoS ONE* **2011**, *6*, e16393. [CrossRef]
- 127. De Palma, G.; Nadal, I.; Medina, M.; Donat, E.; Ribes-Koninckx, C.; Calabuig, M.; Sanz, Y. Intestinal dysbiosis and reduced immunoglobulin-coated bacteria associated with coeliac disease in children. *BMC Microbiol.* **2010**, *10*, 63. [CrossRef]
- 128. Liu, L.; Huh, J.R.; Shah, K. Microbiota and the gut-brain-axis: Implications for new therapeutic design in the CNS. *EBioMedicine* **2022**, 77, 103908. [CrossRef]
- 129. Osadchiy, V.; Martin, C.R.; Mayer, E.A. The Gut-Brain Axis and the Microbiome: Mechanisms and Clinical Implications. *Clin. Gastroenterol. Hepatol.* **2019**, 17, 322–332. [CrossRef]
- 130. Fulling, C.; Dinan, T.G.; Cryan, J.F. Gut Microbe to Brain Signaling: What Happens in Vagus. *Neuron* **2019**, *101*, 998–1002. [CrossRef] [PubMed]
- 131. Blecharz-Klin, K.; Swierczynska, M.; Piechal, A.; Wawer, A.; Joniec-Maciejak, I.; Pyrzanowska, J.; Wojnar, E.; Zawistowska-Deniziak, A.; Sulima-Celinska, A.; Mlocicki, D.; et al. Infection with intestinal helminth (*Hymenolepis diminuta*) impacts exploratory behavior and cognitive processes in rats by changing the central level of neurotransmitters. *PLoS Pathog.* **2022**, *18*, e1010330. [CrossRef] [PubMed]
- 132. Carabotti, M.; Scirocco, A.; Maselli, M.A.; Severi, C. The gut-brain axis: Interactions between enteric microbiota, central and enteric nervous systems. *Ann. Gastroenterol.* **2015**, *28*, 203–209. [PubMed]
- 133. McKenney, E.A.; Williamson, L.; Yoder, A.D.; Rawls, J.F.; Bilbo, S.D.; Parker, W. Alteration of the rat cecal microbiome during colonization with the helminth Hymenolepis diminuta. *Gut Microbes* 2015, *6*, 182–193. [CrossRef] [PubMed]
- 134. Wu, J.; Zhu, Y.; Zhou, L.; Lu, Y.; Feng, T.; Dai, M.; Liu, J.; Xu, W.; Cheng, W.; Sun, F.; et al. Parasite-Derived Excretory-Secretory Products Alleviate Gut Microbiota Dysbiosis and Improve Cognitive Impairment Induced by a High-Fat Diet. *Front. Immunol.* **2021**, *12*, 710513. [CrossRef] [PubMed]
- 135. Defaye, M.; Nourrisson, C.; Baudu, E.; Lashermes, A.; Meynier, M.; Meleine, M.; Wawrzyniak, I.; Bonnin, V.; Barbier, J.; Chassaing, B.; et al. Fecal dysbiosis associated with colonic hypersensitivity and behavioral alterations in chronically *Blastocystis*-infected rats. *Sci. Rep.* **2020**, *10*, 9146. [CrossRef] [PubMed]
- 136. Mayneris-Perxachs, J.; Arnoriaga-Rodriguez, M.; Garre-Olmo, J.; Puig, J.; Ramos, R.; Trelis, M.; Burokas, A.; Coll, C.; Zapata-Tona, C.; Pedraza, S.; et al. Presence of *Blastocystis* in gut microbiota is associated with cognitive traits and decreased executive function. *ISME J.* 2022, *16*, 2181–2197. [CrossRef]
- 137. Borre, Y.E.; Moloney, R.D.; Clarke, G.; Dinan, T.G.; Cryan, J.F. The impact of microbiota on brain and behavior: Mechanisms & therapeutic potential. In *Microbial Endocrinology: The Microbiota-Gut-Brain Axis in Health and Disease*; Springer: New York, NY, USA, 2014; pp. 373–403.
- 138. Leonardi, S.S.; Tan, K.S.-W. Blastocystis: View from atop the gut-brain iceberg. Trends Parasitol. 2023, 40, 1–4. [CrossRef]
- 139. Rieder, R.; Wisniewski, P.J.; Alderman, B.L.; Campbell, S.C. Microbes and mental health: A review. *Brain Behav. Immun.* **2017**, 66, 9–17. [CrossRef]
- 140. Khine, W.W.T.; Voong, M.L.; Ng, T.K.S.; Feng, L.; Rane, G.A.; Kumar, A.P.; Kua, E.H.; Mahendran, R.; Mahendran, R.; Lee, Y.K. Mental awareness improved mild cognitive impairment and modulated gut microbiome. *Aging* **2020**, *12*, 24371–24393. [CrossRef]
- 141. Stensvold, C.R.; van der Giezen, M. Associations between Gut Microbiota and Common Luminal Intestinal Parasites. *Trends Parasitol.* **2018**, *34*, 369–377. [CrossRef]
- 142. Correia, A.S.; Vale, N. Tryptophan metabolism in depression: A narrative review with a focus on serotonin and kynurenine pathways. *Int. J. Mol. Sci.* **2022**, *23*, 8493. [CrossRef]

143. Nozawa, K.; Kawabata-Shoda, E.; Doihara, H.; Kojima, R.; Okada, H.; Mochizuki, S.; Sano, Y.; Inamura, K.; Matsushime, H.; Koizumi, T. TRPA1 regulates gastrointestinal motility through serotonin release from enterochromaffin cells. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 3408–3413. [CrossRef]

- 144. Travers, M.-A.; Florent, I.; Kohl, L.; Grellier, P. Probiotics for the control of parasites: An overview. *J. Parasitol. Res.* **2011**, 2011, 610769. [CrossRef]
- 145. Hill, C.; Guarner, F.; Reid, G.; Gibson, G.R.; Merenstein, D.J.; Pot, B.; Morelli, L.; Canani, R.B.; Flint, H.J.; Salminen, S. Expert consensus document: The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat. Rev. Gastroenterol. Hepatol.* 2014, 11, 506–514. [CrossRef]
- 146. Vitetta, L.; Saltzman, E.T.; Nikov, T.; Ibrahim, I.; Hall, S. Modulating the gut micro-environment in the treatment of intestinal parasites. *J. Clin. Med.* **2016**, *5*, 102. [CrossRef]
- 147. Vallianou, N.; Stratigou, T.; Christodoulatos, G.S.; Tsigalou, C.; Dalamaga, M. Probiotics, Prebiotics, Synbiotics, Postbiotics, and Obesity: Current Evidence, Controversies, and Perspectives. *Curr. Obes. Rep.* **2020**, *9*, 179–192. [CrossRef] [PubMed]
- 148. Sarid, L.; Zanditenas, E.; Ye, J.; Trebicz-Geffen, M.; Ankri, S. Insights into the mechanisms of Lactobacillus acidophilus activity against *Entamoeba histolytica* by using thiol redox proteomics. *Antioxidants* **2022**, *11*, 814. [CrossRef] [PubMed]
- 149. Hardy, H.; Harris, J.; Lyon, E.; Beal, J.; Foey, A.D. Probiotics, prebiotics and immunomodulation of gut mucosal defences: Homeostasis and immunopathology. *Nutrients* **2013**, *5*, 1869–1912. [CrossRef] [PubMed]
- 150. Markowiak, P.; Śliżewska, K. Effects of probiotics, prebiotics, and synbiotics on human health. *Nutrients* **2017**, *9*, 1021. [CrossRef] [PubMed]
- 151. Silva, D.R.; Sardi, J.d.C.O.; de Souza Pitangui, N.; Roque, S.M.; da Silva, A.C.B.; Rosalen, P.L. Probiotics as an alternative antimicrobial therapy: Current reality and future directions. *J. Funct. Foods* **2020**, *73*, 104080. [CrossRef]
- 152. Berrilli, F.; Di Cave, D.; Cavallero, S.; D'Amelio, S. Interactions between parasites and microbial communities in the human gut. *Front. Cell. Infect. Microbiol.* **2012**, *2*, 141. [CrossRef]
- 153. Pryshliak, O.Y.; Protsyk, A.L.; Semaniv, M.V.; Boichuk, O.P.; Gerych, P.R. Effect of probiotics on the intestinal microbiota of patients with giardiasis and ascariasis. *J. Med. Life* **2022**, *15*, 1278. [CrossRef]
- 154. Al-Megrin, W.A.; Mohamed, S.H.; Saleh, M.M.; Yehia, H.M. Preventive role of probiotic bacteria against gastrointestinal diseases in mice caused by Giardia lamblia. *Biosci. Rep.* **2021**, *41*, BSR20204114. [CrossRef]
- 155. Del Coco, V.F.; Sparo, M.D.; Sidoti, A.; Santín, M.; Basualdo, J.A.; Córdoba, M.A. Effects of Enterococcus faecalis CECT 7121 on *Cryptosporidium parvum* infection in mice. *Parasitol. Res.* **2016**, *115*, 3239–3244. [CrossRef] [PubMed]
- 156. Shukla, G.; Sharma, A.; Bhatia, R.; Sharma, M. Prophylactic potential of synbiotic (*Lactobacillus casei* and Inulin) in malnourished murine giardiasis: An immunological and ultrastructural study. *Probiotics Antimicrob. Proteins* **2019**, *11*, 165–174. [CrossRef] [PubMed]
- 157. Saracino, M.P.; Vila, C.C.; Baldi, P.C.; Gonzalez Maglio, D.H. Searching for the one (s): Using probiotics as anthelmintic treatments. *Front. Pharmacol.* **2021**, *12*, 714198. [CrossRef]
- 158. Rooney, J.; Cantacessi, C.; Sotillo, J.; Cortés, A. Gastrointestinal worms and bacteria: From association to intervention. *Parasite Immunol.* 2023, 45, e12955. [CrossRef] [PubMed]
- 159. Andersen, L.O.; Stensvold, C.R. *Blastocystis* in Health and Disease: Are We Moving from a Clinical to a Public Health Perspective? *J. Clin. Microbiol.* **2016**, *54*, 524–528. [CrossRef] [PubMed]
- 160. Mirza, H.; Wu, Z.; Kidwai, F.; Tan, K.S.W. A Metronidazole-Resistant Isolate of *Blastocystis* spp. Is Susceptible to Nitric Oxide and Downregulates Intestinal Epithelial Inducible Nitric Oxide Synthase by a Novel Parasite Survival Mechanism. *Infect. Immun.* **2011**, *79*, 5019–5026. [CrossRef] [PubMed]
- 161. Dunn, L.A.; Tan, K.S.; Vanelle, P.; Juspin, T.; Crozet, M.D.; Terme, T.; Upcroft, P.; Upcroft, J.A. Development of metronidazole-resistant lines of *Blastocystis* sp. *Parasitol. Res.* **2012**, *111*, 441–450. [CrossRef] [PubMed]
- 162. Sekar, U.; Shanthi, M. Blastocystis: Consensus of treatment and controversies. Trop. Parasitol. 2013, 3, 35–39. [CrossRef]
- 163. Roberts, T.; Stark, D.; Harkness, J.; Ellis, J. Update on the pathogenic potential and treatment options for *Blastocystis* sp. *Gut Pathog.* **2014**, *6*, 17. [CrossRef]
- 164. Batista, L.; Perez Jove, J.; Rosinach, M.; Gonzalo, V.; Sainz, E.; Loras, C.; Forne, M.; Esteve, M.; Fernandez-Banares, F. Low efficacy of metronidazole in the eradication of *Blastocystis* hominis in symptomatic patients: Case series and systematic literature review. *Gastroenterol. Hepatol.* **2017**, *40*, 381–387. [CrossRef]
- 165. Dinleyici, E.C.; Eren, M.; Dogan, N.; Reyhanioglu, S.; Yargic, Z.A.; Vandenplas, Y. Clinical efficacy of Saccharomyces boulardii or metronidazole in symptomatic children with *Blastocystis* hominis infection. *Parasitol. Res.* **2011**, *108*, 541–545. [CrossRef] [PubMed]
- 166. Angelici, M.C.; Nardis, C.; Scarpelli, R.; Ade, P. *Blastocystis* hominis transmission by non-potable water: A case report in Italy. *New Microbiol.* **2018**, *41*, 173–177. [PubMed]

167. Méabed, E.M.; Abdelhafez, D.N.; Abdelaliem, Y.F. Saccharomyces boulardii inhibits the expression of pro-inflammatory cytokines and inducible nitric oxide synthase genes in the colonic mucosa of rats experimentally-infected with *Blastocystis* subtype-3 cysts. *Parasitology* **2019**, *146*, 1532–1540. [CrossRef]

168. Lepczynska, M.; Dzika, E. The influence of probiotic bacteria and human gut microorganisms causing opportunistic infections on *Blastocystis* ST3. *Gut Pathog.* **2019**, *11*, 6. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.